Package 'bbknnR'

July 22, 2025

```
Title Perform Batch Balanced KNN in R
Version 2.0.1
Date 2025-06-05
Description A fast and intuitive batch effect removal tool for single-cell data. BBKNN is origi-
     nally used in the 'scanpy' python package, and now can be used with 'Seurat' seamlessly.
License MIT + file LICENSE
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```

panc8_small

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panc8_small

A small example version of the pancreas scRNA-seq dataset

Description

A subsetted version of the pancreas scRNA-seq dataset to test BBKNN

Usage

panc8_small

Format

A Seurat object with the following slots filled

assays Currently only contains one assay ("RNA" - scRNA-seq expression data)

counts - Raw expression data

- data Normalized expression data
- scale.data Scaled expression data
- var.features names of the current features selected as variable
- meta.features Assay level metadata such as mean and variance

meta.data Cell level metadata active.assay Current default assay

active.ident Current default idents

graphs Empty

reductions Dimensional reductions: currently PCA

version Seurat version used to create the object

commands Command history

Source

SeuratData https://github.com/satijalab/seurat-data

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RidgeRegression

Perform ridge regression on scaled expression data

Description

Perform ridge regression on scaled expression data, accepting both technical and biological categorical variables. The effect of the technical variables is removed while the effect of the biological variables is retained. This is a preprocessing step that can aid BBKNN integration.

Usage

```
RidgeRegression(object, ...)
## Default S3 method:
RidgeRegression(
  object,
  latent_data,
  batch_key,
  confounder_key,
  lambda = 1,
  seed = 42,
  verbose = TRUE,
)
## S3 method for class 'Seurat'
RidgeRegression(
 object,
 batch_key,
  confounder_key,
  assay = NULL,
  features = NULL,
  lambda = 1,
  run_pca = TRUE,
  npcs = 50,
  reduction.name = "pca",
  reduction.key = "PC_",
  replace = FALSE,
  seed = 42,
  verbose = TRUE,
)
```

Arguments

object An object

. . . Arguments passed to other methods

latent_data	Extra data to regress out, should be cells x latent data
batch_key	Variables to regress out as technical effects. Must be included in column names of latent_data
confounder_key	Variables to to retain as biological effects. Must be included in column names of latent_data
lambda	A user supplied lambda sequence. pass to glmnet
seed	Set a random seed. By default, sets the seed to 42. Setting NULL will not set a seed.
verbose	Whether or not to print output to the console
assay	Name of Assay ridge regression is being run on
features	Features to compute ridge regression on. If features=NULL, ridge regression will be run using the variable features for the Assay.
run_pca	Whether or not to run pca with regressed expression data (TRUE by default)
npcs	Total Number of PCs to compute and store (50 by default)
reduction.name	Dimensional reduction name (pca by default)
reduction.key	Dimensional reduction key, specifies the string before the number for the dimension names (PC by default)
replace	Whether or not to replace original scale.data with regressed expression data

Value

Returns a Seurat object.

References

Park, Jong-Eun, et al. "A cell atlas of human thymic development defines T cell repertoire formation." Science 367.6480 (2020): eaay3224.

Examples

```
data("panc8_small")
panc8_small <- RidgeRegression(panc8_small, "tech", c("nCount_RNA"))</pre>
```

(TRUE by default)

RunBBKNN	Perform batch balanced KNN

Description

Batch balanced KNN, altering the KNN procedure to identify each cell's top neighbours in each batch separately instead of the entire cell pool with no accounting for batch. The nearest neighbours for each batch are then merged to create a final list of neighbours for the cell. Aligns batches in a quick and lightweight manner.

Usage

```
RunBBKNN(object, ...)
## S3 method for class 'matrix'
RunBBKNN(
  object,
  batch_list,
  neighbors_within_batch = 3,
  n_pcs = 50,
 method = c("annoy", "nndescent"),
 metric = "euclidean",
  n_{\text{trees}} = 10L
  k_build_nndescent = 30,
  trim = NULL,
  set_op_mix_ratio = 1,
  local_connectivity = 1,
  seed = 42,
  verbose = TRUE,
)
## S3 method for class 'Seurat'
RunBBKNN(
  object,
  batch_key,
  assay = NULL,
  reduction = "pca",
  n_pcs = 50L,
  graph_name = "bbknn",
  set_op_mix_ratio = 1,
  local_connectivity = 1,
  run_TSNE = TRUE,
  TSNE_name = "tsne"
  TSNE_key = "tSNE_",
  run_UMAP = TRUE,
 UMAP_name = "umap",
 UMAP_key = "UMAP_",
  return.umap.model = FALSE,
 min_dist = 0.3,
  spread = 1,
  seed = 42,
  verbose = TRUE,
)
```

Arguments

object An object

... Arguments passed to other methods

batch_list A character vector with the same length as nrow(pca)

neighbors_within_batch

How many top neighbours to report for each batch; total number of neighbours in the initial k-nearest-neighbours computation will be this number times the number of batches. This then serves as the basis for the construction of a symmetrical matrix of connectivities.

n_pcs

Number of dimensions to use. Default is 50.

method

Method to find k nearest neighbors (kNNs). One of "annoy" and "nndescent".

metric

Metric to calculate the distances. The options depend on the choice of kNN method. The following metrics are supported in both annoy and nndescent:

- 'euclidean' (the default)
- · 'manhattan'
- · 'hamming'

The following metrics are only supported in nndescent:

- · 'sqeuclidean'
- · 'chebyshev'
- · 'canberra'
- · 'braycurtis'
- · 'cosine'
- · 'correlation'
- · 'jaccard'
- · 'dice'
- · 'matching'
- 'russellrao'
- · 'kulsinski'
- · 'rogerstanimoto'
- · 'sokalmichener'
- 'sokalsneath'
- · 'tsss'
- 'yule'
- · 'hellinger'

n_trees

The number of trees to use in the random projection forest. More trees give higher precision when querying, at the cost of increased run time and resource intensity.

k_build_nndescent

Used with nndescent neighbour identification. The number of neighbours to include when building the approximate nearest neighbors index and neighbor graph. More neighbours give higher precision when querying, at the cost of increased run time and resource intensity.

trim

Trim the neighbours of each cell to these many top connectivities. May help with population independence and improve the tidiness of clustering. The lower the value the more independent the individual populations, at the cost of more conserved batch effect. Default is 10 times neighbors_within_batch times the number of batches. Set to 0 to skip.

set_op_mix_ratio

Pass to 'set_op_mix_ratio' parameter for umap

local_connectivity

Pass to 'local_connectivity' parameter for umap

seed Set a random seed. By default, sets the seed to 42. Setting NULL will not set a

seed.

verbose Whether or not to print output to the console

batch_key Column name in meta.data discriminating between your batches.

assay Used to construct Graph.

reduction Which dimensional reduction to use for the BBKNN input. Default is PCA

graph_name Name of the generated BBKNN graph. Default is "bbknn".
run_TSNE Whether or not to run t-SNE based on BBKNN results.

TSNE_name Name to store t-SNE dimensional reduction.

TSNE_key Specifies the string before the number of the t-SNE dimension names. tSNE by

default.

run_UMAP Whether or not to run UMAP based on BBKNN results.

UMAP_name Name to store UMAP dimensional reduction.

UMAP_key Specifies the string before the number of the UMAP dimension names. tSNE by

default.

return.umap.model

Whether UMAP will return the uwot model.

min_dist Pass to 'min_dist' parameter for umap spread Pass to 'spread' parameter for umap

Value

Returns a Seurat object containing a new BBKNN Graph and Neighbor data. If run t-SNE or UMAP, will also return corresponded reduction objects.

References

Polański, Krzysztof, et al. "BBKNN: fast batch alignment of single cell transcriptomes." Bioinformatics 36.3 (2020): 964-965.

Examples

```
data("panc8_small")
panc8_small <- RunBBKNN(panc8_small, "tech")</pre>
```

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