

Package ‘evola’

July 22, 2025

Version 1.0.5

Date 2025-04-24

Title Evolutionary Algorithm

Maintainer Giovanni Covarrubias-Pazaran <cova_ruber@live.com.mx>

Description Runs an evolutionary algorithm using the 'AlphaSimR' machinery <[doi:10.1093/g3journal/jkaa017](https://doi.org/10.1093/g3journal/jkaa017)> .

Depends R(>= 3.5.0), AlphaSimR (>= 1.4.2), Matrix (>= 1.0), methods, crayon

LazyLoad yes

LazyData yes

License GPL (>= 2)

NeedsCompilation yes

Author Giovanni Covarrubias-Pazaran [aut, cre] (ORCID: <<https://orcid.org/0000-0002-7194-3837>>)

Repository CRAN

Suggests rmarkdown, knitr

VignetteBuilder knitr

Config/testthat/edition 3

Date/Publication 2025-04-26 03:10:03 UTC

Contents

evola-package	2
A.mat	3
addZeros	4
bestSol	5
DT_cpdata	7
DT_technow	8
DT_wheat	10
evolafit	12
evolaPop-class	18

importHaploSparse	19
inbFun	21
Jc	22
Jr	23
logspace	24
nQtl	24
ocsFun	26
overlay	27
pareto	28
pmonitor	29
regFun	30
stan	31
update.evolaFitMod	32
varQ	33
Index	35

evola-package	EVOLutionary Algorithm
---------------	-------------------------------

Description

The evola package is nice wrapper of the AlphaSimR package that enables the use of the evolutionary algorithm to solve complex questions in a simple form.

The `evolafit` function is the core function of the package which allows the user to specify the problem and constraints to find a close-to-optimal solution using the evolutionary forces.

Keeping evola updated

The evola package is updated on CRAN every 4-months due to CRAN policies but you can find the latest source at <https://github.com/covaruber/evola>. This can be easily installed typing the following in the R console:

```
library(devtools)
install_github("covaruber/evola")
```

This is recommended if you reported a bug, was fixed and was immediately pushed to GitHub but not in CRAN until the next update.

Tutorials

For tutorials on how to perform different analysis with evola please look at the vignettes by typing in the terminal:

```
vignette("evola.intro")
```

Getting started

- The package has been equipped with a couple of datasets to learn how to use the evola package:
- * [DT_technow](#) dataset to perform optimal cross selection.
 - * [DT_wheat](#) dataset to perform optimal training population selection.
 - * [DT_cpdata](#) dataset to perform optimal individual.

Models Enabled

The machinery behind the scenes is AlphaSimR.

Bug report and contact

If you have any questions or suggestions please post it in <https://stackoverflow.com> or <https://stats.stackexchange.com>
I'll be glad to help or answer any question. I have spent a valuable amount of time developing this package. Please cite this package in your publication. Type 'citation("evola")' to know how to cite it.

Author(s)

Giovanny Covarrubias-Pazaran

References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. G3 Gene|Genomes|Genetics 11(2):jkaa017. <https://doi.org/10.1093/g3journal/jkaa017>.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. Genome Research, 19, 136-142. <http://genome.cshlp.org/content/19/1/136>.

A.mat	Additive relationship matrix
-------	------------------------------

Description

Calculates the realized additive relationship matrix.

Usage

A.mat(X,min.MAF=NULL)

Arguments

X	Matrix ($n \times m$) of unphased genotypes for n lines and m biallelic markers, coded as $\{-1,0,1\}$. Fractional (imputed) and missing values (NA) are allowed.
min.MAF	Minimum minor allele frequency. The A matrix is not sensitive to rare alleles, so by default only monomorphic markers are removed.

Details

For vanraden method: the marker matrix is centered by subtracting column means $M = X - ms$ where ms is the column means. Then $A = MM'/c$, where $c = \sum_k d_k/k$, the mean value of the diagonal values of the MM' portion.

Value

If `return.imputed = FALSE`, the $n \times n$ additive relationship matrix is returned.

If `return.imputed = TRUE`, the function returns a list containing

\$A the A matrix

References

Giovanny Covarrubias-Pazaran (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
## random population of 200 lines with 1000 markers
X <- matrix(rep(0,200*1000),200,1000)
for (i in 1:200) {
  X[i,] <- ifelse(runif(1000)<0.5,-1,1)
}

A <- A.mat(X)

## take a look at the Genomic relationship matrix
colfunc <- colorRampPalette(c("steelblue4","springgreen","yellow"))
hv <- heatmap(A[1:15,1:15], col = colfunc(100),Colv = "Rowv")
str(hv)
```

addZeros

Function to add zeros before and after a numeric vector to have the same number of characters.

Description

Function to add zeros before and after a numeric vector to have the same number of characters.

Usage

```
addZeros(x, nr=2)
```

Arguments

- x Numeric vector.
- nr number of digits to keep to the right.

Details

A simple apply function to make a matrix of one row and nc columns.

Value

\$res a matrix

References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
addZeros(5)
```

bestSol	<i>Extract the index of the best solution</i>
---------	---

Description

Extracts the index of the best solution for all traits under the constraints specified.

Usage

```
bestSol(object, selectTop=TRUE, n=1)
```

Arguments

- object A resulting object from the function evolafit.
- selectTop Selects highest values for the fitness value if TRUE. Selects lowest values if FALSE.
- n An integer indicating how many solutions should be returned.

Details

A simple apply function looking at the fitness value of all the solution in the last generation to find the maximum value.

Value

\$res the vector of best solutions in **M** for each trait in the problem

References

Giovanny Covarrubias-Pazarán (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
set.seed(1)
# Data
Gems <- data.frame(
  Color = c("Red", "Blue", "Purple", "Orange",
            "Green", "Pink", "White", "Black",
            "Yellow"),
  Weight = round(runif(9,0.5,5),2),
  Value = round(abs(rnorm(9,0,5))+0.5,2),
  Times=c(rep(1,8),0)
)
head(Gems)

# Task: Gem selection.
# Aim: Get highest combined value.
# Restriction: Max weight of the gem combined = 10.
res0<-evolafit(cbind(Weight,Value)~Color, dt= Gems,
               # constraints: if greater than this ignore
               constraintsUB = c(10,Inf),
               # constraints: if smaller than this ignore
               constraintsLB= c(-Inf,-Inf),
               # weight the traits for the selection
               b = c(0,1),
               # population parameters
               nCrosses = 100, nProgeny = 20, recombGens = 1,
               # coancestry parameters
               D=NULL, lambda=c(0,0), nQTLperInd = 1,
               # selection parameters
               propSelBetween = .9, propSelWithin =0.9,
               nGenerations = 50
)
```

```
bestSol(res0$pop, n=2)
```

DT_cpdata

Genotypic and Phenotypic data for a CP population

Description

A CP population or F1 cross is the designation for a cross between 2 highly heterozygote individuals; i.e. humans, fruit crops, breeding populations in recurrent selection.

This dataset contains phenotypic data for 363 siblings for an F1 cross. These are averages over 2 environments evaluated for 4 traits; color, yield, fruit average weight, and firmness. The columns in the CPgeno file are the markers whereas the rows are the individuals. The CPpheno data frame contains the measurements for the 363 siblings, and as mentioned before are averages over 2 environments.

Usage

```
data("DT_cpdata")
```

Format

The format is: chr "DT_cpdata"

Source

This data was simulated for fruit breeding applications.

References

Giovanny Covarrubias-Pazarán (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. *G3 GeneGenomesGenetics* 11(2):jkaa017. <https://doi.org/10.1093/g3journal/jkaa017>.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. *Genome Research*, 19, 136-142. <http://genome.cshlp.org/content/19/1/136>.

Examples

```
data(DT_cpdata)
DT <- DT_cpdata
A <- A[DT$id,DT$id]

# get best 20 individuals weighting variance by ~0.5=(30*pi)/180
res<-evolafit(formula=cbind(Yield, occ)~id, dt= DT,
              # constraints: if sum is greater than this ignore
```

```

constraintsUB = c(Inf,20),
# constraints: if sum is smaller than this ignore
constraintsLB= c(-Inf,-Inf),
# weight the traits for the selection
b = c(1,0),
# population parameters
nCroses = 100, nProgeny = 10,
recombGens=1, nChr=1, mutRate=0,
# coancestry parameters
D=A, lambda= (30*pi)/180 , nQTLperInd = 20,
# selection parameters
propSelBetween = 0.5, propSelWithin =0.5,
nGenerations = 40)

Q <- pullQtlGeno(res$pop, simParam = res$simParam, trait=1); Q <- Q/2
best = bestSol(res$pop)[,"Yield"];best
qa = (Q %%% DT$Yield)[best,]; qa
qDq = Q[best,] %%% A %%% Q[best,]; qDq
sum(Q[best,]) # total # of inds selected

pmonitor(res)

plot(DT$Yield, col=as.factor(Q[best,]),
     pch=(Q[best,]*19)+1)

pareto(res)

```

DT_technow

Genotypic and Phenotypic data from single cross hybrids (Technow et al.,2014)

Description

This dataset contains phenotypic data for 2 traits measured in 1254 single cross hybrids coming from the cross of Flint x Dent heterotic groups. In addition contains the genotypic data (35,478 markers) for each of the 123 Dent lines and 86 Flint lines. The purpose of this data is to demonstrate the prediction of unrealized crosses (9324 unrealized crosses, 1254 evaluated, total 10578 single crosses). We have added the additive relationship matrix (A) but can be easily obtained using the A.mat function on the marker data. Please if using this data for your own research cite Technow et al. (2014) publication (see References).

Usage

```
data("DT_technow")
```

Format

The format is: chr "DT_technow"

Source

This data was extracted from Technow et al. (2014).

References

If using this data for your own research please cite:

Technow et al. 2014. Genome properties and prospects of genomic predictions of hybrid performance in a Breeding program of maize. *Genetics* 197:1343-1355.

Giovanny Covarrubias-Pazarán (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to *Bioinformatics*.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. *G3 Genes|Genomes|Genetics* 11(2):jkaa017. <https://doi.org/10.1093/g3journal/jkaa017>.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. *Genome Research*, 19, 136-142. <http://genome.cshlp.org/content/19/1/136>.

Examples

```
data(DT_technow)
DT <- DT_technow
DT$occ <- 1; DT$occ[1]=0
M <- M_technow

A <- A.mat(M)
A <- A[DT$hy,DT$hy]
# run the genetic algorithm
# we assign a weight to x'Dx of (20*pi)/180=0.34
res<-evolafit(formula = c(GY, occ)~hy,
              dt= DT,
              # constraints: if sum is greater than this ignore
              constraintsUB = c(Inf,100),
              # constraints: if sum is smaller than this ignore
              constraintsLB= c(-Inf,-Inf),
              # weight the traits for the selection
              b = c(1,0),
              # population parameters
              nCrosses = 100, nProgeny = 10,
              recombGens=1, nChr=1, mutRate=0,
              # coancestry parameters
              D=A, lambda= (20*pi)/180 , nQTLperInd = 90,
              # selection parameters
              propSelBetween = 0.5, propSelWithin =0.5,
              nGenerations = 20)

Q <- pullQtlGeno(res$pop, simParam = res$simParam, trait=1); Q <- Q/2
best = bestSol(res$pop[, "GY"]
qa = (Q %*% DT$GY)[best,]; qa
qAq = Q[best,] %*% A %*% Q[best,]; qAq
sum(Q[best,]) # total # of inds selected
```

```
pmonitor(res)
plot(DT$GY, col=as.factor(Q[best,]),
     pch=(Q[best,]*19)+1)

pareto(res)
```

DT_wheat

wheat lines dataset

Description

Information from a collection of 599 historical CIMMYT wheat lines. The wheat data set is from CIMMYT's Global Wheat Program. Historically, this program has conducted numerous international trials across a wide variety of wheat-producing environments. The environments represented in these trials were grouped into four basic target sets of environments comprising four main agroclimatic regions previously defined and widely used by CIMMYT's Global Wheat Breeding Program. The phenotypic trait considered here was the average grain yield (GY) of the 599 wheat lines evaluated in each of these four mega-environments.

A pedigree tracing back many generations was available, and the Browse application of the International Crop Information System (ICIS), as described in (McLaren *et al.* 2000, 2005) was used for deriving the relationship matrix A among the 599 lines; it accounts for selection and inbreeding.

Wheat lines were recently genotyped using 1447 Diversity Array Technology (DArT) generated by Triticaret Pty. Ltd. (Canberra, Australia; <http://www.triticarte.com.au>). The DArT markers may take on two values, denoted by their presence or absence. Markers with a minor allele frequency lower than 0.05 were removed, and missing genotypes were imputed with samples from the marginal distribution of marker genotypes, that is, $x_{ij} = \text{Bernoulli}(\hat{p}_j)$, where $\hat{p}_j$ is the estimated allele frequency computed from the non-missing genotypes. The number of DArT MMs after edition was 1279.

Usage

```
data(DT_wheat)
```

Format

Matrix Y contains the average grain yield, column 1: Grain yield for environment 1 and so on.

Source

International Maize and Wheat Improvement Center (CIMMYT), Mexico.

References

- Giovanny Covarrubias-Pazarán (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to *Bioinformatics*.
- Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. *G3 Genes|Genomes|Genetics* 11(2):jkaa017. <https://doi.org/10.1093/g3journal/jkaa017>.
- Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. *Genome Research*, 19, 136-142. <http://genome.cshlp.org/content/19/1/136>.
- McLaren, C. G., L. Ramos, C. Lopez, and W. Eusebio. 2000. "Applications of the genealogy management system." In *International Crop Information System. Technical Development Manual, version VI*, edited by McLaren, C. G., J.W. White and P.N. Fox. pp. 5.8-5.13. CIMMYT, Mexico: CIMMYT and IRRI.
- McLaren, C. G., R. Bruskiewich, A.M. Portugal, and A.B. Cosico. 2005. The International Rice Information System. A platform for meta-analysis of rice crop data. *Plant Physiology* **139**: 637-642.

Examples

```
# example to optimize a training pop for a validation pop
data(DT_wheat)
DT <- as.data.frame(DT_wheat)
DT$id <- rownames(DT) # IDs
DT$occ <- 1; DT$occ[1]=0 # to track occurrences
DT$dummy <- 1; DT$dummy[1]=0 # dummy trait

# if genomic
GT <- GT_wheat + 1; rownames(GT) <- rownames(DT)
G <- GT%*%t(GT)
G <- G/mean(diag(G))
# if pedigree
A <- A_wheat
A[1:4,1:4]
##Perform eigenvalue decomposition for clustering
##And select cluster 5 as target set to predict
pcNum=25
svdWheat <- svd(A, nu = pcNum, nv = pcNum)
PCWheat <- A %*% svdWheat$v
rownames(PCWheat) <- rownames(A)
DistWheat <- dist(PCWheat)
TreeWheat <- cutree(hclust(DistWheat), k = 5 )
plot(PCWheat[,1], PCWheat[,2], col = TreeWheat,
     pch = as.character(TreeWheat), xlab = "pc1", ylab = "pc2")
vp <- rownames(PCWheat)[TreeWheat == 3]; length(vp)
tp <- setdiff(rownames(PCWheat),vp)

As <- A[tp,tp]
DT2 <- DT[rownames(As),]
DT2$cov <- apply(A[tp,vp],1,mean)

head(DT2)
```

```

# we assign a weight to x'Dx of (60*pi)/180=1
res<-evolafit(formula=cbind(cov, occ)~id, dt= DT2,
              # constraints: if sum is greater than this ignore
              constraintsUB = c(Inf, 100),
              # constraints: if sum is smaller than this ignore
              constraintsLB= c(-Inf, -Inf),
              # weight the traits for the selection
              b = c(1,0),
              # population parameters
              nCrosses = 100, nProgeny = 10,
              recombGens=1, nChr=1, mutRate=0,
              # coancestry parameters
              D=As, lambda= (60*pi)/180 , nQTLperInd = 90,
              # selection parameters
              propSelBetween = 0.5, propSelWithin =0.5,
              nGenerations = 30, verbose = TRUE)

Q <- pullQtlGeno(res$pop, simParam = res$simParam, trait=1); Q <- Q/2
best <- bestSol(res$pop)[,"cov"]
sum(Q[best,]) # total # of inds selected
pareto(res)

```

evolafit

Fits a genetic algorithm for a set of traits and constraints.

Description

Using the AlphaSimR machinery it recreates the evolutionary forces applied to a problem where possible solutions replace individuals and combinations of variables to optimize in the problem replace the genes or QTLs. Then evolutionary forces (mutation, selection and drift) are applied to find a close-to-optimal solution. Although multiple traits are enabled it is assumed that same QTLs are behind all the traits, differing only in their average allelic effects.

Usage

```

evolafit(formula, dt,
          constraintsUB, constraintsLB, b,
          nCrosses=50, nProgeny=20,nGenerations=20,
          recombGens=1, nChr=1, mutRate=0,
          nQTLperInd=NULL, D=NULL, lambda=0,
          propSelBetween=NULL,propSelWithin=NULL,
          fitnessf=NULL, verbose=TRUE, dateWarning=TRUE,
          selectTop=TRUE, tolVarG=1e-6,
          Ne=50, initPop=NULL, simParam=NULL,
          fixQTLperInd=FALSE, traceDelta=TRUE, topN=10,

```

...)

Arguments

formula	Formula of the form $y \sim x$ where y refers to the average allelic substitution effects of the QTLs (alpha) for each trait, and x refers to the variable defining the genes or QTLs to be combined in the possible solutions.
dt	A dataset containing the average allelic effects (a) and classifiers/genes/QTLs.
constraintsUB	A numeric vector specifying the upper bound constraints for the breeding values applied at each trait. The length is equal to the number of traits/features in the formula. If missing is assume an infinite value for all traits. Solutions (individuals in the population) with higher value than the upper bound are assigned a -infinite value if the argument <code>selectTop=TRUE</code> and to +infinite when <code>selectTop=FALSE</code> , which is equivalent to reject/discard a solution based on the fitness function.
constraintsLB	A numeric vector specifying the lower bound constraints for the breeding values applied at each trait. The length is equal to the number of traits/features in the formula. If missing is assume a -infinite value for all traits. Solutions with lower value than the lower bound are assigned a +infinite value if the argument <code>selectTop=TRUE</code> and to -infinite when <code>selectTop=FALSE</code> , which is equivalent to reject/discard a solution based on the fitness function.
b	A numeric vector specifying the weights that each trait has in the fitness function (i.e., a selection index). The length should be equal to the number of traits/features. If missing is assumed equal weight (1) for all traits.
nCrosses	A numeric value indicating how many crosses should occur in the population of solutions at every generation.
nProgeny	A numeric value indicating how many progeny (solutions) each cross should generate in the population of solutions at every generation.
nGenerations	The number of generations that the evolutionary process should run for.
recombGens	The number of recombination generations that should occur before selection is applied. This is in case the user wants to allow for more recombination before selection operates. The default is 1.
nChr	The number of chromosomes where features/genes should be allocated to. The default value is 1 but this number can be increased to mimic more recombination events at every generation and avoid linkage disequilibrium.
mutRate	A value between 0 and 1 to indicate the proportion of random QTLs that should mutate in each individual. For example, a value of 0.1 means that a random 10% of the QTLs will mutate in each individual randomly taking values of 0 or 1. Is important to notice that this implies that the stopping criteria based in variance will never be reached because we keep introducing variance through random mutation.
nQTLperInd	The number of QTLs/genes (classifier x in the formula) that should be fixed for the positive allele at the beginning of the simulation. If not specified it will be equal to the 20% of the QTLs (calculated as the number of rows in the <code>dt</code>

argument over 5). This is just an initial value and will change as the population evolve under the constraints specified by the user. See details section.

D	A relationship matrix between the QTLs (a kind of linkage disequilibrium) specified in the right side of the formula (levels of the x variable). This matrix can be used or ignored in the fitness function. By default the weight to the $q'Dq$ component is 0 though the lambda argument, where x is an individual in the population of a solution.
lambda	A numeric value indicating the weight assigned to the relationship between QTLs in the fitness function. If not specified is assumed to be 0. This can be used or ignored in your customized fitness function.
propSelBetween	A numeric value between 0 and 1 indicating the proportion of families/crosses of solutions/individuals that should be selected. The default is 1, meaning all crosses are selected or passed to the next generation.
propSelWithin	A numeric value between 0 and 1 indicating the proportion of individuals/solutions within families/crosses that should be selected. The default value is 0.5, meaning that 50% of the top individuals are selected.
fitnessf	An alternative fitness function to be applied at the level of individuals or solutions. It could be a linear combination of the trait breeding values. The available variables internally are: Y: matrix of trait breeding values for the individuals/solutions. Of dimensions $s \times t$, s solutions and t traits. b: vector of trait weights, specified in the 'b' argument. Of dimensions $t \times 1$, t traits by 1 Q: matrix with QTLs for the individuals/solutions. Of dimensions $s \times p$, s solutions and p QTL columns. Although multiple traits are enabled it is assumed that same QTLs are behind all the traits, differing only in their average allelic effects. D: matrix of relationship between the QTLs, specified in the 'D' argument. Of dimensions $p \times p$, for p QTL columns lambda: a numeric value indicating the weight assigned to the relationship between QTLs in the fitness function. If not specified is assumed to be 0. This can be used or ignored in your customized fitness function. a: list of vectors with average allelic effects for a given trait. Of dimensions $s \times 1$, s solutions by 1 column If <code>fitnessf=NULL</code>, the default function will be the ocsFun function: <pre>function(Y,b,d,Q,D,a,lambda){(Y%*%b) - d}</pre> where $(Y\%* \%b)$ is equivalent to $[(Q'a)b]$ in genetic contribution theory, and d is equal to the diagonal values from $Q'DQ$ from contribution theory, <i>If you provide your own fitness function please keep in mind that the variables Y, b, Q, D, a, and lambda are already reserved and these variables should always be added to your function (even if you do not use them) in addition to your new variables so the machinery runs.</i> An additional fitness function for accounting only for the group relationship is inbFun when the user wants to find solutions that maximize the representativeness of a sample and the D argument is not NULL. You will need to select the

	solutions with lower values (<code>selectTop=TRUE</code>) which indicate solutions with more representativeness and you may need to indicate lower bound constraints (<code>constraintsLB</code>).
	An additional fitness function available for regression problems is <code>regFun</code> but is not the default since it would require additional arguments not available in a regular genetic algorithm problem (e.g., y and X to compute $y-Xb$).
<code>verbose</code>	A logical value indicating if we should print logs.
<code>dateWarning</code>	A logical value indicating if you should be warned when there is a new version on CRAN.
<code>selectTop</code>	Selects highest values for the fitness value if <code>TRUE</code> . Selects lowest values if <code>FALSE</code> .
<code>tolVarG</code>	A stopping criteria (tolerance for genetic variance) when the variance across traits is smaller than this value, which is equivalent to assume that all solutions having the same QTL profile (depleted variance). The default value is $1e-6$ and is computed as the sum of the diagonal values of the genetic variance covariance matrix between traits.
<code>Ne</code>	initial number of founders in the population (will be important for long term sustainability of genetic variance).
<code>initPop</code>	an object of <code>Pop</code> -class.
<code>simParam</code>	an object of <code>SimParam</code> .
<code>fixQTLperInd</code>	A <code>TRUE/FALSE</code> value to indicate if we should fix the argument <code>nQTLperInd</code> across all generations. This should be used with care since this is not how usually genetic algorithms work and in my experience only using GA for regression problems is a special case where this argument should be set to <code>TRUE</code> . The behavior assumes that if set to <code>TRUE</code> and a particular solution has more QTLs active than <code>nQTLperInd</code> some QTLs will be set to 0 and if a solution has less QTLs active than <code>nQTLperInd</code> some QTLs will be activated. All activations or deactivations are done at random. This only takes place after generation 1.
<code>traceDelta</code>	a logical value indicating if we should compute the rate of coancestry $Q'DQ$ at each iteration. This metric is used by the pareto plot but is not needed for the evolutionary process and it can take a considerable amount of time when the number of QTLs is big.
<code>topN</code>	an integer value indicating the maximum number of solutions to keep in each generation.
<code>...</code>	Further arguments to be passed to the fitness function if required.

Details

Using the AlphaSimR machinery (`runMacs`) it recreates the evolutionary forces applied to a problem where possible solutions replace individuals and combinations of variables in the problem replace the genes. Then evolutionary forces are applied to find a close-to-optimal solution. The number of solutions are controlled with the `nCrosses` and `nProgeny` parameters, whereas the number of initial QTLs activated in a solution is controlled by the `nQTLperInd` parameter. The number of activated QTLs of course will increase if has a positive effect in the fitness of the solutions. The drift force can be controlled by the `recombGens` parameter. The mutation rate can be controlled with the `mutRate` parameter. The recombination rate can be controlled with the `nChr` argument.

The `indivPerformance` output slot contains the columns `id`, `fitness`, `generation`, `nQTLs`, and `deltaC`. These mean the following:

In `fitness` : represents the fitness function value of a solution.

In `deltaC` : it represents the change in coancestry (e.g., inbreeding), it can be thought as the rate of coancestry. It is calculated as $q'Dq$ where ' q ' represents the contribution vector, ' D ' is the linkage disequilibrium matrix between QTNs (whatever the QTNs represent for your specific problem). In practice we do QAQ' and extract the diagonal values.

In `generation` : it represents the generation at which this solution appeared.

In `nQTLs` : it represent the final number of QTNs that are activated in homozygote state for the positive effect.

During the run the columns printed in the console mean the following:

generation: generation of reproduction

constrainedUB: number of solutions constrained by the upper bound specified

constrainedLB: number of solutions constrained by the lower bound specified

varG: genetic variance present in the population due to the QTNs

propB: proportion of families selected during that iteration

propW: proportion of individuals within a family selected in that iteration

time: the time when the iteration has finished.

Value

indivPerformance the matrix of fitness, `deltaC`, `generation`, `nQTLs` per solution per generation. See details section above.

pedBest contains the pedigree of the selected solutions across iterations.

\$score a matrix with scores for different metrics across `n` generations of evolution.

\$pheno the matrix of phenotypes of individuals/solutions present in the last generation.

pop AlphaSimR object used for the evolutionary algorithm at the last iteration.

constCheckUB A matrix with as many rows as solutions and columns as traits to be constrained. 0s indicate that such trait went beyond the bound in that particular solution.

constCheckLB A matrix with as many rows as solutions and columns as traits to be constrained. 0s indicate that such trait went beyond the bound in that particular solution.

traits a character vector corresponding to the name of the variables used in the fitness function.

References

Giovanny Covarrubias-Pazaran (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. *G3 GenesGenomesGenetics* 11(2):jkaa017. <https://doi.org/10.1093/g3journal/jkaa017>.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. *Genome Research*, 19, 136-142. <http://genome.cshlp.org/content/19/1/136>.

See Also

[evolafit](#) – the information of the package

Examples

```
set.seed(1)

# Data
Gems <- data.frame(
  Color = c("Red", "Blue", "Purple", "Orange",
            "Green", "Pink", "White", "Black",
            "Yellow"),
  Weight = round(runif(9,0.5,5),2),
  Value = round(abs(rnorm(9,0,5))+0.5,2),
  Times=c(rep(1,8),0)
)
head(Gems)
#   Color Weight Value
# 1   Red   4.88  9.95
# 2  Blue   1.43  2.73
# 3 Purple   1.52  2.60
# 4 Orange   3.11  0.61
# 5  Green   2.49  0.77
# 6  Pink   3.53  1.99
# 7  White   0.62  9.64
# 8  Black   2.59  1.14
# 9 Yellow   1.77 10.21

# Task: Gem selection.
# Aim: Get highest combined value.
# Restriction: Max weight of the gem combined = 10.

# simple specification
res00<-evolafit(formula=cbind(Weight,Value)~Color, dt= Gems,
               # constraints on traits: if greater than this ignore
               constraintsUB = c(10,Inf), nGenerations = 10
)
best = bestSol(res00$pop)[,"Value"]
Q <- pullQtI geno(res00$pop, simParam = res00$simParam, trait=1); Q <- Q/2
qa = Q[best,] %*% as.matrix(Gems[,c("Weight","Value")]); qa

# more complete specification
res0<-evolafit(formula=cbind(Weight,Value)~Color, dt= Gems,
               # constraints on traits: if greater than this ignore
               constraintsUB = c(10,Inf),
               # constraints on traits: if smaller than this ignore
               constraintsLB= c(-Inf,-Inf),
               # weight the traits for the selection (fitness function)
               b = c(0,1),
```

```

# population parameters
nCcrosses = 100, nProgeny = 20,
# genome parameters
recombGens = 1, nChr=1, mutRate=0, nQTLperInd = 2,
# coancestry parameters
D=NULL, lambda=0,
# selection parameters
propSelBetween = .9, propSelWithin =0.9,
nGenerations = 50
)

Q <- pullQtlGeno(res0$pop, simParam = res0$simParam, trait=2); Q <- Q/2
best = bestSol(res0$pop)[,"Value"]
qa = Q[best,] %*% as.matrix(Gems[,c("Weight", "Value")]); qa
Q[best,]

# $`Genes`
# Red   Blue Purple Orange   Green   Pink   White   Black Yellow
# 1     1     0     0     1     0     0     1     0
#
# $Result
# Weight Value
# 8.74  32.10
pmonitor(res0)
pareto(res0)

```

evolaPop-class

Genetic algorithm pop

Description

A genetic algorithm pop fit by [evolafit](#). This class extends class "[Pop](#)" class and includes some additional slots.

Objects from the Class

Objects are created by calls to the [evolafit](#) function.

Slots

indivPerformance the matrix of q'a (score), deltaC, q'Dq, generation, nQTNs per solution per generation. See details section above. All other slots are inherited from class "[Pop](#)".

pedBest if the argument keepBest=TRUE this contains the pedigree of the selected solutions across iterations. All other slots are inherited from class "[Pop](#)".

\$score a matrix with scores for different metrics across n generations of evolution. All other slots are inherited from class "[Pop](#)".

\$pheno the matrix of phenotypes of individuals/solutions present in the last generation. All other slots are inherited from class "Pop".

\$phenoBest the matrix of phenotypes of top (parents) individuals/solutions present in the last generation. All other slots are inherited from class "Pop".

constCheckUB A matrix with as many rows as solutions and columns as traits to be constrained. 0s indicate that such trait went beyond the bound in that particular solution. All other slots are inherited from class "Pop".

constCheckLB A matrix with as many rows as solutions and columns as traits to be constrained. 0s indicate that such trait went beyond the bound in that particular solution. All other slots are inherited from class "Pop".

traits a character vector corresponding to the name of the variables used in the fitness function. All other slots are inherited from class "Pop".

Extends

Class "Pop", directly.

Methods

update signature(object = "evolaPop"): also a non-method for the same reason as update

See Also

evolafit

Examples

```
showClass("evolaPop")
```

importHaploSparse	<i>Import haplotypes</i>
-------------------	--------------------------

Description

Formats haplotype in a matrix format to an AlphaSimR population that can be used to initialize a simulation. This function serves as wrapper for newMapPop that utilizes a more user friendly input format.

Usage

```
importHaploSparse(haplo, genMap, ploidy = 2L, ped = NULL)
```

Arguments

haplo	a sparse matrix of haplotypes
genMap	genetic map as a data.frame. The first three columns must be: marker name, chromosome, and map position (Morgans). Marker name and chromosome are coerced using as.character. See importGenMap.
ploidy	ploidy level of the organism.
ped	an optional pedigree for the supplied genotypes. See details.

Details

The optional pedigree can be a data.frame, matrix or a vector. If the object is a data.frame or matrix, the first three columns must include information in the following order: id, mother, and father. All values are coerced using as.character. If the object is a vector, it is assumed to only include the id. In this case, the mother and father will be set to "0" for all individuals.

Value

\$res a MapPop-class if ped is NULL, otherwise a NamedMapPop-class

References

Giovanny Covarrubias-Pazaran (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
haplo <- Matrix::Matrix(0, nrow=4, ncol=5)
for (i in 1:4) {
  haplo[i,] <- ifelse(runif(5)<0.2,0,1)
}
colnames(haplo) = letters[1:5]

genMap = data.frame(markerName=letters[1:5],
                    chromosome=c(1,1,1,2,2),
                    position=c(0,0.5,1,0.15,0.4))

ped = data.frame(id=c("a","b"),
                 mother=c(0,0),
                 father=c(0,0))

founderPop = importHaploSparse(haplo=haplo,
                              genMap=genMap,
                              ploidy=2L,
                              ped=ped)
```

inbFun	<i>Fitness function from contribution theory using only the group relationship</i>
--------	--

Description

Simple function for fitness where we only use the group relationship.

Usage

```
inbFun(Y,b,Q,D,a, lambda)
```

Arguments

Y	A matrix of trait values. See details.
b	A vector of trait weights. See details.
Q	A QTL matrix. See details.
D	An LD matrix. See details.
a	A named list with vectors of average allelic effects per trait. See details.
lambda	A numeric value to weight the Q'DQ portion of the objective function (to be provided by the user with the lambda argument). See details.

Details

A simple apply function of a regular index weighted by a vector of relationships.

`Matrix::diag(Q%*%Matrix::tcrossprod(D,Q))` of dimensions $n \times n$

Notice that Q represents the marker of QTLs (columns) for all solutions (rows) and D the LD between QTLs. The user can modify this function as needed and provide it to the `evolafit` function along with other arguments.

Value

\$res a vector of values

References

Giovanny Covarrubias-Pazaran (2024). `evola`: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
Q <- matrix(1,3,3) # QTL matrix available internally
D <- diag(3) # LD matrix
inbFun(Q=Q, D=D) # group relationship
```

Jc	<i>Matrix of ones</i>
----	-----------------------

Description

Makes a matrix of ones with a single row and nc columns.

Usage

```
Jc(nc)
```

Arguments

nc Number of columns to create.

Details

A simple apply function to make a matrix of one row and nc columns.

Value

\$res a matrix

References

Giovanny Covarrubias-Pazaran (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
Jc(5)
```

Jr	<i>Matrix of ones</i>
----	-----------------------

Description

Makes a matrix of ones with a single column and nr rows.

Usage

```
Jr(nr)
```

Arguments

nr Number of rows to create.

Details

A simple apply function to make a matrix of one column and nr rows.

Value

\$res a matrix

References

Giovanny Covarrubias-Pazaran (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
Jr(5)
```

logspace	<i>Decreasing exponential trend</i>
----------	-------------------------------------

Description

logspace creates a vector with decreasing logarithmic trend.

Usage

```
logspace(x, p=2)
```

Arguments

x	sequence of values to pass through the function.
p	power to be applied to the values.

Value

\$res a vector of length n with exponential decrease trend.

Author(s)

Giovanny Covarrubias-Pazaran

References

Covarrubias-Pazaran G (2016) Genome assisted prediction of quantitative traits using the R package sommer. PLoS ONE 11(6): doi:10.1371/journal.pone.0156744

Examples

```
plot(logspace(1:100,p=1))
plot(logspace(1:100,p=2))
plot(logspace(1:100,p=3))
```

nQtl	<i>Matrix of number of activated QTLs</i>
------	---

Description

Makes a matrix indicating how many QTLs were activated for each solution.

Usage

```
nQtl(object)
```


Arguments

object Object returned by the evolafit function.

Details

A simple apply function to count the number of active QTLs per solution (row) per trait (columns).

Value

\$res a matrix

References

Giovanny Covarrubias-Pazarán (2024). evolafit: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
set.seed(1)

# Data
Gems <- data.frame(
  Color = c("Red", "Blue", "Purple", "Orange",
            "Green", "Pink", "White", "Black",
            "Yellow"),
  Weight = round(runif(9,0.5,5),2),
  Value = round(abs(rnorm(9,0,5))+0.5,2),
  Times=c(rep(1,8),0)
)
head(Gems)

# Task: Gem selection.
# Aim: Get highest combined value.
# Restriction: Max weight of the gem combined = 10.

# simple specification
res00<-evolafit(formula=cbind(Weight,Value)~Color, dt= Gems,
                # constraints on traits: if greater than this ignore
                constraintsUB = c(10,Inf), nGenerations = 10
)
nQtl(res00)
```

ocsFun

*Fitness function from contribution theory***Description**

Simple function for fitness where an index of traits is weighted by the group relationship.

Usage

```
ocsFun(Y,b,Q,D,a,lambda,scaled=TRUE)
```

Arguments

Y	A matrix of trait values. See details.
b	A vector of trait weights. See details.
Q	A QTL matrix. See details.
D	An LD matrix. See details.
a	A named list with vectors of average allelic effects per trait. See details.
lambda	A numeric value to weight the Q'DQ portion of the objective function (to be provided by the user with the lambda argument). See details.
scaled	A logical value to indicate if traits should be scaled prior to multiply by the weights.

Details

A simple apply function of a regular index weighted by a vector of relationships.

$$Y \% \% b - d$$

Internally, we use this function in the following way:

The Y matrix is the matrix of trait-GBVs and b is the user-specified trait weights.

$d = qtDq * \lambda$; where $qtDq$ is equal to $\text{Matrix::diag}(Q \% \% \text{Matrix::tcrossprod}(D,Q))$ of dimensions $n \times n$

Notice that Q represents the marker of QTLs (columns) for all solutions (rows) and D the LD between QTLs. The user can modify this function as needed and provide it to the evolafit function along with other arguments.

Notice that a is a list with elements named as the traits specified in your formula.

Value

\$res a vector of values

References

Giovanny Covarrubias-Pazarán (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
Y=matrix(1:12,4,3) # 4 solutions with 3 traits

Q=matrix(sample(0:1,32,replace = TRUE),nrow=4,ncol=8) # coancestry for each solution

D=diag(8)

b=rep(1,3)

lambda=0.5

ocsFun(Y=Y,Q=Q,D=D,b=b, lambda=lambda) # Yb - d where d is QAQ' and A is the LD between QTNs
```

overlay	<i>Overlay Matrix</i>
---------	-----------------------

Description

‘overlay‘ adds r times the design matrix for model term t to the existing design matrix. Specifically, if the model up to this point has p effects and t has a effects, the a columns of the design matrix for t are multiplied by the scalar r (default value 1.0). This can be used to force a correlation of 1 between two terms as in a diallel analysis.

Usage

```
overlay(..., rlist=NULL, prefix=NULL, sparse=FALSE)
```

Arguments

...	as many vectors as desired to overlay.
rlist	a list of scalar values indicating the times that each incidence matrix overlayed should be multiplied by. By default r=1.
prefix	a character name to be added before the column names of the final overlay matrix. This may be useful if you have entries with names starting with numbers which programs such as asreml doesn't like, or for posterior extraction of parameters, that way 'grep'ing is easier.
sparse	a TRUE/FALSE statement specifying if the matrices should be built as sparse or regular matrices.

Value

\$\$3 an incidence matrix with as many columns levels in the vectors provided to build the incidence matrix.

Author(s)

Giovanny Covarrubias-Pazaran

References

Fikret Isik. 2009. Analysis of Diallel Mating Designs. North Carolina State University, Raleigh, USA.

Covarrubias-Pazaran G (2016) Genome assisted prediction of quantitative traits using the R package soevolafit. PLoS ONE 11(6): doi:10.1371/journal.pone.0156744

See Also

The core functions of the package [evolafit](#).

Examples

```
#####
#### For CRAN time limitations most lines in the
#### examples are silenced with one '#' mark,
#### remove them and run the examples
#####
data("DT_technow")
DT <- DT_technow
head(DT)
DT$dentf <- as.factor(DT$dent)
DT$flintf <- as.factor(DT$flint)

with(DT, overlay(dentf,flintf, sparse = TRUE))
with(DT, overlay(dentf,flintf, sparse = FALSE))
```

pareto

plot the change of values across iterations

Description

plot for monitoring.

Usage

```
pareto(object, scaled=TRUE,pch=20, xlim, ...)
```

Arguments

object	model object returned by "evolafit"
scaled	a logical value to specify the scale of the y-axis (gain in merit).
pch	symbol for plotting points as described in par
xlim	upper and lower bound in the x-axis
...	Further arguments to be passed to the plot function.

Value

vector of plot

Author(s)

Giovanny Covarrubias

See Also

[plot](#), [evolafit](#)

pmonitor

plot the change of values across iterations

Description

plot for monitoring.

Usage

```
pmonitor(object, kind, ...)
```

Arguments

object	model object of class "evolafit"
kind	a numeric value indicating what to plot according to the following values: 1: Average and best q'a (contribution) 2. Average q'Dq and deltaC 3. Number of QTLs activated
...	Further arguments to be passed to the plot function.

Value

trace plot

Author(s)

Giovanny Covarrubias

See Also

[plot](#), [evolafit](#)

regFun

Fitness function from linear regressions based on mean squared error.

Description

Simple function for fitness where the mean squared error is computed when the user provides y and X and b are the average allelic effects of the population in the genetic algorithm.

Usage

```
regFun(Y,b,Q,D,a,lambda,X,y)
```

Arguments

Y	A matrix of trait values (internal matrix). See details.
b	A vector of trait weights (provided by users). See details.
Q	A QTL matrix (internal matrix). See details.
D	An LD matrix. See details.
a	A named list with vectors of average allelic effects per trait (internal matrix). See details.
lambda	A numeric value to weight the Q'DQ portion of the objective function (to be provided by the user with the lambda argument). See details.
X	A matrix of covariates or explanatory variables (to be provided by the user in the ... arguments). See details.
y	A vector of the response variable (to be provided by the user in the ... arguments). See details.

Details

A simple apply function of a regular mean squared error.

$$(y - X \% \% b)^2$$

Internally, we use this function in the following way:

The y vector and X matrix are provided by the user and are fixed values that do not change across iterations. The evolutionary algorithm optimizes the b values which are the QTLs and associated average allelic effects that are evolving. The 'b' coefficients in the formula come from the GA and are computed as:

$$b[j] = a[[1]][p[[j]]]$$

where a[[1]] is the list of QTL average allelic effects per trait provided in the original dataset, whereas p is a list (with length equal to the number of solutions) that indicates which QTLs are activated in each solution and the j variable is just a counter so each solution is tested.

Value

\$res a vector of values

References

Giovanny Covarrubias-Pazaran (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
y <- rnorm(40) # 4 responses
X=matrix(rnorm(120),40,3) # covariates
Q=matrix(0,40,30) # QTL matrix with 30 QTLs
for(i in 1:nrow(Q)){Q[i,sample(1:ncol(Q),3)]=1}
a <- matrix(rnorm(30),ncol=1) # 30 average allelic effects in trait 1

mse = regFun(y=y, X=X, Q=Q, a=a, # used
             # ignored, Y is normally available in the evolafit routine
             Y=X)
```

stan	<i>Standardize a vector of values in range 0 to 1</i>
------	---

Description

Simple function to map a vector of values to the range of 0 and 1 values to have a better behavior of the algorithm.

Usage

```
stan(x, lb=0, ub=1)
```

Arguments

- x A vector of numeric values.
- lb Lower bound value to map the x values.
- ub Upper bound value to map the x values.

Details

Simple function to map a vector of values to the range of 0 and 1 values to have a better behavior of the algorithm.

Value

\$res new values in range 0 to 1

References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
x <- rnorm(20, 10, 3);x
stan(x)
```

update.evolaFitMod	<i>update form an evolafit model</i>
--------------------	--------------------------------------

Description

update method for class "evolaFitMod".

Usage

```
## S3 method for class 'evolaFitMod'
update(object, formula., evaluate = TRUE, ...)
```

Arguments

object	an object of class "evolaFitMod"
formula.	an optional formula
evaluate	a logical value to indicate if an evaluation of the call should be done or not
...	Further arguments to be passed

Value

an updated model

Author(s)

Giovanny Covarrubias

Examples

```
set.seed(1)

# Data
Gems <- data.frame(
  Color = c("Red", "Blue", "Purple", "Orange",
            "Green", "Pink", "White", "Black",
            "Yellow"),
  Weight = round(runif(9,0.5,5),2),
  Value = round(abs(rnorm(9,0,5))+0.5,2),
  Times=c(rep(1,8),0)
)
# Task: Gem selection.
# Aim: Get highest combined value.
# Restriction: Max weight of the gem combined = 10.

# simple specification
res<-evolafit(formula=cbind(Weight,Value)~Color, dt= Gems,
              # constraints on traits: if greater than this ignore
              constraintsUB = c(10,Inf), nGenerations = 2
)
resUp=update(res)
```

varQ

Extract the variance existing in the genome solutions

Description

Extracts the variance found across the M element of the resulting object of the evolafit() function which contains the different solution and somehow represents the genome of the population.

Usage

```
varQ(object)
```

Arguments

object A resulting object from the function evolafit.

Details

A simple apply function looking at the variance in each column of the M element of the resulting object of the evolafit function.

Value

\$res a value of variance

References

Giovanny Covarrubias-Pazaran (2024). *evola*: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

See Also

[evolafit](#) – the core function of the package

Examples

```
set.seed(1)
# Data
Gems <- data.frame(
  Color = c("Red", "Blue", "Purple", "Orange",
            "Green", "Pink", "White", "Black",
            "Yellow"),
  Weight = round(runif(9,0.5,5),2),
  Value = round(abs(rnorm(9,0,5))+0.5,2),
  Times=c(rep(1,8),0)
)
head(Gems)

# Task: Gem selection.
# Aim: Get highest combined value.
# Restriction: Max weight of the gem combined = 10.
res0<-evolafit(cbind(Weight,Value)~Color, dt= Gems,
  # constraints: if greater than this ignore
  constraintsUB = c(10,Inf),
  # constraints: if smaller than this ignore
  constraintsLB= c(-Inf,-Inf),
  # weight the traits for the selection
  b = c(0,1),
  # population parameters
  nCrosses = 100, nProgeny = 20, recombGens = 1,
  # coancestry parameters
  D=NULL, lambda=c(0,0), nQTLperInd = 1,
  # selection parameters
  propSelBetween = .9, propSelWithin =0.9,
  nGenerations = 5
)

varQ(res0)
```

Index

- * **R package**
 - evola-package, [2](#)
- * **classes**
 - evolaPop-class, [18](#)
- * **datasets**
 - DT_cpdata, [7](#)
 - DT_technow, [8](#)
 - DT_wheat, [10](#)
- * **models**
 - pareto, [28](#)
 - pmonitor, [29](#)
 - update.evolaFitMod, [32](#)
- A (DT_cpdata), [7](#)
- A.mat, [3](#)
- A_wheat (DT_wheat), [10](#)
- addZeros, [4](#)
- bestSol, [5](#)
- DT_cpdata, [3](#), [7](#)
- DT_technow, [3](#), [8](#)
- DT_wheat, [3](#), [10](#)
- evola (evola-package), [2](#)
- evola-package, [2](#)
- evolafit, [2](#), [4–6](#), [12](#), [17](#), [18](#), [20–23](#), [25](#),
[27–32](#), [34](#)
- evolaPop (evolaPop-class), [18](#)
- evolaPop-class, [18](#)
- GT_wheat (DT_wheat), [10](#)
- importHaploSparse, [19](#)
- inbFun, [14](#), [21](#)
- Jc, [22](#)
- Jr, [23](#)
- logspace, [24](#)
- M_technow (DT_technow), [8](#)
- nQt1, [24](#)
- ocsFun, [14](#), [26](#)
- overlay, [27](#)
- pareto, [28](#)
- plot, [29](#), [30](#)
- pmonitor, [29](#)
- Pop, [18](#), [19](#)
- regFun, [15](#), [30](#)
- stan, [31](#)
- update.evolaFitMod, [32](#)
- varQ, [33](#)