

Package ‘icensmis’

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Title Study Design and Data Analysis in the Presence of Error-Prone
Diagnostic Tests and Self-Reported Outcomes

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Description We consider studies in which information from error-prone diagnostic tests or self-reports are gathered sequentially to determine the occurrence of a silent event. Using a likelihood-based approach incorporating the proportional hazards assumption, we provide functions to estimate the survival distribution and covariate effects. We also provide functions for power and sample size calculations for this setting. Please refer to Xiangdong Gu, Yunsheng Ma, and Raji Balasubramanian (2015) <[doi:10.1214/15-AOAS810](https://doi.org/10.1214/15-AOAS810)>, Xiangdong Gu and Raji Balasubramanian (2016) <[doi:10.1002/sim.6962](https://doi.org/10.1002/sim.6962)>, Xiangdong Gu, Mahlet G Tadesse, Andrea S Foulkes, Yunsheng Ma, and Raji Balasubramanian (2020) <[doi:10.1186/s12911-020-01223-w](https://doi.org/10.1186/s12911-020-01223-w)>.

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bayesmc	<i>Bayesian method for high-dimensional variable selection</i>
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Description

Bayesian method for high-dimensional variable selection

Usage

bayesmc(Dm, Xmat, b, om1, om2, niter, psample, initsurv, nreport, fitsurv)

Arguments

Dm	the D matrix
Xmat	the design matrix
b	the prior distribution parameter for beta, normal std
om1	the prior distribution parameter for omega
om2	the piror distribution parameter for omega
niter	number of iteration
psample	the sampling probability for updating regresson coefficient
initsurv	initial survival probabilities at end of study
nreport	every how many iterations to output parameters
fitsurv	the survival parameters optimization function

datasim	<i>Simulate data including multiple outcomes from error-prone diagnostic tests or self-reports</i>
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Description

This function simulates a data of N subjects with misclassified outcomes, assuming each subject receives a sequence of pre-scheduled tests for disease status ascertainment. Each test is subject to error, characterized by sensitivity and specificity. An exponential distribution is assumed for the time to event of interest. Three kinds of covariate settings can be generated: one sample setting, two group setting, and continuous covariates setting with each covariate sampled from i.i.d. $N(0, 1)$. Two missing mechanisms can be assumed, namely MCAR and NTFP. The MCAR setting assumes that each test is subject to a constant, independent probability of missingness. The NTFP mechanism includes two types of missingness - (1) incorporates a constant, independent, probability of missing for each test prior to the first positive test result; and (2) all test results after first positive are missing. The simulated data is in longitudinal form with one row per test time.

Covariate values, by default, are assumed to be constant. However, this function can simulate a special case of time varying covariates. Under time varying covariates setting, each subject is assumed to have a change time point, which is sampled from the visit times. We assume that each subject has two sets of covariate values. Before his change time point, the covariate values take from the first set, and second set after change time point. Thus, each subject's distribution of survival time is two-piece exponential distribution with different hazard rates.

Usage

```
datasim(  
  N,  
  blambda,  
  testtimes,  
  sensitivity,  
  specificity,  
  betas = NULL,  
  twogroup = NULL,  
  pmiss = 0,  
  pcensor = 0,  
  design = "MCAR",  
  negpred = 1,  
  time.varying = F  
)
```

Arguments

<code>N</code>	total number of subjects to be simulated
<code>blambda</code>	baseline hazard rate
<code>testtimes</code>	a vector of pre-scheduled test times
<code>sensitivity</code>	the sensitivity of test

specificity	the specificity of test
betas	a vector of regression coefficients of the same length as the covariate vector. If <code>betas = NULL</code> then the simulated dataset corresponds to the one sample setting. If <code>betas != NULL</code> and <code>twogroup != NULL</code> then the simulated dataset corresponds to the two group setting, and the first value of <code>betas</code> is used as the coefficient for the treatment group indicator. If <code>betas != NULL</code> and <code>twogroup = NULL</code> , then the covariates are \sim i.i.d. $N(0, 1)$, and the number of covariates is determined by the length of <code>betas</code> .
twogroup	corresponds to the proportion of subjects allocated to the baseline (reference) group in the two-group setting. For the two-group setting, this variable should be between 0 and 1. For the one sample and multiple (≥ 2) covariate setting, this variable should be set to <code>NULL</code> . That is, when <code>betas != NULL</code> , set <code>twogroup</code> to equal the proportion of the subjects in the baseline group to obtain a simulated dataset corresponding to the two-group setting. Else, set <code>twogroup=NULL</code> to obtain either the one sample setting (<code>betas=NULL</code>) or continuous covariates (<code>betas != NULL</code>).
pmiss	a value or a vector (must have same length as <code>testtimes</code>) of the probabilities of each test being randomly missing at each test time. If <code>pmiss</code> is a single value, then each test is assumed to have an identical probability of missingness.
pcensor	a value or a vector (must have same length as <code>testtimes</code>) of the interval probabilities of censoring time at each interval, assuming censoring process is independent on other missing mechanisms. If it is the single value, then we assume same interval probabilities as the value. The sum of <code>pcensor</code> (or <code>pcensor * length(testtimes)</code> if it is single value) must be ≤ 1 . For example, if <code>pcensor = c(0.1, 0.2)</code> , then it means the the probabilities of censoring time in first and second intervals are 0.1, 0.2, and the probability of not being censored is 0.7.
design	missing mechanism: "MCAR" or "NTP"
negpred	baseline negative predictive value, i.e. the probability of being truly disease free for those who were tested (reported) as disease free at baseline. If baseline screening test is perfect, then <code>negpred = 1</code> .
time.varying	indicator whether fitting a time varying covariate model or not

Details

To simulate the one sample setting data, set `betas` to be `NULL`. To simulate the two group setting data, set `twogroup` to equal the proportion of the subjects in the baseline group and set `betas` to equal the coefficient corresponding to the treatment group indicator(i.e. `beta` equals the log hazard ratio of the two groups). To simulate data with continuous i.i.d. $N(0, 1)$ covariates, set `twogroup` to be `NULL` and set `betas` to equal the vector of coefficients of the covariates.

Value

simulated longitudinal form data frame

Examples

```
## One sample setting
```

```

simdata1 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8, sensitivity = 0.7,
  specificity = 0.98, betas = NULL, twogroup = NULL, pmiss = 0.3, design = "MCAR")

## Two group setting, and the two groups have same sample sizes
simdata2 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8, sensitivity = 0.7,
  specificity = 0.98, betas = 0.7, twogroup = 0.5, pmiss = 0.3, design = "MCAR")

## Three covariates with coefficients 0.5, 0.8, and 1.0
simdata3 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8, sensitivity = 0.7,
  specificity = 0.98, betas = c(0.5, 0.8, 1.0), twogroup = NULL, pmiss = 0.3,
  design = "MCAR", negpred = 1)

## NTFP missing mechanism
simdata4 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8, sensitivity = 0.7,
  specificity = 0.98, betas = c(0.5, 0.8, 1.0), twogroup = NULL, pmiss = 0.3,
  design = "NTFP", negpred = 1)

## Baseline misclassification
simdata5 <- datasim(N = 2000, blambda = 0.05, testtimes = 1:8, sensitivity = 0.7,
  specificity = 0.98, betas = c(0.5, 0.8, 1.0), twogroup = NULL, pmiss = 0.3,
  design = "MCAR", negpred = 0.97)

## Time varying covariates
simdata6 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8, sensitivity = 0.7,
  specificity = 0.98, betas = c(0.5, 0.8, 1.0), twogroup = NULL, pmiss = 0.3,
  design = "MCAR", negpred = 1, time.varying = TRUE)

```

fitsurv

Fit survival function, used for Bayesian simulation

Description

Fit survival function, used for Bayesian simulation

Usage

```
fitsurv(parm, Dm, eta)
```

Arguments

parm	the initial parameter value
Dm	the D matrix
eta	equals to X^*bea

icmis	<i>Maximum likelihood estimation for settings of error-prone diagnostic tests and self-reported outcomes</i>
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Description

This function estimates the baseline survival function evaluated at each test time in the presence of error-prone diagnostic tests and self-reported outcomes. If there are covariates included in the dataset, it also estimates their coefficients assuming proportional hazards. The covariate values can be either time independent or time varying. The function can also be used to incorporate misclassification of disease status at baseline (due to an error-prone diagnostic procedure).

Usage

```
icmis(
  subject,
  testtime,
  result,
  data,
  sensitivity,
  specificity,
  formula = NULL,
  negpred = 1,
  time.varying = F,
  betai = NULL,
  initsurv = 0.5,
  param = 1,
  ...
)
```

Arguments

subject	variable in data for subject id.
testtime	variable in data for test time. Assume all test times are non-negative. testtime = 0 refers to baseline visit (only used/needed if the model is time varying covariates)
result	variable in data for test result.
data	the data to analyze.
sensitivity	the sensitivity of test.
specificity	the specificity of test.
formula	a formula to specify what covariates to be included in the model. If there is no covariate or one sample setting, set it to NULL. Otherwise, input like ~x1 + x2 + factor(x3).
negpred	baseline negative predictive value, i.e. the probability of being truly disease free for those who were tested (reported) as disease free at baseline. If baseline screening test is perfect, then negpred = 1.

<code>time.varying</code>	indicator whether fitting a time varying covariate model or not.
<code>betai</code>	a vector of initial values for the regression coefficients corresponding to the vector of covariates. If <code>betai=NULL</code> , then 0s are used for the initial values. Otherwise, the length of <code>betai</code> must equal the number of covariates.
<code>initsurv</code>	initial value for survival function of baseline group in the last visit time. It is used to compute initial values for survival function at all visit times.
<code>param</code>	parameterization for survival function used for optimization, taking values 1, 2, or 3. There are 3 parameterizations available. <code>param = 1</code> : this parameterization uses the change in cumulative incidence in time period j for baseline group as parameters, i.e. $\log(S[j]) - \log(S[j+1])$. <code>param = 2</code> : simply use log of the parameters in <code>param = 1</code> so that those parameters are unbounded. <code>param = 3</code> : the first element is $\log(-\log(S[\text{tau}_1]))$ corresponding to log-log transformation of survival function at first visit, while other parameters are corresponding to the change in log-log of survival function, $\log(-\log(S[j])) - \log(-\log(S[j-1]))$. In most cases, all parameters yield same results, while in some situations especially when two visit times are estimated to have same survival functions, they may differ. Choose the one that works best (check likelihood function)
<code>...</code>	other arguments passed to <code>optim</code> function. For example, if the optimization does not converge, we can increase <code>maxit</code> in the <code>optim</code> function's control argument.

Details

The input data should be in longitudinal form with one row per test time. Use `datasim` to simulate a dataset to see the sample data structure. If time varying model is to be fitted, the baseline visit must be provided so that the baseline covariate information can be extracted. If an error is generated due to the optimization procedure, then we recommend trying different initial values.

This likelihood-based approach is a function of the survival function evaluated at each unique test time in the dataset and the vector of regression coefficients as model parameters. Therefore, it works best for situations where there is a limited number of unique test times in the dataset. If there are a large number of unique test times, one solution is to group several test times together.

Value

A list of fitting results is returned with log-likelihood, estimated coefficients, estimated survival function, and estimated covariance matrix for covariates.

Examples

```
## One sample setting
simdata1 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8,
  sensitivity = 0.7, specificity = 0.98, betas = NULL,
  twogroup = NULL, pmiss = 0.3, design = "MCAR")
fit1 <- icmis(subject = ID, testtime = testtime, result = result,
  data = simdata1, sensitivity = 0.7, specificity = 0.98,
  formula = NULL, negpred = 1)

## Two group setting, and the two groups have same sample sizes
simdata2 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8,
```

```

sensitivity = 0.7, specificity = 0.98, betas = 0.7,
twogroup = 0.5, pmiss = 0.3, design = "MCAR")
fit2 <- icmis(subject = ID, testtime = testtime, result = result,
data = simdata2, sensitivity = 0.7, specificity= 0.98,
formula = ~group)

## Three covariates with coefficients 0.5, 0.8, and 1.0
simdata3 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8,
sensitivity = 0.7, specificity = 0.98, betas = c(0.5, 0.8, 1.0),
twogroup = NULL, pmiss = 0.3, design = "MCAR", negpred = 1)
fit3 <- icmis(subject = ID, testtime = testtime, result = result,
data = simdata3, sensitivity = 0.7, specificity= 0.98,
formula = ~cov1+cov2+cov3, negpred = 1)

## Fit data with NTFP missing mechanism (the fitting is same as MCAR data)
simdata4 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8,
sensitivity = 0.7, specificity = 0.98, betas = c(0.5, 0.8, 1.0),
twogroup = NULL, pmiss = 0.3, design = "NTFP", negpred = 1)
fit4 <- icmis(subject = ID, testtime = testtime, result = result,
data = simdata4, sensitivity = 0.7, specificity= 0.98,
formula = ~cov1+cov2+cov3, negpred = 1)

## Fit data with baseline misclassification
simdata5 <- datasim(N = 2000, blambda = 0.05, testtimes = 1:8,
sensitivity = 0.7, specificity = 0.98, betas = c(0.5, 0.8, 1.0),
twogroup = NULL, pmiss = 0.3, design = "MCAR", negpred = 0.97)
fit5 <- icmis(subject = ID, testtime = testtime, result = result,
data = simdata5, sensitivity = 0.7, specificity= 0.98,
formula = ~cov1+cov2+cov3, negpred = 0.97)

## Fit data with time varying covariates
simdata6 <- datasim(N = 1000, blambda = 0.05, testtimes = 1:8,
sensitivity = 0.7, specificity = 0.98, betas = c(0.5, 0.8, 1.0),
twogroup = NULL, pmiss = 0.3, design = "MCAR", negpred = 1,
time.varying = TRUE)
fit6 <- icmis(subject = ID, testtime = testtime, result = result,
data = simdata6, sensitivity = 0.7, specificity= 0.98,
formula = ~cov1+cov2+cov3, negpred = 1, time.varying = TRUE)

```

icpower

Study design in the presence of error-prone diagnostic tests and self-reported outcomes

Description

This function calculates the power and sample in the presence of error-prone diagnostic tests and self-reported outcomes. Two missing mechanisms can be assumed, namely MCAR and NTFP. The MCAR setting assumes that each test is subject to a constant, independent probability of missingness. The NTFP mechanism includes two types of missingness - (1) incorporates a constant, independent, probability of missing for each test prior to the first positive test result; and (2) all test results after first positive are missing.

Usage

```
icpower(
  HR,
  sensitivity,
  specificity,
  survivals,
  N = NULL,
  power = NULL,
  rho = 0.5,
  alpha = 0.05,
  pmiss = 0,
  pcensor = 0,
  design = "MCAR",
  negpred = 1
)
```

Arguments

HR	hazard ratio under the alternative hypothesis.
sensitivity	the sensitivity of test.
specificity	the specificity of test
survivals	a vector of survival function at each test time for baseline(reference) group. Its length determines the number of tests.
N	a vector of sample sizes to calculate corresponding powers. If one needs to calculate sample size, then set to NULL.
power	a vector of powers to calculate corresponding sample sizes. If one needs to calculate power, then set to NULL.
rho	proportion of subjects in baseline(reference) group.
alpha	type I error.
pmiss	a value or a vector (must have same length as survivals) of the probabilities of each test being randomly missing at each test time. If pmiss is a single value, then each test is assumed to have an identical probability of missingness.
pcensor	a value or a vector (must have same length as survivals) of the interval probabilities of censoring time at each interval, assuming censoring process is independent on other missing mechanisms. If it is the single value, then we assume same interval probabilities as the value. The sum of pcensor (or pcensor * length(survivals) if it is single value) must be ≤ 1 . For example, if pcensor = c(0.1, 0.2), then it means the the probabilities of censoring time in first and second intervals are 0.1, 0.2, and the probability of not being censored is 0.7.
design	missing mechanism: "MCAR" or "NTPP".
negpred	baseline negative predictive value, i.e. the probability of being truly disease free for those who were tested (reported) as disease free at baseline. If baseline screening test is perfect, then negpred = 1.

Details

To calculate sample sizes for a vector of powers, set `N = NULL`. To calculate powers for a vector of sample sizes, set `power = NULL`. One and only one of `power` and `N` should be specified, and the other set to `NULL`. This function uses an enumeration algorithm to calculate the expected Fisher information matrix. The expected Fisher information matrix is used to obtain the variance of the coefficient corresponding to the treatment group indicator.

Value

- `result`: a data frame with calculated sample size and power
- `I1` and `I2`: calculated unit Fisher information matrices for each group, which can be used to calculate more values of sample size and power for the same design without the need to enumerate again

Note

When diagnostic test is perfect, i.e. `sensitivity=1` and `specificity=1`, use [icpowerpf](#) instead to obtain significantly improved computational efficiency.

See Also

[icpowerpf](#)

Examples

```
## First specificity survivals. Assume test times are 1:8, with survival function
## at the end time 0.9
surv <- exp(log(0.9)*(1:8)/8)

## Obtain power vs. N
pow1 <- icpower(HR = 2, sensitivity = 0.55, specificity = 0.99, survivals = surv,
  N = seq(500, 1500, 50), power = NULL, rho = 0.5, alpha = 0.05,
  pmiss = 0, design = "MCAR", negpred = 1)

plot(pow1$result$N, pow1$result$power, type="l", xlab="N", ylab="power")

## Calculate sample size, assuming desired power is 0.9
pow2 <- icpower(HR = 2, sensitivity = 0.55, specificity = 0.99, survivals = surv,
  N = NULL, power = 0.9, rho = 0.5, alpha = 0.05, pmiss = 0, design = "MCAR",
  negpred = 1)

## When missing test is present with MCAR
pow3 <- icpower(HR = 2, sensitivity = 0.55, specificity = 0.99, survivals = surv,
  N = NULL, power = 0.9, rho = 0.5, alpha = 0.05, pmiss = 0.4, design = "MCAR",
  negpred = 1)

## When missing test is present with NTFP
pow4 <- icpower(HR = 2, sensitivity = 0.55, specificity = 0.99, survivals = surv,
  N = NULL, power = 0.9, rho = 0.5, alpha = 0.05, pmiss = 0.4, design = "NTFP",
  negpred = 1)
```

```
## When baseline misclassification is present
pow5 <- icpower(HR = 2, sensitivity = 0.55, specificity = 0.99, survivals = surv,
  N = NULL, power = 0.9, rho = 0.5, alpha = 0.05, pmiss = 0, design = "MCAR",
  negpred = 0.98)

## When test is perfect and no missing test
pow6 <- icpower(HR = 2, sensitivity = 1, specificity = 1, survivals = surv,
  N = NULL, power = 0.9, rho = 0.5, alpha = 0.05, pmiss = 0, design = "MCAR",
  negpred = 1)

## Different missing probabilities at each test time
pow7 <- icpower(HR = 2, sensitivity = 0.55, specificity = 0.99, survivals = surv,
  N = NULL, power = 0.9, rho = 0.5, alpha = 0.05, pmiss = seq(0.1, 0.8, 0.1),
  design = "MCAR")
```

icpower.val

Study design in the presence of error-prone diagnostic tests and self-reported outcomes when sensitivity and specificity are unknown and a validation set is used

Description

This function calculates the power and sample size in the presence of error-prone diagnostic tests and self-reported outcomes when both sensitivity and specificity are unknown. In this case, a subject of the subjects receive both gold standard test and error-prone test at each non-missing visit. The remaining subjects receive only error-prone test. Here, for the validation set, NTFP refers to no test after first positive result from the gold standard test. Both sensitivity and specificity are treated as unknown parameters in this setting.

Usage

```
icpower.val(
  HR,
  sensitivity,
  specificity,
  survivals,
  N = NULL,
  power = NULL,
  rhoval,
  rho = 0.5,
  alpha = 0.05,
  pmiss = 0,
  design = "MCAR",
  designval = "MCAR",
  negpred = 1
)
```

Arguments

HR	hazard ratio under the alternative hypothesis.
sensitivity	the sensitivity of test.
specificity	the specificity of test
survivals	a vector of survival function at each test time for baseline(reference) group. Its length determines the number of tests.
N	a vector of sample sizes to calculate corresponding powers. If one needs to calculate sample size, then set to NULL.
power	a vector of powers to calculate corresponding sample sizes. If one needs to calculate power, then set to NULL.
rhoval	proportion of subjects in validation set.
rho	proportion of subjects in baseline(reference) group.
alpha	type I error.
pmiss	a value or a vector (must have same length as survivals) of the probabilities of each test being randomly missing at each test time. If pmiss is a single value, then each test is assumed to have an identical probability of missingness.
design	missing mechanism: "MCAR" or "NTFP".
designval	missing mechanism of validation set: "MCAR" or "NTFP".
negpred	baseline negative predictive value, i.e. the probability of being truly disease free for those who were tested (reported) as disease free at baseline. If baseline screening test is perfect, then negpred = 1.

Value

- result: a data frame with calculated sample size and power
- IR1 and IR2: calculated unit Fisher information matrices for each group in non-validation set
- IV1 and IV2: calculated unit Fisher information matrices for each group in validation set

Examples

```

surv <- exp(log(0.9)*(1:8)/8)
pow <- icpower.val(HR = 2, sensitivity = 0.55, specificity = 0.99,
  survivals = surv, power = 0.9, rhoval=0.05, design= "NTFP", designval = "NTFP")
pow$result

```

icpowerpf	<i>Study design in the presence of interval censored outcomes (assuming perfect diagnostic tests)</i>
-----------	---

Description

This function implements power and sample size calculations for interval censored time-to-event outcomes, when the diagnostic tests are assumed to be perfect (i.e. sensitivity=1 and specificity=1). This is a special case of the more general study design function [icpower](#). However, for the special case of perfect diagnostic tests, this function can be used with significantly improved computational efficiency.

Usage

```
icpowerpf(
  HR,
  survivals,
  N = NULL,
  power = NULL,
  rho = 0.5,
  alpha = 0.05,
  pmiss = 0
)
```

Arguments

HR	hazard ratio under the alternative hypothesis.
survivals	a vector of survival function at each test time for baseline(reference) group. Its length determines the number of tests.
N	a vector of sample sizes to calculate corresponding powers. If one needs to calculate sample size, then set to NULL.
power	a vector of powers to calculate corresponding sample sizes. If one needs to calculate power, then set to NULL.
rho	proportion of subjects in baseline(reference) group.
alpha	type I error.
pmiss	a value or a vector (must have same length as survivals) of the probabilities of each test being randomly missing at each test time. If pmiss is a single value, then each test is assumed to have an identical probability of missingness.

Value

same form as returned value of [icpower](#)

Note

See [icpower](#) for more details in a general situation.

Examples

```
powpf1 <- icpowerpf(HR =2 , survivals = seq(0.9, 0.1, by=-0.1), N = NULL,
  power = 0.9, pmiss = 0)

powpf2 <- icpowerpf(HR =2 , survivals = seq(0.9, 0.1, by=-0.1), N = NULL,
  power = 0.9, pmiss = 0.7)

## Different missing probabilities at each test time
powpf3 <- icpowerpf(HR =2 , survivals = seq(0.9, 0.1, -0.1), N = NULL,
  power = 0.9, pmiss = seq(0.1, .9, 0.1))
```

icpower_weibull	<i>Study design in the presence of error-prone diagnostic tests and self-reported outcomes for Weibull model</i>
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Description

This functions works same way as [icpower](#) function except that it assumes the survival function follows Weibull distribution. The scale parameter is assumed to be same for both treatment and control groups. This can be used estimate power and sample size for interval censored data using Weibull model, which is a special case when both sensitivity and specificity being 1.

Usage

```
icpower_weibull(
  HR,
  sensitivity,
  specificity,
  shape,
  scale,
  times,
  N = NULL,
  power = NULL,
  rho = 0.5,
  alpha = 0.05,
  pmiss = 0,
  pcensor = 0,
  design = "MCAR",
  negpred = 1
)
```

Arguments

HR	hazard ratio under the alternative hypothesis.
sensitivity	the sensitivity of test.
specificity	the specificity of test

shape	shape parameter of the Weibull distribution for reference group
scale	scale parameter of the Weibull distributions. Same for all groups
times	the visit times
N	a vector of sample sizes to calculate corresponding powers. If one needs to calculate sample size, then set to NULL.
power	a vector of powers to calculate corresponding sample sizes. If one needs to calculate power, then set to NULL.
rho	proportion of subjects in baseline(reference) group.
alpha	type I error.
pmiss	a value or a vector (must have same length as survivals) of the probabilities of each test being randomly missing at each test time. If pmiss is a single value, then each test is assumed to have an identical probability of missingness.
pcensor	a value or a vector (must have same length as testtimes) of the probability of censoring at each visit, assuming censoring process is independent on other missing mechanisms.
design	missing mechanism: "MCAR" or "NTPP".
negpred	baseline negative predictive value, i.e. the probability of being truly disease free for those who were tested (reported) as disease free at baseline. If baseline screening test is perfect, then negpred = 1.

Details

To calculate sample sizes for a vector of powers, set `N = NULL`. To calculate powers for a vector of sample sizes, set `power = NULL`. One and only one of `power` and `N` should be specified, and the other set to `NULL`. This function uses an enumeration algorithm to calculate the expected Fisher information matrix. The expected Fisher information matrix is used to obtain the variance of the coefficient corresponding to the treatment group indicator.

Value

- result: a data frame with calculated sample size and power
- I1 and I2: calculated unit Fisher information matrices for each group, which can be used to calculate more values of sample size and power for the same design without the need to enumerate again

Note

When diagnostic test is perfect, i.e. sensitivity=1 and specificity=1, use [icpowerpf](#) instead to obtain significantly improved computational efficiency.

See Also

[icpowerpf](#) [icpower](#)

Examples

```
icpower_weibull(2, 0.75, 0.98, 1, 0.1, 1:8, power = 0.9)$result  
  
# Interval censoring  
icpower_weibull(2, 1, 1, 1, 0.1, 1:8, power = 0.9)$result
```

plot_surv

Plot survival function

Description

This function plots survival function with confidence interval from model output

Usage

```
plot_surv(obj)
```

Arguments

obj model output object

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