# Package 'renz'

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Type Package

Title R-Enzymology

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#### Description

Contains utilities for the analysis of Michaelian kinetic data. Beside the classical linearization methods (Lineweaver-Burk, Eadie-Hofstee, Hanes-Woolf and Eisenthal-Cornish-Bowden), features include the ability to carry out weighted regression analysis that, in most cases, substantially improves the estimation of kinetic parameters (Aledo (2021) <doi:10.1002/bmb.21522>). To avoid data transformation and the potential biases introduced by them, the package also offers functions to directly fitting data to the Michaelis-Menten equation, either using ([S], v) or (time, [S]) data. Utilities to simulate substrate progress-curves (making use of the Lambert W function) are also provided. The package is accompanied of vignettes that aim to orientate the user in the choice of the most suitable method to estimate the kinetic parameter of an Michaelian enzyme.

**License** GPL ( $\geq 2$ )

**Encoding** UTF-8

LazyData true

RoxygenNote 7.2.3

**Depends** R (>= 4.0.0)

Imports graphics, stats, VGAM

Suggests knitr, rmarkdown, testthat

VignetteBuilder knitr

NeedsCompilation no

**Repository** CRAN

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bibi

Kinetic Mechanisms and Parameters for Bi-Bi Reactions

# Description

Discriminates between sequential and ping-pong mechanisms and estimates the kinetic parameters.

#### Usage

bibi(data, unit\_a = "mM", unit\_b = "mM", unit\_v = "ua", vice\_versa = FALSE)

# Arguments

data	either a dataframe or the path to a text file containing the data (see details).
unit_a	concentration unit for substrate A.
unit_b	concentration unit for substrate B.
unit_v	velocity unit.
vice_versa	logical. When FALSE the variable substrate is A. If TRUE, then the variable substrate is B.

#### Details

Either the txt file or the dataframe containing the data must conform to the following format: a table with three columns and as many rows as conditions were assessed. The first and second columns are named 'a' and 'b' and they give the concentrations for substrate A and B, respectively. The third column, named 'rate', provides the assessed rates.

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#### dir.MM

# Value

A list with three elements: (i) a character vector giving the kinetic parameters Vmax, KiA, Km\_A and Km\_B values; (ii) a numeric vector giving the apparent inverse of Vmax for each concentration of substrate B (intercepts of primary representation); and (iii) a numeric vector giving the apparent specificity constant for each concentration of substrate B (slopes from primary representations).

#### Examples

bibi(data = hk)

dir.MM

Non-linear Least-squares Fitting of the MM equation

#### Description

Non-linear least-squares fitting of the Michaelis-Menten equation.

#### Usage

dir.MM(data, unit\_S = 'mM', unit\_v = 'au', plot = TRUE)

#### Arguments

data	a dataframe with two columns. The first column contains the values of the inde- pendent variable (substrate concentration), and the second column contains the initial rates.
unit_S	concentration unit.
unit_v	time unit.
plot	logical. If true, the data and fitted curve are plotted.

# Details

This function invokes nls() to carry out the fitting.

# Value

A list of two elements. The first one is a vector containing the enzyme kinetic parameters. The second one is a dataframe with the original data plus the fitted value of v.

# Examples

dir.MM(ONPG[, c(1,2)])

#### Description

Obtains Km and Vm using the Eisenthal & Cornish-Bowden method.

#### Usage

ecb(data, unit\_S = 'mM', unit\_v = 'au', plot = TRUE)

# Arguments

data	a dataframe where the first column is the independent variable, [S], and the remaining columns (as many as experiment replicates) correspond to the dependent variable, v.
unit_S	concentration unit.
unit_v	time unit.
plot	logical. If TRUE data are plotted.

#### Details

For each experimental replicate the observations (S, v) are plotted as lines in the Km-Vm parameter space, instead of points in observation space. Afterwards, the lines tend to intersect at a common point, whose coordinates provide the kinetic parameters. Nevertheless, since the observations are subject to error, there is no unique intersection point for all the lines. In this case, the method computes all the pair-wise intersections. Then, the median value from each series is taken to be the best estimate of Km and Vm. This procedure is repeated as many times as replicates and finally the mean and sd is returned.

#### Value

Returns a list with the estimated values of Km and Vm.

#### References

Biochem.J.(1974) 139:715-720 (10.1042/bj1390715)

# See Also

lb(), hw(), eh()

#### Examples

ecb(ONPG[, c(1,2)])

#### ecb

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# Description

Obtain Km and Vm using the Eadie-Hofstee transformation.

#### Usage

eh(data, unit\_S = 'mM', unit\_v = 'au', plot = TRUE)

# Arguments

data	a dataframe where the first column is the independent variable, [S], and the remaining columns (as many as experiment replicates) correspond to the dependent variable, v.
unit_S	concentration unit.
unit_v	time unit.
plot	logical. If TRUE the data and fitted line are plotted.

#### Value

A dataframe with the values of the transformed variables is returned. The fitted Km and Vm are given as attributes of this dataframe.

#### See Also

lb(), hw(), ecb()

# Examples

eh(ONPG[, c(1,2)])

fE.progress	Fitted Progress Curve for Enzyme-Catalyzed Reaction
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# Description

Fits the progress curve of an enzyme-catalyzed reaction.

#### Usage

```
fE.progress(data, unit_S = 'mM', unit_t = 'min')
```

#### eh

#### Arguments

data	a dataframe where the first column is the time and the second column is the substrate concentration.
unit_S	concentration unit.
unit_t	time unit.

# Value

Returns a list with two elements. The first one contains the fitted kinetic parameters, the second one is a dataframe giving the fitted substrate concentration time course.

#### References

Biochem Mol Biol Educ.39:117-25 (10.1002/bmb.20479).

#### See Also

sEprogress(), int.MM()

#### Examples

```
data <- sE.progress(So = 10, time = 5, Km = 4, Vm = 50, plot = FALSE)
fE.progress(data[, c(1,3)])</pre>
```

hk	Kinetic data for the phosphorylation of glucose catalyzed by hexoki-
	nase.

# Description

The variable 'a' is the concentration of ATP-Mg2+ in mM. The variable 'b' is the glucose concentration in mM. 'rate' is given in arbitrary units.

# Usage

hk

# Format

A dataframe with 25 rows (conditions assayed) and 3 columns (variables).

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# Description

Obtains Km and Vm using the Hanes-Woolf transformation.

# Usage

hw(data, unit\_S = 'mM', unit\_v = 'au', plot = TRUE)

# Arguments

data	a dataframe where the first column is the independent variable, [S], and the remaining columns (as many as experiment replicates) correspond to the dependent variable, v.
unit_S	concentration unit.
unit_v	time unit.
plot	logical. If TRUE the data and fitted line are plotted.

# Value

A dataframe with the values of the transformed variables is returned. The fitted Km and Vm are given as attributes of this dataframe.

#### See Also

lb(), eh(), ecb()

# Examples

hw(ONPG[, c(1,2)])

int.MM

Linearization of The Integrated Michaelis-Menten Equation

# Description

Estimates the kinetic parameters using an linearized form of the integrated Michaelis-Menten equation.

# Usage

```
int.MM(data, unit_S = 'mM', unit_t = 'min')
```

hw

# Arguments

trations.	data	a dataframe with two columns. The first column contains the values of the inde- pendent variable time, t, and the second column contains the substrate concen- trations.
unit_S concentration unit.	unit_S	concentration unit.
unit_t time unit.	unit_t	time unit.

#### Details

The r-squared value of the model can be checked using attributes().

# Value

A list of two elements. The first element is named vector containing the Km and Vm. The second element is a dataframe where the first two columns are the original data and the last two columns are the transformed variables. Also a linear plot of the transformed variables together with the parameters values are provided.

#### Examples

int.MM(data = sE.progress(So = 10, time = 5, Km = 4, Vm = 50)[, c(1,3)])

lb

Lineweaver-Burk Transformation

# Description

Obtains Km and Vm using double reciprocal transformation

# Usage

```
lb(data, unit_S = 'mM', unit_v = 'au', weighting = FALSE, plot = TRUE)
```

#### Arguments

data	a dataframe where the first column is the independent variable, [S], and the remaining columns (as many as experiment replicates) correspond to the dependent variable, v.
unit_S	concentration unit.
unit_v	time unit.
weighting	logical. When TRUE the weight v <sup>4</sup> is employed.
plot	logical. If TRUE the data and fitted line are plotted.

#### **ONPG**

#### Value

A double reciprocal plot and the Km and Vm computed using averaged 1/v (when more than one replicate is provided). In addition, this function returns a list of five elements. The first and second ones are vectors with the Km and Vm, respectively, computed individually for each replicate. The third one provides the R-squared values of the fits. The fourth element of the list gives the fitted Km and Vm. The last element of the list is a dataframe with the values of the transformed variables.

#### References

J. Am. Chem. Soc.1934, 56, 3,658-666 (doi.org/10.1021/ja01318a036)

#### See Also

hw(), eh(), ecb()

#### Examples

lb(ONPG[, c(1,2)], weighting = TRUE)

ONPG

Kinetic data for the hydrolysis of ONPG catalyzed by Betagalactosidase (EC. 3.2.1.23)

#### Description

In the University of Málaga, Enzymology is a second-year subject that all Biochemistry students must take. In the context of this subject, students carry out different experiments in the laboratory, using Beta-galactosidase (EC. 3.2.1.23) as an enzyme model, to illustrate the effect of different variables on the rate of the enzyme-catalyzed reaction (hydrolysis of o-nitrophenyl-Beta-d-galactopyranoside, ONPG). One of these experiments consists in assessing the effect of the substrate (ONPG) concentration on the initial rate. The current dataframe shows the results obtained by eight different student groups, as were presented in their reports.

# Usage

ONPG

#### Format

A dataframe with 10 rows (one per substrate concentration) and 9 columns. The first column give the ONPG concentrations assayed (in mM). The remaining columns provide the determined initial rates. Please, note that rates are given using different units, which can be checked typing in the console: attributes(ONPG).

sE.progress

#### Description

Simulates the evolution of the substrate concentration along time.

# Usage

# Arguments

So	initial substrate concentration.
time	reaction timespan.
Km	Michaelis constant.
Vm	maximal velocity.
unit_S	concentration unit.
unit_t	time unit.
I	inhibitor concentration.
Kic	competitive inhibition constant.
Kiu	uncompetitive inhibition constant.
replicates	number of replicates for the dependent variable
error	it should be one among c('absolute', 'relative').
sd	standard deviation of the error.
plot	logical. If TRUE, the progress curve is plotted.

#### Details

When sd is different to 0, then an absolute error normally distributed is added to the variable St.

#### Value

Returns a dataframe where the two first columns are time and St (without error). The two last columns are the mean and sd of the variable St.

#### See Also

fE.progress()

# Examples

sE.progress(So = 10, time = 5, Km = 4, Vm = 50, plot = FALSE)

# TcTS

# Description

The variable 'a' is the concentration of p-nitrophenyl alpha-sialoside (as donor) in mM. The variable 'b' is the lactose (as acceptor) concentration in mM. 'rate' is given in mM/min. Biochemistry 2008, 47, 3507–3512 (https://pubmed.ncbi.nlm.nih.gov/18284211)

# Usage

TcTS

# Format

A dataframe with 16 rows (conditions assayed) and 3 columns (variables).

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