# Package 'detectRUNS'

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**Description** Detection of runs of homozygosity and of heterozygosity in diploid genomes using two methods: sliding windows (Purcell et al (2007) <doi:10.1086/519795>) and consecutive runs (Marras et al (2015) <doi:10.1111/age.12259>).

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consecutiveRUNS.run

Main function to detect genomic RUNS (ROHom/ROHet) using the consecutive method

## Description

This is the main detectRUNS function to scan the genome for runs (of homozygosity or heterozygosity) using the consecutive method (Marras et al. 2015, Animal Genetics 46(2):110-121). All parameters to detect runs (e.g. minimum n. of SNP, max n. of missing genotypes, max n. of opposite genotypes etc.) are specified here. Input data are in the ped/map Plink format (https://www.cog-genomics.org/plink/1.9/input#ped)

#### Usage

```
consecutiveRUNS.run(genotypeFile, mapFile, ROHet = FALSE,
maxOppRun = 0, maxMissRun = 0, minSNP = 15, minLengthBps = 1000,
maxGap = 10<sup>6</sup>)
```

## Arguments

genotypeFile	genotype (.ped) file path
mapFile	map file (.map) file path
ROHet	should we look for ROHet or ROHom? (default = FALSE)
maxOppRun	max n. of opposite genotype SNPs in the run (default = $0$ )
maxMissRun	max n. of missing SNPs in the run (default = $0$ )
minSNP	minimum n. of SNP in a RUN (default = 15)
minLengthBps	minimum length of run in bps (defaults to 1000 bps = 1 kbps)
maxGap	max distance between consecutive SNP in a window to be still considered a potential run (defaults to 10 <sup>6</sup> )

#### Details

This function scans the genome (diploid) for runs using the consecutive method. This is a wrapper function for many component functions that handle the input data (ped/map files), performs internal conversions, accepts parameters specifications, selects the statistical method to detect runs (sliding windows, consecutive loci) and whether runs of homozygosity (RoHom) or of heterozygosity (RoHet) are looked for.

In the ped file, the groups samples belong to can be specified (first column). This is important if comparisons between human ethnic groups or between animal breeds or plant varieties or biological populations are to be performed. Also, if cases and controls are to be compared, this is the place where this information needs to be specified.

This function returns a data frame with all runs detected in the dataset. This data frame can then be written out to a csv file. The data frame is, in turn, the input for other functions of the detectRUNS package that create plots and produce statistics of the results (see plot and statistic functions in this manual, and/or refer to the vignette of detectRUNS).

#### Value

A dataframe with RUNs of Homozygosity or Heterozygosity in the analysed dataset. The returned dataframe contains the following seven columns: "group", "id", "chrom", "nSNP", "from", "to", "lengthBps" (group: population, breed, case/control etc.; id: individual identifier; chrom: chromosome on which the run is located; nSNP: number of SNPs in the run; from: starting position of the run, in bps; to: end position of the run, in bps; lengthBps: size of the run)

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")</pre>
```

```
# calculating runs with consecutive run approach
## Not run:
# skipping runs calculation
runs <- consecutiveRUNS.run(genotypeFile, mapFile, minSNP = 15, ROHet = FALSE,
maxOppRun = 0, maxMissRun = 0, maxGap=10^6,
minLengthBps = 100000
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.consecutive.csv", package="detectRUNS")
colClasses <- c(rep("character", 3), rep("numeric", 4) )
runs <- read.csv2(runsFile, header = TRUE, stringsAsFactors = FALSE,
colClasses = colClasses)</pre>
```

consecutiveRunsCpp	Function to detect consecutive runs in a vector (individual's geno-
	types)

## Description

This is a core function. It implements the consecutive method for detection of runs in diploid genomes (see Marras et al. 2015)

## Usage

```
consecutiveRunsCpp(indGeno, individual, mapFile, ROHet = TRUE,
minSNP = 3L, maxOppositeGenotype = 1L, maxMiss = 1L,
minLengthBps = 1000L, maxGap = 1000000L)
```

#### Arguments

indGeno	vector of 0/1/NAs of individual genotypes (0: homozygote; 1: heterozygote)	
individual	list of group (breed, population, case/control etc.) and ID of individual sample	
mapFile	Plink map file (for SNP position)	
ROHet	shall we detect ROHet or ROHom?	
minSNP maxOppositeGeno	minimum number of SNP in a run notype	
	max n. of homozygous/heterozygous SNP	
maxMiss	max. n. of missing SNP	
minLengthBps	min length of a run in bps	
maxGap	max distance between consecutive SNP in a window to be still considered a potential run	

```
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```

#### createRUNdf

#### Details

The consecutive method detect runs by consecutively scanning SNP loci along the genome. No sliding windows are used. Checks on minimum n. of SNP, max n. of opposite and missing geno-types, max gap between adjacent loci and minimum length of the run are implemented (as in the sliding window method). Both runs of homozygosity (RoHom) and of heterozygosity (RoHet) can be search for (option ROHet: TRUE/FALSE)

## Value

A data frame of runs per individual sample

createRUNdf	Function to create a dataframe of RUNS per individual animal Re-
	quires a map file (other filename to read or R object) Parameters on
	maximum number of missing and opposite genotypes in the run (not
	the window) are implemented here

## Description

Function to create a dataframe of RUNS per individual animal Requires a map file (other filename to read or R object) Parameters on maximum number of missing and opposite genotypes in the run (not the window) are implemented here

#### Usage

```
createRUNdf(snpRun, mapFile, minSNP = 3, minLengthBps = 1000,
minDensity = 1/10, oppositeAndMissingSNP, maxOppRun = NULL,
maxMissRun = NULL)
```

#### Arguments

snpRun	vector of TRUE/FALSE (is the SNP in a RUN?)
mapFile	Plink-like map file (data.frame)
minSNP	minimum n. of SNP to call a RUN
minLengthBps	minimum length of run in bps (defaults to 1000 bps = 1 kbps)
minDensity	minimum n. of SNP per kbps (defaults to $0.1 = 1$ SNP every 10 kbps)
oppositeAndMiss	ingSNP
	indexed array of missing and opposite genotypes (SNP order in the genome is the index)
maxOppRun	max n. of opposite genotype SNPs in the run (not in the window!)
maxMissRun	max n. of missing SNPs in the run (not in the window!)

#### Value

a data.frame with RUNS per animal

```
findOppositeAndMissing
```

Function to calculate oppositeAndMissingGenotypes array

## Description

This is an helper function, this will be called by another function

## Usage

```
findOppositeAndMissing(data, ROHet = TRUE)
```

## Arguments

data	vector of 0/1/2 genotypes
ROHet	TRUE in ROHet evaluation, FALSE for ROHom

## Value

character array; names will be index in which opposite and missing snps are found in data array

Froh_inbreeding	Function to calculated Froh genome-wide or chromosome-wide
-----------------	--

## Description

This function calculates the individual inbreeding coefficients based on runs of homozygosity (ROH), either per-chromosome (chromosome-wide) or based on the entire genome (genome-wide). See details of calculations below

## Usage

```
Froh_inbreeding(runs, mapFile, genome_wide = TRUE)
```

## Arguments

runs	R object (dataframe) with results on runs
mapFile	Plink map file (to retrieve SNP position)
genome_wide	vector of TRUE/FALSE (genome-wide or chromosome-wide; defaults to TRUE/genome-wide)

#### Details

Froh is calculated as:

$$F_{ROH} = \frac{\sum ROH_{length}}{Length_{genome}}$$

Depending on whether genome-wide or chromosome-wide calculations are required, the terms in the numerator and denominator will refer to the entire genome or will be restricted to specific chromosomes.

#### Value

A data frame with the inbreeding coefficients of each individual sample

### Examples

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
Froh_inbreeding(runs = runsDF, mapFile = mapFile)
Froh_inbreeding(runs = runsDF, mapFile = mapFile, genome_wide=FALSE)
```

Froh\_inbreedingClass Function to calculated Froh using a ROH-class

#### Description

This function calculates the individual inbreeding coefficients based on runs of homozygosity (ROH) using only ROH of specific size classes. The parameter class specify the size interval to split up calculations. For example, if class = 2 Froh based on ROH 0-2, 2-4, 4-8, 80-16, >16 Mbps long will be calculated.

#### Usage

```
Froh_inbreedingClass(runs, mapFile, Class = 2)
```

#### Arguments

runs	R object (dataframe) with ROH results
mapFile	Plink map file (for SNP position)
Class	base ROH-length interval (in Mbps) (default: 0-2, 2-4, 4-8, 8-16, >16)

## Value

A data frame with individual inbreeding coefficients based on ROH-length of specific size. The sum of ROH-length of specific size in each individual is reported alongside

#### Examples

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
```

```
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')</pre>
```

```
Froh_inbreedingClass(runs = runsDF, mapFile = mapFile, Class = 2)
```

genoConvertCpp Convert 0/1/2 genotypes to 0/1

#### Description

This is a utility function, that convert 0/1/2 genotypes (AA/AB/BB) into 0/1 (either homozy-gous/heterozygous)

## Usage

```
genoConvertCpp(genotype)
```

#### Arguments

genotype vector of 0/1/2 genotypes

## Value

converted vector of genotypes (0/1)

heteroZygotTest

## Description

This is a core function within the sliding-window workflow. Parameters on how to consider a window heterozygous are here (maxHom, maxMiss)

## Usage

```
heteroZygotTest(x, gaps, maxHom, maxMiss, maxGap, i, windowSize)
```

#### Arguments

Х	vector of 0/1 genotypes (from genoConvert())
gaps	vector of differences between consecutive positions (gaps) in bps
maxHom	max n. of homozygous SNP in a heterozygous window
maxMiss	max n. of missing in a window
maxGap	max distance between consecutive SNP in a window to be still considered a potential run
i	index along the genome (genome-vector for each individual)
windowSize	size of window (n. of SNP)

## Value

a list: i) TRUE/FALSE (whether a window is heterozygous or NOT); ii) indexes of "opposite and missing" genotype

heteroZygotTestCpp Function to check whether a window is (loosely) heterozygous or not

## Description

This is a core function. Parameters on how to consider a window heterozygous are here (maxHom, maxMiss)

#### Usage

```
heteroZygotTestCpp(x, gaps, maxHom, maxMiss, maxGap)
```

## Arguments

х	vector of 0/1 genotypes (from genoConvert())
gaps	vector of differences between consecutive positions (gaps) in bps
maxHom	max n. of homozygous SNP in a heterozygous window
maxMiss	max n. of missing in a window
maxGap	max distance between consecutive SNP in a window to be still considered a potential run

## Value

TRUE/FALSE (whether a window is heterozygous or NOT)

homoZygotTest	Function to check whether a window is (loosely) homozygous or not
---------------	---

## Description

This is a core function. Parameters on how to consider a window homozygous are here (maxHet, maxMiss)

## Usage

homoZygotTest(x, gaps, maxHet, maxMiss, maxGap, i, windowSize)

## Arguments

х	vector of 0/1 genotypes (from genoConvert())
gaps	vector of differences between consecutive positions (gaps) in bps
maxHet	max n. of heterozygous SNP in a homozygous window
maxMiss	max n. of missing in a window
maxGap	max distance between consecutive SNP in a window to be still considered a potential run
i	index along the genome (genome-vector for each individual)
windowSize	size of window (n. of SNP)

### Value

a list: i) TRUE/FALSE (whether a window is heterozygous or NOT); ii) indexes of "opposite and missing" genotype

homoZygotTestCpp Function to check whether a window is (loosely) homozygous or not

## Description

This is a core function. Parameters on how to consider a window homozygous are here (maxHet, maxMiss)

## Usage

homoZygotTestCpp(x, gaps, maxHet, maxMiss, maxGap)

## Arguments

Х	vector of 0/1 genotypes (from genoConvert())
gaps	vector of differences between consecutive positions (gaps) in bps
maxHet	max n. of heterozygous SNP in a homozygous window
maxMiss	max n. of missing in a window
maxGap	max distance between consecutive SNP in a window to be still considered a potential run

#### Value

TRUE/FALSE (whether a window is homozygous or NOT)

pedConvertCpp Convert ped genotypes to 0/1

## Description

This is a utility function, that convert ped genotypes (AA/AB/BB) into 0/1 (either homozygous/heterozygous)

## Usage

```
pedConvertCpp(genotype)
```

## Arguments

genotype vector of pair of genotypes (01, AA, AG)

## Value

converted vector of genotypes (0/1)

plot\_DistributionRuns Plot Distribution of runs

#### Description

This function the distribution of runs per group. The average run length per size-class, the average run length per chromosome (and group), the percent distribution of runs per size-class and group, and the proportion of runs per chromosome are plotted. With style="All" all three plots are produced.

#### Usage

```
plot_DistributionRuns(runs, mapFile, groupSplit = TRUE,
   style = c("MeanClass", "MeanChr", "RunsPCT", "RunsPCT_Chr", "All"),
   savePlots = FALSE, outputName = NULL, plotTitle = NULL,
   Class = 2)
```

#### Arguments

runs	R object (dataframe) with results on detected runs
mapFile	Plink map file (for SNP position)
groupSplit	plots split by group (defaults to TRUE)
style	type of plot: MeanClass, MeanChr, RunsPCT, RunsPCT_Chr, All (all plots)
savePlots	should plots be saved out to files or plotted in the graphical terminal (default)?
outputName	title prefix (the base name of graph, if savePlots is TRUE)#'
plotTitle	title in plot (default NULL)
Class	group of length (in Mbps) by class (default: 0-2, 2-4, 4-8, 8-16, >16)

#### Value

plot Distribution Runs

# loading pre-calculated data

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
```

```
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
plot_InbreedingChr(runs = runsDF, mapFile = mapFile, style='All')</pre>
```

plot\_InbreedingChr Plot Froh-based inbreeding coefficients by group

#### Description

The function plots the distribution of inbreeding/consanguinity coefficients per chromosome and/or group. Three types of plots can be produces: barplots, boxplots, violin plots. With style="All" all three plots are produced.

#### Usage

```
plot_InbreedingChr(runs, mapFile, groupSplit = TRUE,
    style = c("ChrBarPlot", "ChrBoxPlot", "FrohBoxPlot", "All"),
    outputName = NULL, plotTitle = NULL, savePlots = FALSE)
```

#### Arguments

runs	R object (dataframe) with results on detected runs
mapFile	Plink map file (for SNP position)
groupSplit	plots split by group (defaults to TRUE)
style	type of plot: ChrBarPlot, ChrBoxPlot, FrohBoxPlot, All (all plots)
outputName	title prefix (the base name of graph, if savePlots is TRUE)
plot⊺itle	title in plot (default NULL)
savePlots	should plots be saved out to files or plotted in the graphical terminal (default)?

#### Value

plots of the distribution of inbreeding by chromosome and group

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)</pre>
```

```
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
plot_InbreedingChr(runs = runsDF, mapFile = mapFile, style='All')</pre>
```

plot\_manhattanRuns Plot the proportion of times SNPs are inside runs - MANHATTAN PLOT

#### Description

Function to plot the proportion of times/percentage each SNP in inside a run (population-specific signals) against SNP position in all chromosomes together Proportions on the y-axis, bps on the x-axis for all analysed chromosomes This is similar to the familiar GWAS Manhattan plot

## Usage

```
plot_manhattanRuns(runs, genotypeFile, mapFile, savePlots = FALSE,
    outputName = NULL, plotTitle = NULL)
```

#### Arguments

runs	a data.frame with runs per individual (group, id, chrom, nSNP, start, end, length)
genotypeFile	genotype (.ped) file path
mapFile	map file (.map) file path
savePlots	should plots be saved out in files (default) or plotted in the graphical terminal?
outputName	title prefix (the base name of graph, if savePlots is TRUE)
plotTitle	title in plot (default)

## Value

Manhattan plots of proportion of times SNPs are inside runs, per population (pdf files)

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)</pre>
```

#### plot\_PatternRuns

```
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
# plot runs per animal (interactive)
plot_manhattanRuns(runs = runsDF, genotypeFile = genotypeFile, mapFile = mapFile,
savePlots = FALSE, plotTitle = "ROHom")</pre>
```

plot\_PatternRuns

Plot sum of run-lengths (or average run-lengths) against the number of runs per individual

## Description

Function to plot the sum of run lengths (or the average run length) per individual against the average number of runs per individual. Points can be differentially coloured by group/population. This plot can be useful to identify patterns in the distribution of runs in different groups (e.g. few long runs vs many short runs)

#### Usage

```
plot_PatternRuns(runs, mapFile, method = c("sum", "mean"),
    outputName = NULL, savePlots = FALSE, plotTitle = NULL)
```

#### Arguments

runs	a data.frame with runs per individual (group, id, chrom, nSNP, start, end, length)
mapFile	map file (.map) file path
method	"sum" or "mean" of run lengths per individual sample
outputName	title prefix (the base name of graph, if savePlots is TRUE)#'
savePlots	should plots be saved out to files or plotted in the graphical terminal (default)?
plot⊺itle	title in plot (default NULL)

#### Value

plot of number of runs vs run-length sum/mean per individual sample

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:</pre>
```

```
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
plot_PatternRuns(runs = runsDF, mapFile = mapFile, method = 'sum')
plot_PatternRuns(runs = runsDF, mapFile = mapFile, method = 'mean')
```

plot\_Runs

Function to plot runs per individual

#### Description

Function to plot runs per individual (see Williams et al. 2016, Animal Genetics, for an example with animal data) Individual IDs on the y-axis, bps on the x-axis (position along the chromosome)

#### Usage

```
plot_Runs(runs, suppressInds = FALSE, savePlots = FALSE,
    separatePlots = FALSE, outputName = NULL)
```

## Arguments

runs	a data.frame with runs per individual (group, id, chrom, nSNP, start, end, length)
suppressInds	shall we suppress individual IDs on the y-axis? (defaults to FALSE)
savePlots	should plots be saved out to files (one pdf file for all chromosomes) or plotted in the graphical terminal (default)?
separatePlots	should plots for each chromosome be saved out to separate files?
outputName	title prefix (the base name of graph, if savePlots is TRUE)

#### Value

plot of runs by chromosome

## Examples

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation</pre>
```

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```
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
# plot runs per animal (interactive)
plot_Runs(runs = runsDF, suppressInds = FALSE, savePlots = FALSE, outputName = "ROHom")
```

plot\_SnpsInRuns Plot the number of times each SNP falls inside runs

#### Description

Function to plot the number of times/percentage each SNP is inside a run (population-specific signals) against the SNP positions in the genome. Proportions on the y-axis, bps on the x-axis

#### Usage

```
plot_SnpsInRuns(runs, genotypeFile, mapFile, savePlots = FALSE,
    separatePlots = FALSE, outputName = NULL)
```

#### Arguments

runs	a data.frame with runs per individual (group, id, chrom, nSNP, start, end, length)
genotypeFile	genotype (.ped) file path
mapFile	map file (.map) file path
savePlots	should plots be saved out in files (default) or plotted in the graphical terminal?
separatePlots	should plots for each chromosome be saved out to separate files?
outputName	title prefix (the base name of graph, if savePlots is TRUE)

#### Value

plot number of times a SNP is in a run by chromosome and population (pdf files)

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
# skipping runs calculation
## Not run:
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,</pre>
```

```
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
# plot runs per animal (interactive)
plot_SnpsInRuns(runs = runsDF, genotypeFile = genotypeFile, mapFile = mapFile,
savePlots = FALSE, outputName = "ROHom")</pre>
```

plot\_StackedRuns Plot stacked runs

#### Description

Function to plot stacked runs along the chromosome (signaling presence of large numbers of runs in specific regions of a chromosome) Counts on the y-axis, bps on the x-axis (position along the chromosome)

#### Usage

```
plot_StackedRuns(runs, savePlots = FALSE, separatePlots = FALSE,
    outputName = NULL)
```

#### Arguments

runs	a data.frame with runs per individual (group, id, chrom, nSNP, start, end, length)
savePlots	should plots be saved out in files (default) or plotted in the graphical terminal?
separatePlots	should plots for chromosomes be saved out to separate files?
outputName	title prefix (the base name of graph, if savePlots is TRUE)

#### Value

plot of stacked runs by population and by chromosome (pdf files)

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)</pre>
```

#### plot\_ViolinRuns

```
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
# plot runs per animal (interactive)
plot_StackedRuns(runs = runsDF, savePlots = FALSE, outputName = "ROHom")</pre>
```

plot\_ViolinRuns *Violin plot of run length per individual (either sum or mean)* 

## Description

Function to produce violin plots of the distribution of runs lengths per group The sum of run lengths, or its average, per individual sample is used to characterize the distribution of runs

#### Usage

```
plot_ViolinRuns(runs, method = c("sum", "mean"), outputName = NULL,
    plotTitle = NULL, savePlots = FALSE)
```

#### Arguments

runs	a data.frame with runs per individual (group, id, chrom, nSNP, start, end, length)
method	"sum" or "mean" of run lengths per individual samples
outputName	title prefix (the base name of graph, if savePlots is TRUE)
plot⊺itle	title in plot (default NULL)
savePlots	should plots be saved out to files or plotted in the graphical terminal (default)?

#### Value

Violin plot of the distribution of runs-lengths (sum or mean)

## Examples

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)</pre>
```

## End(Not run)

```
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
plot_ViolinRuns(runs = runsDF, method = "sum", savePlots = FALSE)
plot_ViolinRuns(runs = runsDF, method = "mean", savePlots = FALSE)</pre>
```

readExternalRuns Read runs from external files

#### Description

Function to read in, from external files, the output of software for ROH:

- 1. detectRUNS: output saved out to a file (e.g. write.table)
- 2. Plink: output from the --homozyg option (.hom files)
- 3. BCFtools: output from the roh option

## Usage

```
readExternalRuns(inputFile = NULL, program = c("plink", "BCFtools",
    "detectRUNS"))
```

#### Arguments

inputFile	name of (path to) external file
program	source program that produced the ROH file (one of detect RUNS, Plink, BCF tools)

#### Value

dataframe in the correct format to be used with plots and statistics functions from detectRUNS

#### Examples

```
# getting map and ped paths
## Not run:
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxMissRun = 1, maxMissWindow = 1, minLengthBps = 100000, minDensity = 1/10000)
write.table(x= runs,file = 'RunsFileTest.txt', quote=F, row.names = F)
newData=readRunsFromFile(runsFile = 'RunsFileTest.txt', program = 'detectRUNS')
## End(Not run)</pre>
```

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readPOPCpp

## Description

This is a core function. Read PED file and returns a data.frame with the first two columns

## Usage

```
readPOPCpp(genotypeFile)
```

## Arguments

genotypeFile genotype (.ped) file location

## Value

a dataframe of POP, ID

reorderDF

Function to reorder data frames by CHROMOSOME

## Description

The data frame will be reordered according to chromosome: from 1 to n, then X, Y, XY, MT The data frame needs to have a column with name "CHROMOSOME"

## Usage

```
reorderDF(dfx)
```

## Arguments

dfx

data frame to be reordered (with column "CHROMOSOME")

## Details

Reorder results based on chromosome

#### Value

A reordered data frame by chromosome

slidingRUNS.run

## Description

This is one of the main function of detectRUNS and is used to detect runs (of homozygosity or heterozygosity) in the genome (diploid) with the sliding-window method. All parameters to detect runs (e.g. minimum n. of SNP, max n. of missing genotypes, max n. of opposite genotypes etc.) are specified here. Input data are in the ped/map Plink format (https://www.cog-genomics.org/plink/1.9/input#ped)

#### Usage

```
slidingRUNS.run(genotypeFile, mapFile, windowSize = 15,
threshold = 0.05, minSNP = 3, ROHet = FALSE, maxOppWindow = 1,
maxMissWindow = 1, maxGap = 10<sup>6</sup>, minLengthBps = 1000,
minDensity = 1/1000, maxOppRun = NULL, maxMissRun = NULL)
```

## Arguments

genotypeFile	genotype (.ped) file path
mapFile	map file (.map) file path
windowSize	the size of sliding window (number of SNP loci) (default = 15)
threshold	the threshold of overlapping windows of the same state (homozygous/heterozygous) to call a SNP in a RUN (default = 0.05)
minSNP	minimum n. of SNP in a RUN (default = 3)
ROHet	should we look for ROHet or ROHom? (default = FALSE)
maxOppWindow	max n. of homozygous/heterozygous SNP in the sliding window (default = 1)
maxMissWindow	max. n. of missing SNP in the sliding window (default = $1$ )
maxGap	max distance between consecutive SNP to be still considered a potential run (default = $10^{6}$ bps)
minLengthBps	minimum length of run in bps (defaults to 1000 bps = 1 kbps)
minDensity	minimum n. of SNP per kbps (defaults to $0.1 = 1$ SNP every 10 kbps)
maxOppRun	max n. of opposite genotype SNPs in the run (optional)
maxMissRun	max n. of missing SNPs in the run (optional)

#### Details

This function scans the genome (diploid) for runs using the sliding-window method. This is a wrapper function for many component functions that handle the input data (ped/map files), perform internal conversions, accept parameters specifications, select whether runs of homozygosity (RoHom) or of heterozygosity (RoHet) are looked for.

#### slidingWindow

In the ped file, the groups samples belong to can be specified (first column). This is important if comparisons between human ethnic groups or between animal breeds or plant varieties or biological populations are to be performed. Also, if cases and controls are to be compared, this is the place where this information needs to be specified.

This function returns a data frame with all runs detected in the dataset. This data frame can then be written out to a csv file. The data frame is, in turn, the input for other functions of the detectRUNS package that create plots and produce statistics from the results (see plots and statistics functions in this manual, and/or refer to the detectRUNS vignette).

## Value

A dataframe with RUNs of Homozygosity or Heterozygosity in the analysed dataset. The returned dataframe contains the following seven columns: "group", "id", "chrom", "nSNP", "from", "to", "lengthBps" (group: population, breed, case/control etc.; id: individual identifier; chrom: chromosome on which the run is located; nSNP: number of SNPs in the run; from: starting position of the run, in bps; to: end position of the run, in bps; lengthBps: size of the run)

## Examples

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs with sliding window approach
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1,
minSNP = 15, ROHet = FALSE, maxOppWindow = 1, maxMissWindow = 1, maxGap=10^6,
minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
colClasses <- c(rep("character", 3), rep("numeric", 4) )
runs <- read.csv2(runsFile, header = TRUE, stringsAsFactors = FALSE,</pre>
```

```
colClasses = colClasses)
```

slidingWindow	Function to slide	a window over a vector	(individual's genotypes)
---------------	-------------------	------------------------	--------------------------

## Description

This is a core function. The functions to detect RUNS are slid over the genome

#### Usage

```
slidingWindow(data, gaps, windowSize, step, maxGap, ROHet = TRUE,
maxOppositeGenotype = 1, maxMiss = 1)
```

## Arguments

data	vector of 0/1/2 genotypes	
gaps	vector of differences between consecutive positions (gaps) in bps	
windowSize	size of window (n. of SNP)	
step	by which (how many SNP) is the window slid	
maxGap	max distance between consecutive SNP in a window to be still considered a potential run	
ROHet maxOppositeGeno	ROHet shall we detect ROHet or ROHom? naxOppositeGenotype	
	max n. of homozygous/heterozygous SNP	
maxMiss	max. n. of missing SNP	

## Value

vector of TRUE/FALSE (whether a window is homozygous or NOT)

slidingWindowCpp Function to slide a window over a vector (individual's genotypes)

## Description

This is a core function. The functions to detect RUNS are slid over the genome

## Usage

```
slidingWindowCpp(data, gaps, windowSize, step, maxGap, ROHet = TRUE,
maxOppositeGenotype = 1L, maxMiss = 1L)
```

## Arguments

data	vector of 0/1/2 genotypes
gaps	vector of differences between consecutive positions (gaps) in bps
windowSize	size of window (n. of SNP)
step	by which (how many SNP) is the window slid
maxGap	max distance between consecutive SNP in a window to be still considered a potential run
ROHet maxOppositeGeno	shall we detect ROHet or ROHom? type
	max n. of homozygous/heterozygous SNP
maxMiss	max. n. of missing SNP

## Value

vector of TRUE/FALSE (whether a window is homozygous or NOT)

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snpInRun Function to return a vector of T/F for whether a SNP is or not in a RUN

## Description

This is a core function. The function to determine whether a SNP is or not in a RUN. The ratio between homozygous/heterozygous windows and total n. of windows is computed here

## Usage

snpInRun(RunVector, windowSize, threshold)

## Arguments

RunVector	vector of TRUE/FALSE (is a window homozygous/heterozygous?)
windowSize	size of window (n. of SNP)
threshold	threshold to call a SNP in a RUN

## Value

vector of TRUE/FALSE (whether a SNP is in a RUN or NOT)

snpInRunCpp	Function to return a vector of T/F for whether a SNP is or not in a
	RUN

## Description

This is a core function. The function to determine whether a SNP is or not in a RUN. The ratio between homozygous/heterozygous windows and total n. of windows is computed here

## Usage

```
snpInRunCpp(RunVector, windowSize, threshold)
```

## Arguments

RunVector	vector of TRUE/FALSE (is a window homozygous/heterozygous?)
windowSize	size of window (n. of SNP)
threshold	threshold to call a SNP in a RUN

## Value

vector of TRUE/FALSE (whether a SNP is in a RUN or NOT)

snpInsideRuns

#### Description

Function to count number of times a SNP is in a RUN

#### Usage

snpInsideRuns(runsChrom, mapChrom, genotypeFile)

#### Arguments

runsChrom	R object (dataframe) with results per chromosome (column names: "POPULATION", "IND", "CHROMOS
mapChrom	R object (dataframe) with SNP name and position per chromosome (map file) (column names: "CHR", "SNP_NAME", "x", "POSITION")
genotypeFile	genotype (.ped) file location

## Value

dataframe with counts per SNP in runs (per population)

## Examples

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")</pre>
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")</pre>
# defining mapChrom
mappa <- data.table::fread(mapFile, header = FALSE)</pre>
names(mappa) <- c("CHR","SNP_NAME","x","POSITION")</pre>
mappa$x <- NULL</pre>
chrom <- "24"
mapChrom <- mappa[mappa$CHR==chrom, ]</pre>
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")</pre>
colClasses <- c(rep("character", 3), rep("numeric", 4) )</pre>
```

# fix column names and define runsChrom

colClasses = colClasses)

runs <- read.csv2(runsFile, header = TRUE, stringsAsFactors = FALSE,</pre>

## snpInsideRunsCpp

```
names(runs) <- c("POPULATION","IND","CHROMOSOME","COUNT","START","END","LENGTH")
runsChrom <- runs[runs$CHROMOSOME==chrom, ]</pre>
```

```
snpInsideRuns(runsChrom, mapChrom, genotypeFile)
```

snpInsideRunsCpp Function to count number of times a SNP is in a RUN

## Description

Function to count number of times a SNP is in a RUN

## Usage

snpInsideRunsCpp(runsChrom, mapChrom, genotypeFile)

#### Arguments

runsChrom	R object (dataframe) with results per chromosome
mapChrom	R map object with SNP per chromosome
genotypeFile	genotype (.ped) file location

## Value

dataframe with counts per SNP in runs (per population)

summaryRuns	Summary statistics on detected runs
-------------	-------------------------------------

## Description

This function processes the results from slidingRUNS.run and consecutiveRUNS.run and produces a number of interesting descriptives statistics on results.

#### Usage

```
summaryRuns(runs, mapFile, genotypeFile, Class = 2, snpInRuns = FALSE)
```

#### Arguments

runs	R object (dataframe) with results on detected runs
mapFile	Plink map file (for SNP position)
genotypeFile	Plink ped file (for SNP position)
Class	group of length (in Mbps) by class (default: 0-2, 2-4, 4-8, 8-16, >16)
snpInRuns	TRUE/FALSE (default): should the function snpInsideRuns be called to compute the proportion of times each SNP falls inside a run in the group/population?

#### Details

summaryRuns calculates: i) the number of runs per chromosome and group/population; ii) the percent distribution of runs per chromosome and group; iii) the number of runs per size-class and group; iv) the percent distribution of runs per size-class and group; v) the mean length of runs per chromosome and group; vi) the mean length of runs per size-class and group; vii) individual inbreeding coefficient estimated from ROH; viii) individual inbreeding coefficient estimated from ROH per chromosome; ix) individual inbreeding coefficient estimated from ROH per size-class

## Value

A list of dataframes containing the most relevant descriptives statistics on detected runs. The list conveniently contains 9 dataframes that can be used for further processing and visualization, or can be written out to text files

#### Examples

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF <- readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
summaryRuns(runs = runsDF, mapFile = mapFile, genotypeFile = genotypeFile, Class = 2,
snpInRuns = FALSE)
```

tableRuns

Function to retrieve most common runs in the population

#### Description

This function takes in input either the run results or the output from the function snpInsideRuns (proportion of times a SNP is inside a run) in the population/group, and returns a subset of the runs most commonly found in the group/population. The parameter threshold controls the definition of most common (e.g. in at least 50%, 70% etc. of the sampled individuals)

#### Usage

```
tableRuns(runs = NULL, SnpInRuns = NULL, genotypeFile, mapFile,
threshold = 0.5)
```

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## writeRUN

#### Arguments

runs	R object (dataframe) with results on detected runs
SnpInRuns	dataframe with the proportion of times each SNP falls inside a run in the population (output from snpInsideRuns)
genotypeFile	Plink ped file (for SNP position)
mapFile	Plink map file (for SNP position)
threshold	value from 0 to 1 (default 0.7) that controls the desired proportion of individuals carrying that run (e.g. 70%)

#### Value

A dataframe with the most common runs detected in the sampled individuals (the group/population, start and end position of the run, chromosome and number of SNP included in the run are reported in the output dataframe)

#### Examples

```
# getting map and ped paths
genotypeFile <- system.file("extdata", "Kijas2016_Sheep_subset.ped", package = "detectRUNS")
mapFile <- system.file("extdata", "Kijas2016_Sheep_subset.map", package = "detectRUNS")
# calculating runs of Homozygosity
## Not run:
# skipping runs calculation
runs <- slidingRUNS.run(genotypeFile, mapFile, windowSize = 15, threshold = 0.1, minSNP = 15,
ROHet = FALSE, maxOppositeGenotype = 1, maxMiss = 1, minLengthBps = 100000, minDensity = 1/10000)
## End(Not run)
# loading pre-calculated data
runsFile <- system.file("extdata", "Kijas2016_Sheep_subset.sliding.csv", package="detectRUNS")
runsDF = readExternalRuns(inputFile = runsFile, program = 'detectRUNS')
```

tableRuns(runs = runsDF, genotypeFile = genotypeFile, mapFile = mapFile, threshold = 0.5)

writeRUN

Function to write out RUNS per individual animal

## Description

Function to write out RUNS per individual animal

#### Usage

```
writeRUN(ind, dRUN, ROHet = TRUE, group, outputName)
```

## Arguments

ind	ID of animals
dRUN	data.frame with RUNS per animal
ROHet	shall we detect ROHet or ROHom?
group	group (factor): population, breed, ethnicity, case/control etc.
outputName	output filename

## Value

TRUE/FALSE if RUNS are written out or not

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