

Package ‘tcpl’

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Title ToxCast Data Analysis Pipeline

Version 3.3.0

Description The ToxCast Data Analysis Pipeline ('tcpl') is an R package that manages, curve-fits, plots, and stores ToxCast data to populate its linked MySQL database, 'invitrodb'. The package was developed for the chemical screening data curated by the US EPA's Toxicity Forecaster (ToxCast) program, but 'tcpl' can be used to support diverse chemical screening efforts.

URL <https://github.com/USEPA/CompTox-ToxCast-tcpl>,
<https://www.epa.gov/comptox-tools/toxicity-forecasting-toxcast>

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tidyr, plotly, tcplfit2, ggplot2, gridExtra, stringr, rlang,
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Author Dayne L Filer [aut],
Jason Brown [ctb] (ORCID: <<https://orcid.org/0009-0000-2294-641X>>),
Madison Feshuk [cre] (ORCID: <<https://orcid.org/0000-0002-1390-6405>>),
Carter Thunes [ctb],
Sarah E Davidson-Fritz [ctb] (ORCID:
<<https://orcid.org/0000-0002-2891-9380>>),

Kelly Carstens [ctb] (ORCID: <<https://orcid.org/0000-0002-1746-5379>>),
 Elizabeth Gilson [ctb],
 Lindsay Knupp [ctb],
 Lori Kolaczowski [ctb],
 Ashley Ko [ctb],
 Zhihui Zhao [ctb],
 Kurt Dunham [ctb],
 Todd Zurlinden [ctb] (ORCID: <<https://orcid.org/0000-0003-1372-3913>>),
 Parth Kothiya [ctb],
 Woodrow R Setzer [ctb],
 Matthew T Martin [ctb, ths],
 Richard S Judson [ctb, ths] (ORCID:
 <<https://orcid.org/0000-0002-2348-9633>>),
 Katie Paul Friedman [ctb] (ORCID:
 <<https://orcid.org/0000-0002-2710-1691>>)

Maintainer Madison Feshuk <feshuk.madison@epa.gov>

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Contents

.buildAssayQ	4
.ChemListQ	5
.ChemQ	5
.convertNames	6
.load6DR	6
.plateHeat	7
.prepField	7
blinShift	8
check_tcpl_db_schema	8
Configure functions	9
dynamic_table_trunc	10
flareFunc	11
get_plot_caption	12
get_plot_title	12
get_verbose_tables	13
Hill model utilites	13
interlaceFunc	14
invitrodb_dd	15
is.odd	16
Load assay information	16
load_data_columns	17
lu	18
lw	19
mc1	20
mc2	20
MC2_Methods	21

mc3	23
MC3_Methods	24
mc4	27
MC4_Methods	28
mc5	29
MC5_Methods	30
mc6	33
MC6_Methods	34
mcdat	35
mc_test	36
mc_vignette	39
Method functions	47
Models	48
MORELETTERS	50
mthd_list_defaults	51
Query functions	52
Register/update annotation	53
registerMthd	54
round_n	55
sc1	55
SC1_Methods	56
sc2	58
SC2_Methods	59
scdat	61
sc_test	62
sc_vignette	64
sink.reset	65
tcplAddModel	66
tcplAICProb	67
tcplAppend	68
tcplCascade	69
tcplCode2CASN	69
tcplCytoPt	70
tcpldbStats	72
tcplDefine	73
tcplDelete	74
tcplFit	74
tcplFit2	76
tcplfit2_fun	76
tcplFit2_nest	77
tcplFit2_unnest	77
tcplGetAeid	78
tcplggplot2	78
tcplHit2	79
tcplLegacyPlot	80
tcplListFlds	80
tcplLoadChem	81
tcplLoadChemList	82

tcplLoadConcUnit	83
tcplLoadData	83
tcplLoadUnit	85
tcplLvlCount	86
tcplMakeAeidMultiPlts	87
tcplMakeAeidPlts	88
tcplMakeChidMultiPlts	89
tcplMultiplot	90
tcplPlot	90
tcplPlotCalcAspectRatio	95
tcplPlotFitc	96
tcplPlotFits	96
tcplPlotLoadData	98
tcplPlotLoadWlIt	99
tcplPlotlyPlot	100
tcplPlotM4ID	100
tcplPlotPlate	101
tcplPlotSetYRange	102
tcplPlotValidate	103
tcplPrepOtp	104
tcplRun	105
tcplSubsetChid	106
tcplVarMat	107
tcplWriteData	109
tcplWriteLvl0	110
test_api	111
write_lvl_4	112

Index	113
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.buildAssayQ	<i>Generate query for assay information</i>
--------------	---

Description

.buildAssayQ generates a query string to load assay information

Usage

```
.buildAssayQ(out, tblo, fld = NULL, val = NULL, add.fld = NULL)
```

Arguments

out	Character, the default fields to include
tblo	Integer, the order to send the fields to prepOutput
fld	Character, the field(s) to query/subset on

val	List, vectors of values for each field to query/subset on. Must be in the same order as 'fld'.
add.fld	Character, additional field(s) to include, but not query/ subset on

Value

A character containing the query to send to tcplQuery

.ChemListQ	<i>Function to support queries by chemical</i>
------------	--

Description

.ChemListQ creates tcplLoadChemList query string

Usage

.ChemListQ(field, val)

Arguments

field	Character, the field to query on
val	Vector of values to subset on

Note

This function is not exported and not intended to be used by the user.

See Also

[tcplLoadChemList](#)

.ChemQ	<i>Function to support queries by chemical</i>
--------	--

Description

.ChemQ creates tcplLoadChem query string

Usage

.ChemQ(field, val, exact)

Arguments

field	Character, the field to query on
val	Vector of values to subset on
exact	Logical, should chemical names be considered exact?

Note

This function is not exported and not intended to be used by the user.

See Also

[tcplLoadData](#)
[tcplLoadChem](#)

.convertNames	<i>Convert assay names to their abbreviations</i>
---------------	---

Description

.convertNames converts the assay names as they appear in the tcpl database to their respective abbreviations

Usage

```
.convertNames(names)
```

Arguments

names	Character, strings to convert
-------	-------------------------------

Value

The same character vector given with any name strings converted to the abbreviated version

.load6DR	<i>Load data for tcpl6</i>
----------	----------------------------

Description

.load6DR loads dose-response data for tcpl6.

Usage

```
.load6DR(ae)
```

Arguments

ae	String acid to query on
----	-------------------------

<code>.plateHeat</code>	<i>Plot plate heatmap</i>
-------------------------	---------------------------

Description

Plot plate heatmap, to be used with `tcplPlotPlate`

Usage

```
.plateHeat(vals, rowi, coli, wllt, wllq, rown, coln, main, arng)
```

Arguments

<code>vals</code>	Numeric, the well values
<code>rowi</code>	Integer, the row index
<code>coli</code>	Integer, the column index
<code>wllt</code>	Character, the well type
<code>wllq</code>	Logical, the well quality
<code>rown</code>	Integer, the number of rows on the plate
<code>coln</code>	Integer, the number of columns on the plate
<code>main</code>	Character of length 1, the title/main
<code>arng</code>	Numeric of length 2, the minimum and maximum values to constrain the color scale

Note

Optimized for an output with height = 20/3, width = 10, and pointsize = 10

<code>.prepField</code>	<i>Paste appropriate table name to field name</i>
-------------------------	---

Description

Paste appropriate table name to field name

Usage

```
.prepField(fld, tbl, db)
```

Arguments

<code>fld</code>	Character, the table fields
<code>tbl</code>	Character, the possible tables
<code>db</code>	Character, the database containing the tables

Details

The function loops through the given tables, and for each field *i* it assigns the last table containing *i* to *i*. ORDER OF FLD MATTERS!!

blineShift	<i>Shift the baseline to 0</i>
------------	--------------------------------

Description

blineShift Takes in dose-response data and shifts the baseline to 0 based on the window.

Usage

```
blineShift(resp, conc, wndw)
```

Arguments

resp	Numeric, the response values
conc	Numeric, the concentration values
wndw	Numeric, the threshold window

Value

A numeric vector containing the shifted response values

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc3_mthds](#), [mc3](#)

check_tcpl_db_schema	<i>Function that checks if the most recent v3 table schema is used in the database schema</i>
----------------------	---

Description

Function that checks if the most recent v3 table schema is used in the database schema

Usage

```
check_tcpl_db_schema()
```


Value

boolean TRUE if param tables are listed in schema FALSE otherwise

Examples

```
## Not run:
#connect to database first with tcplConf
tcplConf(user=user,
  pass= pass,
  db=dbname,
  drvr='MySQL',
  host=hostname)

#check if it is part of the new schema
new_schema <- check_tcpl_db_schema()

## End(Not run)
```

Configure functions *Functions for configuring the tcpl package*

Description

These functions are used to configure the tcpl settings.

Usage

```
tcplConf(drvr = NULL, user = NULL, pass = NULL, host = NULL, db = NULL, ...)

tcplConfDefault()

tcplConfList(show.pass = FALSE)

tcplConfLoad(list.new = TRUE)

tcplConfReset()

tcplConfSave()
```

Arguments

drvr	Character of length 1, which database driver to use
user	Character of length 1, the database server username
pass	Character of length 1, the database server password
host	Character of length 1, the database server
db	Character of length 1, the name of the tcpl database

...	Additional arguments that should be passed to dbConnect function
show.pass	Logical, should the password be returned
list.new	Logical of length 1, should the new settings be printed?

Details

Currently, the tcpl package supports the "MySQL", "example", and "API" database drivers.

The settings can be stored in a configuration file to make the using the package more user-friendly. To create the configuration file, the user must first create a system environment variable ('TCPL_CONF') that points to the file. There is more information about system environment variables in [Startup](#) and [Sys.getenv](#). Briefly, the user needs to modify the '.Renvirom' file in their home directory. If the file does not exist, create it, and add the following line:

TCPL_CONF=path/to/confFile.conf

Here 'path/to/confFile.conf' can be any path to a file. One suggestion would be to include .tcplConf in the home directory, e.g. TCPL_CONF=~/.tcplConf. Note, '~' may not indicate the home directory on every operating system. Once the environment variable is added, the user can change the settings using tcplConf, then save the settings to the file given by the TCPL_CONF environment variable running tcplConfSave().

tcplConf changes options to set the tcpl-specific options, most importantly to configure the connection to the tcpl databases. tcplConf will only change non-null values, and can be used to change a single value if needed.

tcplConfSave modifies the configuration file to reflect the current tcpl settings.

tcplConfList lists the values assigned to the tcpl global options.

tcplConfLoad updates the tcpl settings to reflect the current configuration file.

tcplConfDefault changes the options to reflect the default settings for the API connection, but does not alter the configuration file.

tcplConfReset is used to generate the initial configuration script, and can be used to reset or regenerate the configuration script by the user.

dynamic_table_trunc	<i>dynamic_table_trunc Dynamically truncate lengths of column values of they are longer than a calculated width. Strings contained in verbose table output can be very long, and this function ensures that the 128 character row limit is kept while growing/shrinking default columns widths that may or may not need the space.</i>
---------------------	--

Description

dynamic_table_trunc Dynamically truncate lengths of column values of they are longer than a calculated width. Strings contained in verbose table output can be very long, and this function ensures that the 128 character row limit is kept while growing/shrinking default columns widths that may or may not need the space.

Usage

```
dynamic_table_trunc(tbl = NULL, all_cols)
```

Arguments

tbl	data.table with potential long values to truncate
all_cols	every annotation column to assign lengths

Value

altered data table with truncated strings

flareFunc	<i>Calculate the weighted mean of a square to detect plate flares</i>
-----------	---

Description

flareFunc calculates the weighted mean of square regions to detect plate flares.

Usage

```
flareFunc(val, coli, rowi, apid, r)
```

Arguments

val	Numeric, the well values
coli	Integer, the well column index
rowi	Integer, the well row index
apid	Character, the assay plate id
r	Integer, the number of wells from the center well (in one direction) to make the square

See Also

[MC6_Methods](#), [Method functions](#), [mc6](#)

get_plot_caption	<i>get_plot_caption</i> Generate plot caption given number of flags and length of unique elements
------------------	---

Description

get_plot_caption Generate plot caption given number of flags and length of unique elements

Usage

```
get_plot_caption(dat = NULL)
```

Arguments

dat	data table including all required flags
-----	---

Value

a string caption for the individual plot

get_plot_title	<i>get_plot_title</i> Generate plot title given number of rows and length of unique elements
----------------	--

Description

get_plot_title Generate plot title given number of rows and length of unique elements

Usage

```
get_plot_title(dat = NULL, type = "mc", compare = "m4id", verbose = TRUE)
```

Arguments

dat	data table with all required conc/resp data; each row will extend comparison
type	string of mc or sc indicating if it is single or multi conc
compare	Character vector, the field(s) samples were joined on for comparison
verbose	bool if hitc should be included in the title

Value

a string title for the individual plot

get_verbose_tables	<i>get_verbose_tables Generates 'gt' package tables containing annotations and level 5 metrics</i>
--------------------	--

Description

get_verbose_tables Generates 'gt' package tables containing annotations and level 5 metrics

Usage

```
get_verbose_tables(dat = NULL, type = "mc", compare = "m4id", flags = FALSE)
```

Arguments

dat	data table with all level 5 metrics and mapped annotations
type	string of mc or sc indicating if it is single or multi conc
compare	Character vector, the field(s) samples were joined on for comparison
flags	bool if flags should be included in the table footer

Value

list of 2 GT tables

Hill model utilites	<i>Functions to solve the Hill model</i>
---------------------	--

Description

These functions solve for Hill model parameters.

Usage

```
tcplHillACXX(XX, tp, ga, gw, bt = 0)
tcplHillConc(val, tp, ga, gw, bt = 0)
tcplHillVal(logc, tp, ga, gw, bt = 0)
```

Arguments

XX	Numeric, the activity level (percentage of the top value)
tp	Numeric, the top value from the Hill model
ga	Numeric, the logAC50 value from the Hill model
gw	Numeric, the Hill coefficient from the Hill model
bt	Numeric, the bottom value from the Hill model
val	Numeric, the activity value
logc	Numeric, the log concentration

Details

tcplHillVal computes the value of the Hill model for a given log concentration.

tcplHillACXX computes the activity concentration for a Hill model for a given activity level.

tcplHillConc computes the Hill model concentration for a given value.

Examples

```
## The following code gives examples for a Hill model with a top of 50,
## bottom of 0, AC50 of 1 and Hill coefficient of 1.
## tcplHillVal calculates activity value given a concentration. tcplHillVal
## will return the tp/2 when logc equals ga:
tcplHillVal(logc = 1, tp = 50, ga = 1, gw = 1, bt = 0)

## Here, tcplHillConc returns the concentration where the value equals 20
tcplHillConc(val = 20, tp = 50, ga = 1, gw = 1, bt = 0)

## Note how this differs from tcplHillACXX:
tcplHillACXX(XX = 20, tp = 50, ga = 1, gw = 1, bt = 0)

## tcplHillACXX is based on the top value and allows the user to calculate
## specific activity concentrations based on a percentage of the top value

## For example, we can calculate the value for the concentration 0.25, then
## use that value to check the other two functions.

value <- tcplHillVal(logc = 0.25, tp = 50, ga = 1, gw = 1, bt = 0)
c1 <- tcplHillConc(val = value, tp = 50, ga = 1, gw = 1, bt = 0)
c2 <- tcplHillACXX(XX = value/50*100, tp = 50, ga = 1, gw = 1, bt = 0)
all.equal(0.25, c1, c2)

## Notice, the value had to be transformed to a percentage of the top value
## when using tcplHillACXX
```

interlaceFunc

Calculate the weighted mean of a square to detect interlace effect

Description

interlaceFunc calculates the distance weighted mean of square regions from a 384-well plate that is interlaced onto a 1536 well plate to detect non-random signals coming from the source plate

Usage

```
interlaceFunc(val, intq, coli, rowi, apid, r)
```

Arguments

val	Numeric, the well values
intq	Numeric, interlace quadrant
coli	Integer, the well column index
rowi	Integer, the well row index
apid	Character, the assay plate id
r	Integer, the number of wells from the center well (in one direction) to make the square

See Also

[MC6_Methods](#), [Method functions](#), [mc6](#)

invitrodb_dd	<i>Short descriptions of fields for different tables are stored in a data dictionary.</i>
--------------	---

Description

Short descriptions of fields for different tables are stored in a data dictionary.

Usage

```
invitrodb_dd
```

Format

A data frame with 44 rows and 3 variables:

invitrodb_table Table of the data dictionary

invitrodb_field Field of the data dictionary

description Description

Source

ToxCast database

<code>is.odd</code>	<i>Check for odd numbers</i>
---------------------	------------------------------

Description

`is.odd` takes an integer vector, `x`, and returns TRUE for odd integers.

Usage

```
is.odd(x)
```

Arguments

<code>x</code>	An integer
----------------	------------

Value

TRUE for odd integers and FALSE for even integers.

See Also

Other tcpl abbreviations: [lu\(\)](#), [lw\(\)](#), [sink.reset\(\)](#)

Load assay information

Functions for loading assay information

Description

These functions query the tcpl databases and returns a `data.table` with assay ID and name information. More information about the assay hierarchy is available in the overview vignette.

Usage

```
tcplLoadAcid(fld = NULL, val = NULL, add.fld = NULL)
```

```
tcplLoadAeid(fld = NULL, val = NULL, add.fld = NULL)
```

```
tcplLoadAid(fld = NULL, val = NULL, add.fld = NULL)
```

```
tcplLoadAsid(fld = NULL, val = NULL, add.fld = NULL)
```


Arguments

fld	Character, the field(s) to query/subset on
val	List, vectors of values for each field to query/subset on. Must be in the same order as 'fld'.
add.fld	Character, additional field(s) to include, but not query/ subset on

Details

Each element in the assay hierarchy has its own function, loading the ID and name for the given assay element. For example, tcplLoadAsid will return the assay source ID (asid) and assay source name (asnm).

Value

A data.table containing the ID, name, and any additional fields.

Examples

```
## Not run:
## The load assay functions can be used without any parameters to list the
## full list of registered assay elements:
tcplLoadAsid()
tcplLoadAeid()

## Similarly, the user can add fields without doing any element selection:
tcplLoadAeid(add.fld = c("asid", "aid", "acid"))

## Or, the user can look only at a subset:
tcplLoadAeid(fld = "aeid", val = 1, add.fld = "asid")

## The field can be any value in one of the corresponding assay element
## tables, but the functions also recognize the abbreviated version of
## the name fields.
tcplListFlds("assay")
a1 <- tcplLoadAeid(fld = "anm", val = "Steroidogenesis")
a2 <- tcplLoadAeid(fld = "assay_name", val = "Steroidogenesis")
identical(a1, a2)

## End(Not run)
```

load_data_columns	<i>Lists of column names returned from tcplLoadData invitrodb v4.1 (same as CCTE Bioactivity API version).</i>
-------------------	--

Description

Lists of column names returned from tcplLoadData invitrodb v4.1 (same as CCTE Bioactivity API version).

Usage

load_data_columns

Format

A list with 12 items:

- mc0** Column names returned requesting mc lvl 0 data
- mc1** Column names returned requesting mc lvl 1 data
- mc2** Column names returned requesting mc lvl 2 data
- mc3** Column names returned requesting mc lvl 3 data
- mc4** Column names returned requesting mc lvl 4 data
- mc5** Column names returned requesting mc lvl 5 data
- mc6** Column names returned requesting mc lvl 6 data
- mcagg** Column names returned requesting mc lvl "agg" data
- sc0** Column names returned requesting sc lvl 0 data
- sc1** Column names returned requesting sc lvl 1 data
- sc2** Column names returned requesting sc lvl 2 data
- scagg** Column names returned requesting sc lvl "agg" data

Source

ToxCast database

lu	<i>Abbreviation for</i> length(unique(x))
----	---

Description

lu takes a logical vector, x, and returns length(unique(x)).
lu takes a logical vector, x, and returns length(unique(x)).

Usage

lu(x)

lu(x)

Arguments

x A logical

Value

The unique of the TRUE values in x
The unique of the TRUE values in x

See Also

[unique, which](#)
[unique, which](#)
Other tcpl abbreviations: [is.odd\(\)](#), [lw\(\)](#), [sink.reset\(\)](#)
Other tcpl abbreviations: [is.odd\(\)](#), [lw\(\)](#), [sink.reset\(\)](#)

lw	<i>Abbreviation for</i> <code>length(which(x))</code>
----	---

Description

lw takes a logical vector, x, and returns `length(which(x))`.
lw takes a logical vector, x, and returns `length(which(x))`.

Usage

`lw(x)`
`lw(x)`

Arguments

x A logical

Value

The length of the TRUE values in x
The length of the TRUE values in x

See Also

[length, which](#)
[length, which](#)
Other tcpl abbreviations: [is.odd\(\)](#), [lu\(\)](#), [sink.reset\(\)](#)
Other tcpl abbreviations: [is.odd\(\)](#), [lu\(\)](#), [sink.reset\(\)](#)

mc1

*Perform level 1 multiple-concentration processing***Description**

mc1 loads level 0 data from the tcpl database for the given id and performs level 1 multiple-concentration processing. The processed data is then loaded into the mc1 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
mc1(ac, wr = FALSE)
```

Arguments

ac	Integer of length 1, assay component id (acid) for processing.
wr	Logical, whether the processed data should be written to the tcpl database

Details

Level 1 processing includes defining the concentration and replicate index, cndx and repi, respectively.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

Other multiple-concentration: [mc2\(\)](#), [mc3\(\)](#), [mc4\(\)](#), [mc5\(\)](#), [mc6\(\)](#)

mc2

*Perform level 2 multiple-concentration processing***Description**

mc2 loads level 1 data from the tcpl database for the given id and performs level 2 multiple-concentration processing. The processed data is then loaded into the mc2 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
mc2(ac, wr = FALSE)
```

Arguments

ac Integer of length 1, assay component id (acid) for processing.

wr Logical, whether the processed data should be written to the tcpl database

Details

Level 2 multiple-concentration processing includes defining the corrected value, `cval`, based on the correction methods listed in the `mc2_acid` and `mc2_methods` tables.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a `data.table` containing the processed data

See Also

[Method functions, MC2_Methods](#)

Other multiple-concentration: [mc1\(\)](#), [mc3\(\)](#), [mc4\(\)](#), [mc5\(\)](#), [mc6\(\)](#)

MC2_Methods

List of level 2 multiple-concentration correction functions

Description

`mc2_mthds` returns a list of correction/transformation functions to be used during level 2 multiple-concentration processing.

Usage

```
mc2_mthds()
```

Details

The functions contained in the list returned by `mc2_mthds` return a list of expressions to be executed in the `mc2` (not exported) function environment. The functions are described here for reference purposes, The `mc2_mthds` function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the function/method name.

Value

A list functions

Available Methods

More information about the level 2 multiple-concentration processing is available in the package vignette, "Data_processing."

Correction Methods:

log2 Transform the corrected response value (*cval*) to log-scale (base 2).

log10 Transform the corrected response value (*cval*) to log-scale (base 10).

rmneg Exclude wells with negative corrected response values (*cval*) and downgrading their well quality (*wllq*); if $cval < 0$, $wllq = 0$.

rmzero Exclude wells with corrected response values (*cval*) equal to zero and downgrading their well quality (*wllq*); if $cval = 0$, $wllq = 0$.

mult25 Multiply corrected response value (*cval*) by 25; $25 * cval$.

mult100 Multiply corrected response value (*cval*) by 100; $100 * cval$.

negshift Shift corrected response values (*cval*) by subtracting the minimum *cval* and adding 1, such that the new minimum is 1; $cval - min + 1$.

mult2.5 Multiply corrected response value (*cval*) by 2.5; $2.5 * cval$.

mult3 Multiply corrected response value (*cval*) by 3; $3 * cval$.

mult6 Multiply corrected response value (*cval*) by 6; $6 * cval$.

sub100 Center data around zero by subtracting the corrected response value (*cval*) from 100; $100 - cval$. Typically used if data was pre-normalized around 100 with responses decreasing to 0.

zscore.npwlls Convert the corrected response value (*cval*) to an absolute Z-Score based on the neutral and positive control wells (*wllts* = n and p), by assay plate ID (*apid*)

sub1 Center data around zero by subtracting the corrected response value (*cval*) from 1; $1 - cval$. Typically used if data was pre-normalized around 1 with responses decreasing to 0.

Aggregation Methods:

agg.mean.rep.apid Aggregate technical test replicates (*wllt*=t) by taking the plate-wise mean per sample id (*spid*), assay plate (*apid*), and concentration index (*cndx*).

agg.median.rep.apid Aggregate technical test replicates (*wllt*=t) by taking the plate-wise median per sample id (*spid*), assay plate (*apid*), and concentration index (*cndx*).

agg.percent.rep.spid Use for binary data. Aggregate technical replicates as percentage by taking the sum of *rval* (raw values) relative to total replicates per sample id (*spid*) and concentration index (*cndx*); $cval = (sum(rval) / .N) * 100$.

agg.percent.rep.spid.min1 Use for binary data with variable number of replicates. Aggregate technical replicates as percentage by taking the sum of *rval* (raw values) relative to total replicates per per sample id (*spid*) and concentration index (*cndx*), where there is more than one replicate; $cval = (sum(rval) / .N) * 100$, where $.N > 1$. *Rvals* are collapsed to one value per *cndx*.

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc2, Method functions](#) to query what methods get applied to each acid

`mc3`*Perform level 3 multiple-concentration processing*

Description

`mc3` loads level 2 data from the `tcpl` database for the given id and performs level 3 multiple-concentration processing. The processed data is then loaded into the `mc3` table and all subsequent data is deleted with `tcplCascade`. See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the `tcplRun` wrapper function.

Usage

```
mc3(ac, wr = FALSE)
```

Arguments

<code>ac</code>	Integer of length 1, assay component id (acid) for processing.
<code>wr</code>	Logical, whether the processed data should be written to the <code>tcpl</code> database

Details

Level 3 multiple-concentration processing includes mapping assay component to assay endpoint, duplicating the data when the assay component has multiple assay endpoints, and any normalization of the data. Data normalization based on methods listed in `mc3_aeid` and `mc3_methods` tables.

Value

A boolean of length 1, indicating the success of the processing, or when `'wr'` is `FALSE`, a list where the first element is a boolean indicating the success of processing and the second element is a `data.table` containing the processed data

See Also

[Method functions](#), [MC3_Methods](#)

Other multiple-concentration: [mc1\(\)](#), [mc2\(\)](#), [mc4\(\)](#), [mc5\(\)](#), [mc6\(\)](#)

Description

`mc3_mthds` returns a list of normalization methods to be used during level 3 multiple-concentration processing.

Usage

```
mc3_mthds()
```

Details

The functions contained in the list returned by `mc3_mthds` take `aids` (a numeric vector of aid values) and returns a list of expressions to be executed in the `mc3` (not exported) function environment. The functions are described here for reference purposes. The `mc3_mthds` function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the type of function and the function/method name.

Value

A list of functions

Available Methods

The methods are broken into three types, based on what fields they define. Different methods are used to define "bval" (the baseline value), "pval" (the positive control value), and "resp" (the final response value).

Although it does not say so specifically in each description, all methods are applied by aid.

More information about the level 3 multiple-concentration processing is available in the package vignette, "Data_processing."

bval Methods:

bval.apid.nwlls.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for neutral control wells (wlft = n).

bval.apid.lowconc.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for test compound wells (wlft = t) with a concentration index (cndx) of 1 or 2.

bval.apid.twlls.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) of test compound wells (wlft = t).

bval.apid.tn.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for test compound wells (wlft = t) and neutral control wells (wlft = n).

bval.apid.nwllslowconc.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) of test compound wells (wlIt = t) with a concentration index (cndx) of 1 or 2 or neutral control wells (wlIt = n).

bval.spid.lowconc.med Calculate the baseline value (bval) as the sample-wise median, by sample ID (spid), of the corrected values (cval) of the three lowest concentration test compound wells (wlIt = t and cndx = 1, 2, & 3).

bval.apid.nwllstcwllslowconc.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for neutral control wells (wlIt = n) or wells with a concentration index (cndx) of 1 or 2 and well type of test compound (wlIt = t) or gain-of-signal control in multiple concentrations (wlIt = c).

bval.aeid.nwlls.med Calculate the baseline value (bval) as the endpoint-wise median, by assay component endpoint ID (aeid), corrected value (cval) for neutral control wells (wlIt = n).

pval Methods:

pval.apid.pwlls.med Calculate the positive control value (pval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for single-concentration gain-of-signal positive control wells (wlIt = p).

pval.apid.mwlls.med Calculate the positive control value (pval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for multiple-concentration loss-of-signal negative control wells (wlIt = m).

pval.apid.medpcbyconc.max Calculate the positive control value (pval) as the plate-wise maximum, by assay plate ID (apid), of the medians of the corrected values (cval) for gain-of-signal single- or multiple-concentration negative control wells (wlIt = m or o) by apid, well type, and concentration.

pval.apid.medpcbyconc.min Calculate the positive control value (pval) as the plate-wise minimum, by assay plate ID (apid), of the medians of corrected value (cval) of gain-of-signal single- or multiple-concentration positive control wells (wlIt = p or c) by apid, well type, and concentration.

pval.apid.medncbyconc.min Calculate the positive control value (pval) as the plate-wise minimum, by assay plate ID (apid), of the medians of the corrected values (cval) for gain-of-signal single- or multiple-concentration negative control wells (wlIt = m or o) by apid, well type, and concentration.

pval.apid.pmv.min Calculate the positive control value (pval) as the plate-wise minimum, by assay plate ID (apid), of the medians of the corrected values (cval) for single-concentration gain-of-signal, multiple-concentration loss-of-signal, or viability control wells (wlIt = p, m, or v) by apid, well type, and concentration.

pval.apid.pmv.max Calculate the positive control value (pval) as the plate-wise maximum, by assay plate ID (apid), of the medians of the corrected values (cval) for single-concentration gain-of-signal, multiple-concentration loss-of-signal, or viability control wells (wlIt = p, m, or v) by apid, well type, and concentration.

pval.apid.f.max Calculate the positive control value (pval) as the plate-wise maximum, by assay plate ID (apid), of the medians of important reference wells (wlIt = f) values by apid and concentration.

pval.apid.f.min Calculate the positive control value (pval) as the plate-wise minimum, by assay plate ID (apid), of the medians of important reference wells (wlIt = f) values by apid and concentration.

pval.apid.p.max Calculate the positive control value (pval) as the plate-wise maximum, by assay plate ID (apid), of the medians of the corrected values (cval) for single-concentration gain-of-signal control wells (wlIt = p) by apid.

pval.apid.p.min Calculate the positive control value (pval) as the plate-wise minimum, by assay plate ID (apid), of the medians of the corrected values (cval) for single-concentration gain-of-signal control wells (wlIt = p) by apid.

pval.apid.v.min Calculate the positive control value (pval) as the plate-wise minimum, by assay plate ID (apid), of the medians of the corrected values (cval) for viability control wells (wlIt = v) by apid and concentration.

pval.zero Set the positive control value (pval) to 0; $pval = 0$.

pval.apid.owlls.med Calculate the positive control value (pval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for single-concentration negative control wells (wlIt = o).

pval.2bval Calculate the positive control value (pval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for neutral control wells (wlIt = n) multiplied by 2.

pval.maxp Calculate the positive control value (pval) as the endpoint-wise maximum, by assay component ID (acid), of the corrected values for single-concentration gain-of-signal wells (wlIt = p).

pval.apid.bwlls.med Calculate the positive control value (pval) as the plate-wise median, by assay plate ID (apid), of the corrected values (cval) for blank wells (wlIt = b).

pval.twlls.99pct Calculate positive control value (pval) as the 99th percentile of all corrected value (cvals) of the test compound wells (wlIt = t).

pval.neg.100 Calculate positive control value (pval) as -100 for endpoints in the down direction; $pval = -100$.

resp Methods:

resp.pc Calculate the normalized response (resp) as a percent of control, i.e. the ratio of the difference between the corrected (cval) and baseline (bval) values divided the difference between the positive control (pval) and baseline (bval) values multiplied by 100; $resp = (cval - bval) / (pval - bval) * 100$.

resp.pc.pval.cor Calculate the normalized response (resp) as a percent of control, i.e. the ratio of the difference between the corrected (cval) and baseline (bval) values divided the positive control (pval) value multiplied by 100; $resp = (cval - bval) / pval * 100$.

resp.fc Calculate the normalized response (resp) as the fold change, i.e. the ratio of the corrected (cval) and baseline (bval) values; $resp = cval / bal$.

resp.logfc Calculate the normalized response (resp) as the fold change of logged, i.e. the difference between corrected (cval) and baseline (bval) log-scale values.

resp.log2 Transform the response values to log-scale (base 2).

resp.mult25 Multiply the normalized response value (resp) by 25; $25 * resp$.

resp.scale.mad.log2fc Scale the normalized response value (resp) by the ratio of $\log_2(1.2)$ and 3 multiplied by the baseline median absolute deviation (bmad) of the unscaled normalized response values (resp); $(\log_2 1.2) / 3 * bmad * resp$.

resp.scale.quant.log2fc Scale the normalized response value (resp). First, determine the maximum difference (md) by finding the maximum between the absolute difference of the 1st percentile minus the 50th percentile and the absolute difference of the 99th percentile minus the 50th percentile. Then multiply resp by $\log_2(1.2)$ divided by 20 percent of md; $(\log_2 1.2) / 0.2 * md * resp$.

resp.multneg1 Multiply the normalized response value (resp) by -1; $-1 * resp$.

resp.shiftneg.3bmad Shift all the normalized response values (resp) less than -3 multiplied by the baseline median absolute deviation (bmad) to 0; if $resp < -3 * bmad$, $resp = 0$.

resp.shiftneg.6bmad Shift all the normalized response values (resp) less than -6 multiplied by the baseline median absolute deviation (bmad) to 0; if $resp < -6 * bmad$, $resp = 0$.

resp.shiftneg.10bmad Shift all the normalized response values (resp) less than 10 multiplied by the baseline median absolute deviation (bmad) to 0; if $resp < -10 * bmad$, $resp = 0$.

resp.blinesshift.3bmad.repi Shift the normalized response value (resp) with a baseline correction, by replicate index (repi), with a window of 3 multiplied by the baseline median absolute deviation (bmad).

resp.blinesshift.50.repi Shift the normalized response value (resp) with a baseline correction, by replicate index (repi), with a window of 50.

resp.blinesshift.3bmad.spid Shift the normalized response value (resp) with a baseline correction, by sample ID (spid), with a window of 3 multiplied by the baseline median absolute deviation (bmad).

resp.blinesshift.50.spid Shift the normalized response value (resp) with a baseline correction, by sample ID (spid), with a window of 50.

none Set the corrected response value (cval) as the normalized response value (resp); $cval = resp$. No additional mc3 methods needed for endpoint-specific normalization.

resp.zerocenter.fc Calculate the normalized response (resp) as a zero center fold change, i.e. 1 minus the ratio of corrected (cval) and baseline (bval) values; $resp = 1 - cval/bval$. Typically used for increasing responses.

resp.incr.zerocenter.fc Calculate the normalized response (resp) as a zero center fold change, i.e. the ratio of the the corrected (cval) and baseline (bval) values minus 1; $resp = cval/bval - 1$. Typically used for increasing responses.

resp.mult100 Multiply the normalized response value (resp) by 100; $100 * resp$.

resp.censormed.neg25 Censor (remove) response values from concentrations which median falls below -25.

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc3, Method functions](#) to query what methods get applied to each acid

mc4

Perform level 4 multiple-concentration processing

Description

mc4 loads level 3 data from the tcpl database for the given id and performs level 4 multiple-concentration processing. The processed data is then loaded into the mc4 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
mc4(ae, wr = FALSE)
```

Arguments

ae Integer of length 1, assay endpoint id (aeid) for processing.

wr Logical, whether the processed data should be written to the tcpl database

Details

Level 4 multiple-concentration modeling takes the dose-response data for chemical-assay pairs, and fits three models to the data: constant, hill, and gain-loss. For more information about the models see [Models](#). When a chemical has more than one sample, the function fits each sample separately.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

[tcplFit](#), [Models](#)

Other multiple-concentration: [mc1\(\)](#), [mc2\(\)](#), [mc3\(\)](#), [mc5\(\)](#), [mc6\(\)](#)

MC4_Methods

List of level 4 multiple-concentration methods for calculating bmad

Description

mc4_mthds returns a list of methods to be used during level 4 multiple-concentration processing for calculating bmad

Usage

```
mc4_mthds()
```

Details

The functions contained in the list returned by mc4_mthds take aeids (a numeric vector of aeid values) and returns a list of expressions to be executed in the mc4 (not exported) function environment. The functions are described here for reference purposes, The mc4_mthds function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the type of function and the function/method name.

Value

A list of functions

Available Methods

Although it does not say so specifically in each description, all methods are applied by `aeid`.

More information about the level 4 multiple-concentration processing is available in the package vignette, "Data_processing."

bmad.aeid.lowconc.twells Calculate the baseline median absolute value (`bmad`) as the median absolute deviation of normalized response values (`rep`) for test compound wells (`wllt = t`) with concentration index (`cndx`) equal to 1 or 2. Calculate one standard deviation of the normalized response for test compound wells (`wllt = t`) with a concentration index (`cndx`) of 1 or 2; `onesd = sqrt(sum((resp - mean resp)^2)/sample size - 1)`. `Onesd` is used to establish BMR and therefore required for `tcplfit2` processing.

bmad.aeid.lowconc.nwells Calculate the baseline median absolute value (`bmad`) as the median absolute deviation of normalized response values (`resp`) for neutral control wells (`wllt = n`). Calculate one standard deviation of the normalized response for neutral control wells (`wllt = n`); `onesd = sqrt(sum((resp - mean resp)^2)/sample size - 1)`. `Onesd` is used to establish BMR and therefore required for `tcplfit2` processing.

bidirectional.false Limits bidirectional fitting and processes data in positive analysis direction only. Use for gain-of-signal or inverted data.

bmad5.onesd16.static Replace baseline median absolute deviation (`bmad`) with 5 and one standard deviation (`osd`) of the normalized response for test compound wells (`wllt = t`) with a concentration index (`cndx`) of 1 or 2 with 16. Typically used for binary data where values would otherwise be 0; non-zero values are required for `tcplfit2` processing.

no.unbounded.models Exclude unbounded models and only fit data to bounded models (`hill`, `gnls`, `exp4` and `exp5`).

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc4](#), [Method functions](#) to query what methods get applied to each `aeid`

mc5

Perform level 5 multiple-concentration processing

Description

`mc5` loads level 4 data from the `tcpl` database for the given `id` and performs level 5 multiple-concentration processing. The processed data is then loaded into the `mc5` table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Arguments

ae	Integer of length 1, assay endpoint id (aeid) for processing.
wr	Logical, whether the processed data should be written to the tcpl database

Details

Level 5 multiple-concentration hit-calling uses the fit parameters and the activity cutoff methods from mc5_aeid and mc5_methods to make an activity call and identify the winning model for each fit.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a data.table containing the processed data

See Also

[Method functions](#), [MC5_Methods](#)

Other multiple-concentration: [mc1\(\)](#), [mc2\(\)](#), [mc3\(\)](#), [mc4\(\)](#), [mc6\(\)](#)

MC5_Methods

Load list of level 5 multiple-concentration cutoff methods

Description

mc5_mthds returns a list of additional activity cutoff methods to be used during level 5 multiple-concentration processing.

Usage

```
mc5_mthds(ae)
```

Arguments

ae	Integer of length 1, the assay endpoint id
----	--

Details

The functions contained in the list returned by mc5_mthds take aeids (a numeric vector of aeid values) and returns a list of expressions to be executed in the mc5 (not exported) function environment. The functions are described here for reference purposes, The mc5_mthds function is not exported, nor is it intended for use.

All available methods are described in the "Available Methods" section, listed by the cutoff type in ascending order of cutoff value.

Value

A list of functions

Available Methods

The methods are broken down into five categories based on the type of cutoff they assign. Different methods are used to define cutoffs for "bmad" (baseline median absolute value), "fc" (fold change), "log" (\log_2 or \log_{10}), "pc" (percent of control), and "other" (uncategorized cutoffs).

All methods are applied by acid.

Although there are method exceptions (notably within the "other" category), only highest calculated cutoff value based on assigned methods will be selected for hitcalling. Therefore, only the largest cutoff method per method type should be assigned.

More information about the level 5 multiple-concentration processing is available in the package vignette, "Data_processing."

BMAD Methods:

bmad1 Add a cutoff value of 1 multiplied by baseline median absolute value (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad2 Add a cutoff value of 2 multiplied by the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad3 Add a cutoff value of 3 multiplied by the baseline median absolute deviation (bmad) as defined at Level 4.

bmad4 Add a cutoff value of 4 multiplied the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad5 Add a cutoff value of 5 multiplied the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad6 Add a cutoff value of 6 multiplied by the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad10 Add a cutoff value of 10 multiplied by the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

Fold Change Methods:

fc0.2 Add a cutoff value of 0.2. Typically for zero centered fold change data.

fc0.25 Add a cutoff value of 0.25. Typically for zero centered fold change data.

fc0.3 Add a cutoff value of 0.3. Typically for zero centered fold change data.

fc0.5 Add a cutoff value of 0.5. Typically for zero centered fold change data.

Log Methods: Log Base 2

neglog2_0.88 Add a cutoff value of $-\log_2 0.88$.

log2_1.2 Add a cutoff value of $\log_2 1.2$. Typically for fold change data.

log2_2 Add a cutoff value $\log_2 2$. Typically for fold change data.

Log Base 10

log10_1.2 Add a cutoff value of $\log_{10} 1.2$. Typically for fold change data.

log10_2 Add a cutoff value of $\log_{10} 2$. Typically for fold change data.

Percent of Control Methods:

- pc05** Add a cutoff value of 5. Typically for percent of control data.
- pc10** Add a cutoff value of 10. Typically for percent of control data.
- pc16** Add a cutoff value of 16. Typically for percent of control data.
- pc20** Add a cutoff value of 20. Typically for percent of control data.
- pc25** Add a cutoff value of 25. Typically for percent of control data.
- pc30** Add a cutoff value of 30. Typically for percent of control data.
- pc40** Add a cutoff value of 40. Typically for percent of control data.
- pc50** Add a cutoff value of 50. Typically for percent of control data.
- pc70** Add a cutoff value of 70. Typically for percent of control data.
- pc95** Add a cutoff value of 95. Typically for percent of control data.

Other Methods:

- maxmed20pct** Add a cutoff value of 20 percent of the maximum of all endpoint maximal average response values (max_med).
- coff_2.32** Add a cutoff value of 2.32.
- ow_bidirectional_loss** Multiply winning model hitcall (hitc) by -1 for models fit in the positive analysis direction. Typically used for endpoints where only negative responses are biologically relevant.
- ow_bidirectional_gain** Multiply winning model hitcall (hitc) by -1 for models fit in the negative analysis direction. Typically used for endpoints where only positive responses are biologically relevant.
- osd_coff_bmr** Overwrite the osd value so that bmr equals cutoff.
- ow_loec.coff** Identify the lowest observed effective concentration (loec) where the values of all responses are outside the cutoff band (i.e. $\text{abs}(\text{resp}) > \text{cutoff}$). loec is stored alongside winning model and potency estimates. If loec exists, assume hit call = 1, fitc = 100, model_type = 1, and if not, assume hit call = 0.
- include_loec.coff** Identify the lowest observed effective concentration (loec) where the values of all responses are outside the cutoff band (i.e. $\text{abs}(\text{resp}) > \text{cutoff}$). loec is stored alongside winning model and potency estimates.

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc5, Method functions](#) to query what methods get applied to each acid.

`mc6`*Perform level 6 multiple-concentration processing*

Description

`mc6` loads level 5 data from the `tcpl` database for the given `id` and performs level 6 multiple-concentration processing. The processed data is then loaded into the `mc6` table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
mc6(ae, wr = FALSE)
```

Arguments

<code>ae</code>	Integer of length 1, assay endpoint id (<code>aeid</code>) for processing.
<code>wr</code>	Logical, whether the processed data should be written to the <code>tcpl</code> database

Details

Level 6 multiple-concentration flagging uses both the plate level concentration-response data and the modeled parameters to flag potential false positives and false negative results.

Value

A boolean of length 1, indicating the success of the processing, or when `'wr'` is `FALSE`, a list where the first element is a boolean indicating the success of processing and the second element is a `data.table` containing the processed data

See Also

[Method functions](#), [MC6_Methods](#)

Other multiple-concentration: [mc1\(\)](#), [mc2\(\)](#), [mc3\(\)](#), [mc4\(\)](#), [mc5\(\)](#)

Description

`mc6_mthds` returns a list of flag methods to be used during level 6 multiple-concentration processing.

Usage

```
mc6_mthds()
```

Value

A list functions

Available Methods

More information about the level 6 multiple-concentration processing is available in the package vignette, "Data_processing."

modl.directionality.fail Flag series if model directionality is questionable, i.e. if the winning model direction was opposite, more responses (*resp*) would have exceeded the cutoff (*coff*). If loss was winning directionality ($\text{top} < 0$), flag if $\text{count}(\text{resp} < -1 * \text{coff}) < 2 * \text{count}(\text{resp} > \text{coff})$. If gain was winning directionality ($\text{top} > 0$), flag if $\text{count}(\text{resp} > \text{coff}) < 2 * \text{count}(\text{resp} < -1 * \text{coff})$.

low.nrep Flag series if the average number of replicates per concentration is less than 2; $nrep < 2$.

low.nconc Flag series if 4 concentrations or less were tested; $nconc \leq 4$.

bmd.high Flag series if modeled benchmark dose (BMD) is greater than AC50 (concentration at 50 percent maximal response). This indicates high variability in baseline response in excess of more than half of the maximal response.

singlept.hit.high Flag single-point hit that's only at the highest conc tested, where series is an active hit call ($\text{hitc} \geq 0.9$) with the median response observed above baseline occurring only at the highest tested concentration tested.

singlept.hit.mid Flag single-point hit that's not at the highest conc tested, where series is an active hit call ($\text{hitc} \geq 0.9$) with the median response observed above baseline occurring only at one concentration and not the highest concentration tested.

multipt.neg Flag multi-point miss, where series is an inactive hit call ($\text{hitc} < 0.9$) with multiple median responses observed above baseline.

gnls.lowconc Flag series where winning model is gain-loss (gnls) and the gain AC50 is less than the minimum tested concentration, and the loss AC50 is less than the mean tested concentration.

noise Flag series as noisy if the quality of fit as calculated by the root mean square error (rmse) for the series is greater than the cutoff (*coff*); $\text{rmse} > \text{coff}$.

border Flag series if borderline activity is suspected based on modeled top parameter (*top*) relative to cutoff (*coff*); $0.8 * \text{coff} \leq |\text{top}| \leq 1.2 * \text{coff}$.

overfit.hit Method not yet updated for tcpl implementation. Flag hit-calls that would get changed after doing the small N correction to the aic values.

efficacy.50 Flag low efficacy hits if series has an active hit call ($hitc \geq 0.9$) and efficacy values (e.g. top and maximum median response) less than 50 percent; intended for biochemical assays. If $hitc \geq 0.9$ and $coff \geq 5$, then flag when $top < 50$ or $maxmed < 50$. If $hitc \geq 0.9$ and $coff < 5$, then flag when $top < \log_2 1.5$ or $maxmed < \log_2 1.5$.

ac50.lowconc Flag series with an active hit call ($hitc \geq 0.9$) if AC50 (concentration at 50 percent maximal response) is less than the lowest concentration tested; if $hitc \geq 0.9$ and $ac50 < 10^{\log_{e^{min}}}$, then flag.

viability.gnls Flag series with an active hit call ($hitc \geq 0.9$) if denoted as cell viability assay with winning model is gain-loss (gnls); if $hitc \geq 0.9$, $modl = "gnls"$ and $cell_viability_assay = 1$, then flag.

no.med.gt.3bmad Flag series where no median response values are greater than baseline as defined by 3 times the baseline median absolute deviation (bmad); $nmed_gtbl_pos$ and $nmed_gtbl_neg$ both = 0, where $nmed_gtbl_pos/_neg$ is the number of medians greater than $3 * bmad$ /less than $-3 * bmad$.

Note

This function is not exported and is not intended to be used by the user.

See Also

[mc6](#), [Method functions](#) to query what methods get applied to each aeid.

mcdat

A subset of ToxCast data showing changes in the activity of the intra-cellular estrogen receptor.

Description

The example dataset is used to illustrate how the user can pipeline multiple-concentration data from chemical screening using tcpl.

Usage

```
mcdat
```

Format

A data frame with 14183 rows and 10 variables:

spid sample ID

apid assay plate ID

rowi well-plate row number

coli well-plate column number

wllt well type
wllq well quality
conc concentration in micromolar
rval raw assay component readout value
srcf source file containing the data
acsn assay component source name

Source

ToxCast database

mc_test	<i>List of lists containing queries sent to tcplQuery associated with each test case. Each list also contains the associated ids with each case. Only meant to be used with automated testing with mocking for mc data.</i>
---------	---

Description

List of lists containing queries sent to tcplQuery associated with each test case. Each list also contains the associated ids with each case. Only meant to be used with automated testing with mocking for mc data.

Usage

mc_test

Format

A list with 30 items:

tcplConfQuery Data table with 1 row and 2 columns used for each test case for establishing connection using tcplConf. This data table mocks the response one would get from connecting with invitrodb.

mc0_by_m0id List containing the queries used for loading mc0 data by m0id via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'm0id' labeled item storing the id used to load the data, for use in tests.

mc0_by_acid List containing the queries used for loading mc0 data by acid via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'acid' labeled item storing the id used to load the data, for use in tests.

mc1_by_m1id List containing the queries used for loading mc1 data by m1id via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'm1id' labeled item storing the id used to load the data, for use in tests.

mc1_by_acid List containing the queries used for loading mc1 data by acid via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'acid' labeled item storing the id used to load the data, for use in tests.

- mc2_by_m2id** List containing the queries used for loading mc2 data by m2id via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'm2id' labeled item storing the id used to load the data, for use in tests.
- mc2_by_aeid** List containing the queries used for loading mc2 data by aeid via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- mc3_by_m3id** List containing the queries used for loading mc3 data by m3id via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'm3id' labeled item storing the id used to load the data, for use in tests.
- mc3_by_aeid** List containing the queries used for loading mc3 data by aeid via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- mc4_by_m4id** List containing the queries used for loading mc4 data by m4id via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'm4id' labeled item storing the id used to load the data, for use in tests.
- mc4_by_aeid** List containing the queries used for loading mc4 data by aeid via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- mc5_by_m5id** List containing the queries used for loading mc5 data by m5id via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'm5id' labeled item storing the id used to load the data, for use in tests.
- mc5_by_aeid** List containing the queries used for loading mc5 data by aeid via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- mc6_by_m6id** List containing the queries used for loading mc6 data by m6id via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'm6id' labeled item storing the id used to load the data, for use in tests.
- mc6_by_aeid** List containing the queries used for loading mc6 data by aeid via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- mc7_by_m7id** List containing the queries used for loading mc7 data by m7id via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'm7id' labeled item storing the id used to load the data, for use in tests.
- mc7_by_aeid** List containing the queries used for loading mc7 data by aeid via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- mcagg_by_aeid** List containing the queries used for loading mc 'agg' data by aeid via tcpLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- plot_single_m4id** List containing the queries used for loading and plotting mc data by m4id via tcpPlot. Each query has an associated data table response for mocking an actual connection. Contains one 'm4id' labeled item storing the id used to load the data, for use in tests.
- plot_multiple_m4id** List containing the queries used for loading and plotting mc data by multiple m4ids via tcpPlot. Each query has an associated data table response for mocking an actual connection. Contains one 'm4id' labeled item storing the ids used to load the data, for use in tests.

- plot_single_aeid** List containing the queries used for loading and plotting mc data by aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- plot_multiple_aeid** List containing the queries used for loading and plotting mc data by multiple aeids via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the ids used to load the data, for use in tests.
- plot_single_spid** List containing the queries used for loading and plotting mc data by spid/aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'spid' and 'aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_multiple_spid** List containing the queries used for loading and plotting mc data by multiple spids/aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'spid' and 'aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_single_m4id_compare** List containing the queries used for loading and plotting compared mc data by m4id via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'm4id' and 'compare.m4id' labeled items storing the ids used to load the data, for use in tests.
- plot_multiple_m4id_compare** List containing the queries used for loading and plotting compared mc data by multiple m4ids via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'm4id' and 'compare.m4id' labeled items storing the ids used to load the data, for use in tests.
- plot_single_aeid_compare** List containing the queries used for loading and plotting compared mc data by aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'aeid' and 'compare.aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_multiple_aeid_compare** List containing the queries used for loading and plotting compared mc data by multiple aeids via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'aeid' and 'compare.aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_single_spid_compare** List containing the queries used for loading and plotting compared mc data by spid/aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'spid', 'compare.spid', 'aeid', and 'compare.aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_multiple_spid_compare** List containing the queries used for loading and plotting compared mc data by multiple spids/aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'spid', 'compare.spid', 'aeid', and 'compare.aeid' labeled items storing the ids used to load the data, for use in tests.

Source

ToxCast database

mc_vignette*List with multi-concentration data for the vignette*

Description

This dataset is a list with 6 data.tables (mc0,mc1,mc2,mc3,mc4,mc5).

Usage

mc_vignette

Format

1. **mc0** A data frame with 78 rows and 18 columns containing level 0 formatted raw data.
 - spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - acid** Assay Component ID
 - acnm** Assay Component Name
 - m0id** Level 0 (mc0) ID
 - apid** Assay plate ID
 - rowi** Row Index
 - coli** Column Index
 - wllt** Well Type
 - wllq** Well Quality (0 or 1)
 - conc** Concentration in micromolar
 - rval** Raw assay component readout value
 - srcf** Source file containing the raw data
 - conc_unit** Concentration Units
2. **mc1** A data frame with 78 rows and 21 columns containing level 1 replicate and concentration level indicated data.
 - spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - acid** Assay Component ID
 - acnm** Assay Component Name

- m0id** Level 0 (mc0) ID
 - m1id** Level 1 (mc1) ID
 - apid** Assay plate ID
 - rowi** Row Index
 - coli** Column Index
 - wllt** Well Type
 - wllq** Well Quality (0 or 1)
 - conc** Concentration in micromolar
 - rval** Raw assay component readout value
 - cndx** Concentration index defined by ranking the unique concentrations, with the lowest concentration starting at 1.
 - repi** Temporary replicate ID is defined, the data are scanned from top to bottom and increment the replicate index every time a replicate ID is duplicated
 - srcf** Source file containing the raw data
 - conc_unit** Concentration Units
3. **mc2** A data frame with 78 rows and 20 columns containing level 2 assay component-specific corrections.
- spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - acid** Assay Component ID
 - acnm** Assay Component Name
 - m0id** Level 0 (mc0) ID
 - m1id** Level 1 (mc1) ID
 - m2id** Level 2 (mc2) ID
 - apid** Assay plate ID
 - rowi** Row Index
 - coli** Column Index
 - wllt** Well Type
 - conc** Concentration in micromolar
 - cval** Corrected Value
 - cndx** Concentration index defined by ranking the unique concentrations, with the lowest concentration starting at 1.
 - repi** Temporary replicate ID is defined, the data are scanned from top to bottom and increment the replicate index every time a replicate ID is duplicated
 - conc_unit** Concentration Units
4. **mc3** A data frame with 78 rows and 22 columns containing level 3 assay endpoint normalized data.
- spid** Sample ID

chid Unique chemical ID number for tcpl
casn Chemical Abstract Service(CAS) number
chnm Chemical name
dsstox_substance_id Chemical-specific DTXSID
code CAS number compressed into numeric string
aeid Assay Component Endpoint ID
aenm Assay endpoint name (i.e., assay_component_endpoint_name)
m0id Level 0 (mc0) ID
m1id Level 1 (mc1) ID
m2id Level 2 (mc2) ID
m3id Level 3 (mc3) ID
logc Log base 10 concentration
resp Normalized response value
cndx Concentration index defined by ranking the unique concentrations, with the lowest concentration starting at 1.
wllt Well Type
apid Assay plate ID
rowi Row Index
coli Column Index
repi Temporary replicate ID is defined, the data are scanned from top to bottom and increment the replicate index every time a replicate ID is duplicated
resp_unit Response Units
conc_unit Concentration Units

5. **mc4** A data frame with 5 rows and 149 columns containing level 4 concentration-response fitting data (all fits).

spid Sample ID
chid Unique chemical ID number for tcpl
casn Chemical Abstract Service(CAS) number
chnm Chemical name
dsstox_substance_id Chemical-specific DTXSID
code CAS number compressed into numeric string
aeid Assay Component Endpoint ID
aenm Assay endpoint name (i.e., assay_component_endpoint_name)
m4id Level 4 (mc4) ID
bmad The median absolute deviation of all treatment wells (default option) or blank wells
resp_max Maximum observed response
resp_min Minimum observed response
max_mean Maximum mean response
max_mean_conc Concentration of the maximum mean response
max_med Maximum median response
max_med_conc Concentration of the maximum median response
logc_max Maximum concentration on the log scale

logc_min Minimum concentration on the log scale
nconc The total number of concentration groups
npts Total number of observed responses (i.e. data points in the concentration series)
nrep Number of replicates in concentration groups
nmed_gtbl The number of median responses greater than 3BMAD
cnst_success Success indicator for the Constant model; 1 if the optimization was successful, otherwise 0
cnst_aic Akaike Information Criteria (AIC) for the Constant model
cnst_rme Root mean square error for the Constant model
cnst_er Error term for the Constant model
hill_success Success indicator for the Hill model; 1 if the optimization was successful, otherwise 0
hill_aic Akaike Information Criteria (AIC) for the Hill model
hill_cov Success indicator for the Hill model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0
hill_rme Root mean square error for the Hill model
hill_tp The top parameter indicating the maximal estimated response
hill_ga The gain parameter for the Hill model, gain AC50
hill_p The power parameter for the Hill model
hill_er Error term for the Hill model
hill_tp_sd Standard deviation of the Hill model top parameter
hill_ga_sd Standard deviation of the Hill model gain parameter
hill_p_sd Standard deviation of the Hill model power parameter
hill_er_sd Standard deviation of the Hill model error term
hill_top The maximal response on the resulting Hill model fit
hill_ac50 Concentration at 50% of the maximal response on the Hill model fit
gnls_success Success indicator for the Gain-loss model; 1 if the optimization was successful, otherwise 0
gnls_aic Akaike Information Criteria (AIC) for the Gain-loss model
gnls_cov Success indicator for the Gain-loss model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0
gnls_rme Root mean square error for the Gain-loss model
gnls_tp The top parameter indicating the maximal estimated response
gnls_ga The gain parameter for the Gain-loss model, gain AC50
gnls_p The gain power parameter for the Gain-loss model
gnls_la The loss parameter for the Gain-loss model, loss AC50
gnls_q The loss power parameter for the Gain-loss model
gnls_er Error term for the Gain-loss model
gnls_tp_sd Standard deviation of the Gain-loss model top parameter
gnls_ga_sd Standard deviation of the Gain-loss model gain parameter
gnls_p_sd Standard deviation of the Gain-loss model gain power parameter
gnls_la_sd Standard deviation of the Gain-loss model loss parameter
gnls_q_sd Standard deviation of the Gain-loss model loss power parameter

gnls_er_sd Standard deviation of the Gain-loss model error term

gnls_top The maximal response on the resulting Gain-loss model fit

gnls_ac50 Concentration at 50% of the maximal response on the Gain-loss model fit, gain AC50

gnls_ac50_loss Concentration at 50% of the maximal response on the Gain-loss model fit, loss AC50

poly1_success Success indicator for the Polynomial 1 model; 1 if the optimization was successful, otherwise 0

poly1_aic Akaike Information Criteria (AIC) for the Polynomial 1 model

poly1_cov Success indicator for the Polynomial 1 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

poly1_rme Root mean square error for the Polynomial 1 model

poly1_a The y-scale parameter for the Polynomial 1 model

poly1_er Error term for the Polynomial 1 model

poly1_a_sd Standard deviation of the Polynomial 1 model y-scale parameter

poly1_er_sd Standard deviation of the Polynomial 1 model error term

poly1_top The maximal response on the resulting Polynomial 1 model fit

poly1_ac50 Concentration at 50% of the maximal response on the Polynomial 1 model fit

poly2_success Success indicator for the Polynomial 2 model; 1 if the optimization was successful, otherwise 0

poly2_aic Akaike Information Criteria (AIC) for the Polynomial 2 model

poly2_cov Success indicator for the Polynomial 2 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

poly2_rme Root mean square error for the Polynomial 2 model

poly2_a The y-scale parameter for the Polynomial 2 model

poly2_b The x-scale parameter for the Polynomial 2 model

poly2_er Error term for the Polynomial 2 model

poly2_a_sd Standard deviation of the Polynomial 2 model y-scale parameter

poly2_b_sd Standard deviation of the Polynomial 2 model x-scale parameter

poly2_er_sd Standard deviation of the Polynomial 2 model error term

poly2_top The maximal response on the resulting Polynomial 2 model fit

poly2_ac50 Concentration at 50% of the maximal response on the Polynomial 2 model fit

pow_success Success indicator for the Power model; 1 if the optimization was successful, otherwise 0

pow_aic Akaike Information Criteria (AIC) for the Power model

pow_cov Success indicator for the Power model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

pow_rme Root mean square error for the Power model

pow_a The y-scale parameter for the Power model

pow_p The power parameter for the Power model

pow_er Error term for the Power model

pow_a_sd Standard deviation of the Power model y-scale parameter

pow_p_sd Standard deviation of the Power model power parameter

pow_er_sd Standard deviation of the Power model error term

pow_top The maximal response on the resulting Power model fit

pow_ac50 Concentration at 50% of the maximal response on the Power model fit

exp2_success Success indicator for the Exponential 2 model; 1 if the optimization was successful, otherwise 0

exp2_aic Akaike Information Criteria (AIC) for the Exponential 2 model

exp2_cov Success indicator for the Exponential 2 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

exp2_rme Root mean square error for the Exponential 2 model

exp2_a The y-scale parameter for the Exponential 2 model

exp2_b The x-scale parameter for the Exponential 2 model

exp2_er Error term for the Exponential 2 model

exp2_a_sd Standard deviation of the Exponential 2 model y-scale parameter

exp2_b_sd Standard deviation of the Exponential 2 model x-scale parameter

exp2_er_sd Standard deviation of the Exponential 2 model error term

exp2_top The maximal response on the resulting Exponential 2 model fit

exp2_ac50 Concentration at 50% of the maximal response on the Exponential 2 model fit

exp3_success Success indicator for the Exponential 3 model; 1 if the optimization was successful, otherwise 0

exp3_aic Akaike Information Criteria (AIC) for the Exponential 3 model

exp3_cov Success indicator for the Exponential 3 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

exp3_rme Root mean square error for the Exponential 3 model

exp3_a The y-scale parameter for the Exponential 3 model

exp3_b The x-scale parameter for the Exponential 3 model

exp3_p The power parameter for the Exponential 3 model

exp3_er Error term for the Exponential 3 model

exp3_a_sd Standard deviation of the Exponential 3 model y-scale parameter

exp3_b_sd Standard deviation of the Exponential 3 model x-scale parameter

exp3_p_sd Standard deviation of the Exponential 3 model power parameter

exp3_er_sd Standard deviation of the Exponential 3 model error term

exp3_top The maximal response on the resulting Exponential 3 model fit

exp3_ac50 Concentration at 50% of the maximal response on the Exponential 3 model fit

exp4_success Success indicator for the Exponential 4 model; 1 if the optimization was successful, otherwise 0

exp4_aic Akaike Information Criteria (AIC) for the Exponential 4 model

exp4_cov Success indicator for the Exponential 4 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0

exp4_rme Root mean square error for the Exponential 4 model

exp4_tp The top parameter indicating the maximal estimated response

exp4_ga The gain parameter for the Exponential 4 model, gain AC50

exp4_er Error term for the Exponential 4 model

exp4_tp_sd Standard deviation of the Exponential 4 model top parameter

- exp4_ga_sd** Standard deviation of the Exponential 4 model gain parameter
 - exp4_er_sd** Standard deviation of the Exponential 4 model error term
 - exp4_top** The maximal response on the resulting Exponential 4 model fit
 - exp4_ac50** Concentration at 50% of the maximal response on the Exponential 4 model fit
 - exp5_success** Success indicator for the Exponential 5 model; 1 if the optimization was successful, otherwise 0
 - exp5_aic** Akaike Information Criteria (AIC) for the Exponential 5 model
 - exp5_cov** Success indicator for the Exponential 5 model covariance calculation; 1 if the Hessian matrix inversion is successful, otherwise 0
 - exp5_rme** Root mean square error for the Exponential 5 model
 - exp5_tp** The top parameter indicating the maximal estimated response
 - exp5_ga** The gain parameter for the Exponential 5 model, gain AC50
 - exp5_p** The power parameter for the Exponential 5 model
 - exp5_er** Error term for the Exponential 5 model
 - exp5_tp_sd** Standard deviation of the Exponential 5 model top parameter
 - exp5_ga_sd** Standard deviation of the Exponential 5 model gain parameter
 - exp5_p_sd** Standard deviation of the Exponential 5 model power parameter
 - exp5_er_sd** Standard deviation of the Exponential 5 model error term
 - exp5_top** The maximal response on the resulting Exponential 5 model fit
 - exp5_ac50** Concentration at 50% of the maximal response on the Exponential 5 model fit
 - all_onesd** Standard deviation of the baseline response for all models
 - all_bmed** Median noise estimation of the baseline response for all models
 - resp_unit** Response Units
 - conc_unit** Concentration Units
6. **mc5** A data frame with 5 rows and 54 columns containing level 5 best curve-fit and hitcall data.
- spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - aeid** Assay Component Endpoint ID
 - aenm** Assay endpoint name (i.e., assay_component_endpoint_name)
 - m5id** Level 5 (mc5) ID
 - m4id** Level 4 (mc4) ID
 - bmad** The median absolute deviation of all treatment wells (default option) or blank wells
 - resp_max** Maximum observed response
 - resp_min** Minimum observed response
 - max_mean** Maximum mean response
 - max_mean_conc** Concentration of the maximum mean response
 - max_med** Maximum median response

max_med_conc Concentration of the maximum median response
logc_max Maximum concentration on the log scale
logc_min Minimum concentration on the log scale
nconc The total number of concentration groups
npts Total number of observed responses (i.e. data points in the concentration series)
nrep Number of replicates in concentration groups
nmed_gtbl The number of median responses greater than 3BMAD
hitc Hitcall
modl Best model fit from tcplFit2 curve-fitting
fitc Fit category
coff Cutoff
top_over_cutoff Ratio of the top of the best model fit curve and the cutoff
rmse Root mean squared error
a The y-scale parameter for poly1, poly2, pow, exp2, or exp3 model
er Error term
bmr Benchmark response
bmdl Lower 95% confidence bound on the benchmark dose/concentration estimate
caikwt Akaike Information Criteria weight of constant model relative to the best model fit
mll Maximum log-likelihood of the best model fit
hitcall Continuous hitcall
ac50 Concentration where 50% of the maximal response occurs - if 'modl' is the Hill or Gain-loss model this is for the "gain" side of the response
top The maximal response on the best model curve fit - i.e. top of the curve fit
ac5 Concentration where 5% of the maximal response occurs
ac10 Concentration where 10% of the maximal response occurs
ac20 Concentration where 20% of the maximal response occurs
acc Concentration where the efficacy cutoff response occurs
ac1sd Concentration where one standard deviation of the background response occurs
bmd Benchmark response/concentration estimate - concentration where the benchmark response occurs
bmdu Upper 95% confidence bound on the benchmark dose/concentration estimate
tp The top curve parameter for the exp4, exp5, hill, or gnls model
ga The gain parameter for the hill or gnls model - gain AC50
p The power parameter for the pow, exp3, exp5, gnls, or hill model - for gnls this is the gain power parameter
q The loss power parameter for the gnls model
la The loss parameter for the gnls model, loss AC50
ac50_loss Concentration where 50% of the maximal response occurs - if 'modl' is the Hill or Gain-loss model this is for the "loss" side of the response
b The x-scale parameter for poly2, exp2, or exp3 model
resp_unit Response Units
conc_unit Concentration Units

Method functions

Functions for managing processing methods

Description

These functions are used to manage which methods are used to process data. They include methods for assigning, clearing, and loading the assigned methods. Also, `tcplMthdList` lists the available methods.

Usage

```
tcplMthdAssign(lvl, id, mthd_id, ordr = NULL, type)
```

```
tcplMthdClear(lvl, id, mthd_id = NULL, type)
```

```
tcplMthdList(lvl, type = "mc")
```

```
tcplMthdLoad(lvl, id = NULL, type = "mc")
```

Arguments

<code>lvl</code>	Integer of length 1, the method level
<code>id</code>	Integer, the assay component or assay endpoint id(s)
<code>mthd_id</code>	Integer, the method id(s)
<code>ordr</code>	Integer, the order in which to execute the analysis methods, must be the same length as <code>mthd_id</code> , does not apply to levels 5 or 6
<code>type</code>	Character of length 1, the data type, "sc" or "mc"

Details

`tcplMthdLoad` loads the assigned methods for the given level and ID(s). Similarly, `tcplMthdList` displays the available methods for the given level. These two functions do not make any changes to the database.

Unlike the `-Load` and `-List` functions, the `-Assign` and `-Clear` functions alter the database and trigger a delete cascade. `tcplMthdAssign` assigns methods to the given ID(s), and `tcplMthdClear` removes methods. In addition to the method ID (`'mthd_id'`), assigning methods at some levels require an order (`'ordr'`). The `'ordr'` parameter is necessary to allow progression of methods at level one for single-concentration processing, and levels two and three for multiple-concentration processing. More information about method assignments and the delete cascade are available in the package vignette.

Examples

```
## Not run:
## tcplListMthd allows the user to display the available methods for
## a given level and data type
```

```

head(tcplMthdList(lvl = 2, type = "mc"))

## tcplLoadMthd shows which methods are assigned for the given ID, level,
## and data type. Here we will show how to register, load, and clear methods
## using an acid not in the example database. Note: There is no check for
## whether an ID exists before assigning/clearing methods.
tcplMthdLoad(lvl = 2, id = 55, type = "mc")

## ACID 55 does not have any methods. Assign methods from the list above.
tcplMthdAssign(lvl = 2,
               id = 55,
               mthd_id = c(3, 4, 2),
               ord = 1:3,
               type = "mc")
## Method assignment can be done for multiple assays, too.
tcplMthdAssign(lvl = 2,
               id = 53:54,
               mthd_id = c(3, 4, 2),
               ord = 1:3,
               type = "mc")

## Cleanup example method assignments
tcplMthdClear(lvl = 2, id = 53:55, type = "mc")

## End(Not run)

```

Models

Model objective functions

Description

These functions take in the dose-response data and the model parameters, and return a likelihood value. They are intended to be optimized using `constrOptim` in the `tcplFit` function.

Usage

```

tcplObjCnst(p, resp)

tcplObjGnls(p, lconc, resp)

tcplObjHill(p, lconc, resp)

tcplObjCnst(p, resp)

tcplObjGnls(p, lconc, resp)

tcplObjHill(p, lconc, resp)

```


Arguments

p	Numeric, the parameter values. See details for more information.
resp	Numeric, the response values
lconc	Numeric, the log10 concentration values

Details

These functions produce an estimated value based on the model and given parameters for each observation. Those estimated values are then used with the observed values and a scale term to calculate the log-likelihood.

Let $t(z, \nu)$ be the Student's t-distribution with ν degrees of freedom, y_i be the observed response at the i^{th} observation, and μ_i be the estimated response at the i^{th} observation. We calculate z_i as:

$$z_i = \frac{y_i - \mu_i}{e^\sigma}$$

where σ is the scale term. Then the log-likelihood is:

$$\sum_{i=1}^n [\ln(t(z_i, 4)) - \sigma]$$

Where n is the number of observations.

Value

The log-likelihood.

Constant Model (cnst)

tcpl0bjCnst calculates the likelihood for a constant model at 0. The only parameter passed to tcpl0bjCnst by p is the scale term σ . The constant model value μ_i for the i^{th} observation is given by:

$$\mu_i = 0$$

tcpl0bjCnst calculates the likelihood for a constant model at 0. The only parameter passed to tcpl0bjCnst by p is the scale term σ . The constant model value μ_i for the i^{th} observation is given by:

$$\mu_i = 0$$

Gain-Loss Model (gnls)

tcpl0bjGnls calculates the likelihood for a 5 parameter model as the product of two Hill models with the same top and both bottoms equal to 0. The parameters passed to tcpl0bjGnls by p are (in order) top (tp), gain log AC50 (ga), gain hill coefficient (gw), loss log AC50 (la), loss hill coefficient (lw), and the scale term (σ). The gain-loss model value μ_i for the i^{th} observation is given by:

$$g_i = \frac{1}{1 + 10^{(ga - x_i)gw}}$$

$$l_i = \frac{1}{1 + 10^{(x_i - la)lw}}$$

$$\mu_i = tp(g_i)(l_i)$$

where x_i is the log concentration for the i^{th} observation.

`tcp1objGnls` calculates the likelihood for a 5 parameter model as the product of two Hill models with the same top and both bottoms equal to 0. The parameters passed to `tcp1objGnls` by `p` are (in order) top (tp), gain log AC50 (ga), gain hill coefficient (gw), loss log AC50 la , loss hill coefficient lw , and the scale term (σ). The gain-loss model value μ_i for the i^{th} observation is given by:

$$g_i = \frac{1}{1 + 10^{(ga-x_i)gw}}$$

$$l_i = \frac{1}{1 + 10^{(x_i-la)lw}}$$

$$\mu_i = tp(g_i)(l_i)$$

where x_i is the log concentration for the i^{th} observation.

Hill Model (hill)

`tcp1objHill` calculates the likelihood for a 3 parameter Hill model with the bottom equal to 0. The parameters passed to `tcp1objHill` by `p` are (in order) top (tp), log AC50 (ga), hill coefficient (gw), and the scale term (σ). The hill model value μ_i for the i^{th} observation is given by:

$$\mu_i = \frac{tp}{1 + 10^{(ga-x_i)gw}}$$

where x_i is the log concentration for the i^{th} observation.

`tcp1objHill` calculates the likelihood for a 3 parameter Hill model with the bottom equal to 0. The parameters passed to `tcp1objHill` by `p` are (in order) top (tp), log AC50 (ga), hill coefficient (gw), and the scale term (σ). The hill model value μ_i for the i^{th} observation is given by:

$$\mu_i = \frac{tp}{1 + 10^{(ga-x_i)gw}}$$

where x_i is the log concentration for the i^{th} observation.

MORELETTERS

MORELETTERS Extends LETTERS recursively to any number of letters

Description

MORELETTERS Extends LETTERS recursively to any number of letters

Usage

`MORELETTERS(i)`

Arguments

`i` numeric vector of all needed letter combinations, i.e. `1:nrow(dat)`

Value

char vector containing all letter/combinations

Author(s)

<https://stackoverflow.com/a/25881167>

mthd_list_defaults	<i>Lists of data frames returned from tcplMthdList invitrodb v4.2</i>
--------------------	---

Description

Lists of data frames returned from tcplMthdList invitrodb v4.2

Usage

mthd_list_defaults

Format

A list with 7 items:

mc2 displays the available methods for mc lvl 2 data

mc3 displays the available methods for mc lvl 3 data

mc4 displays the available methods for mc lvl 4 data

mc5 displays the available methods for mc lvl 5 data

mc6 displays the available methods for mc lvl 6 data

sc1 displays the available methods for sc lvl 1 data

sc2 displays the available methods for sc lvl 2 data

Source

ToxCast database

Query functions

Wrappers for sending queries and fetching results

Description

These functions send a query to the given database, and are the access point for all tcpl functions that query or update the tcpl database.

Usage

```
tcplQuery(
  query,
  db = getOption("TCPL_DB"),
  drvr = getOption("TCPL_DRVR"),
  tbl = NULL
)

tcplQueryAPI(resource = "data", fld = NULL, val = NULL, return_flds = NULL)

tcplSendQuery(
  query,
  db = getOption("TCPL_DB"),
  drvr = getOption("TCPL_DRVR"),
  tbl = NULL,
  delete = F
)
```

Arguments

query	Character of length 1, the query string
db	Character of length 1, the name of the tcpl database
drvr	Character of length 1, which database driver to use
tbl	Tables to be read queried
resource	must be either data or assay to determine which api endpoint to hit
fld	field that should be used to query the api
val	value for specified field to query on
return_flds	optional list of fields that should be returned
delete	Logical of length 1, execute delete on queried table

Details

Currently, the tcpl package supports the "MySQL", "example", and "API" database drivers.

tcplQuery returns a data.table object with the query results. tcplSendQuery sends a query, but does not fetch any results, and returns 'TRUE' or the error message given by the database. tcplQueryAPI returns a data.table object with the query results when connected using "API" as driver.

Examples

```
## Not run:
# only with MySQL driver
tcplQuery("SELECT 'Hello World';")

# only with API driver
tcplConfDefault()
tcplQueryAPI(resource = "data", fld = "aeid", val = 2)

## End(Not run)
```

Register/update annotation

Functions for registering & updating annotation information

Description

These functions are used to register and update the chemical and assay annotation information.

Usage

```
tcplRegister(what, flds)

tcplUpdate(what, id, flds)
```

Arguments

what	Character of length 1, the name of the ID to register or update
flds	Named list, the other fields and their values
id	Integer, the ID value(s) to update

Details

These functions are used to populate the tcpl database with the necessary annotation information to complete the processing. As shown in the package vignette, the package requires some information about the samples and assays before data can be loaded into the tcpl database.

Depending on what is being registered, different information is required. The following table lists the fields that can be registered/updated by these functions, and the minimal fields required for registering a new ID. (The database table affected is in parentheses.)

- asid (assay_source): assay_source_name
- aid (assay): asid, assay_name, assay_footprint
- acid (assay_component): aid, assay_component_name
- aeid (assay_component_endpoint): acid, assay_component_endpoint_name, normalized_data_type
- acsn (assay_component_map): acid, acsn

- spid (sample): spid, chid
- chid (chemical): chid, casn
- clib (chemical_library): chid, clib

Note: The functions accept the abbreviated forms of the names, ie. "aenm" rather than the full "assay_component_endpoint_name." More information about the registration process and all of the fields is available in the vignette.

Examples

```
## Not run:
## Load current ASID information
tcplLoadAsid()

## Register a new assay source
tcplRegister(what = "asid", flds = list(asnm = "example_asid"))

## Show the newly registered ASID
tcplLoadAsid(add.fld = "assay_source_desc")

## Notice that the newly created ASID does not have an assay_source_desc.
## The field could have been defined during the registration process, but
## can also be updated using tcplUpdate
i1 <- tcplLoadAsid()[asnm == "example_asid", asid]
tcplUpdate(what = "asid",
           id = i1,
           flds = list(assay_source_desc = "example asid description"))
tcplLoadAsid(add.fld = "assay_source_desc")

## Remove the created ASID. Note: Manually deleting primary keys can cause
## serious database problems and should not generally be done.

## End(Not run)
```

registerMthd	<i>Add a new analysis method</i>
--------------	----------------------------------

Description

registerMthd registers a new analysis method to the tcpl databases.

Usage

```
registerMthd(lvl, mthd, desc, naddr = 0L, type)
```

Arguments

lvl	Integer of length 1, the level for the analysis method
mthd	Character, the name of the method
desc	Character, same length as mthd, the method description
nddr	Integer, 0 or 1, 1 if the method requires loading the dose- response data
type	Character of length 1, the data type, "sc" or "mc"

Details

'mthd' must match a corresponding function name in the functions that load the methods, ie. mc2_mthds. 'nddr' only applies to level 6 methods.

round_n	<i>round_n General function to round/shorten values for plotting tables</i>
---------	---

Description

round_n General function to round/shorten values for plotting tables

Usage

```
round_n(x, n = 3)
```

Arguments

x	numeric value
n	numeric number of decimal places to round to

Value

format string in decimal or scientific notation

sc1	<i>Perform level 1 single-concentration processing</i>
-----	--

Description

sc1 loads level 0 data from the tcpl database for the given id and performs level 1 single-concentration processing. The processed data is then loaded into the sc1 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
sc1(ac, wr = FALSE)
```

Arguments

- ac Integer of length 1, assay component id (acid) for processing.
- wr Logical, whether the processed data should be written to the tcpl database

Details

Level 1 single-concentration processing includes mapping assay component to assay endpoint, duplicating the data when the assay component has multiple assay endpoints, and any normalization of the data. Data normalization based on methods listed in `sc1_aeid` and `sc1_methods` tables.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a `data.table` containing the processed data

See Also

[Method functions, SC1_Methods](#)
Other single-concentration: [sc2\(\)](#)

SC1_Methods	<i>List of level 1 single-concentration normalization functions</i>
-------------	---

Description

`sc1_mthds` returns a list of functions to be used during level 1 single-concentration processing.

Usage

```
sc1_mthds()
```

Details

The functions contained in the list returned by `sc1_mthds` return a list of expressions to be executed in the `sc2` (not exported) function environment. The functions are described here for reference purposes, The `sc1_mthds` function is not exported, nor is it intended for use.
All available methods are described in the Available Methods section, listed by the function/method name.

Value

A list functions

Available Methods

The methods are broken into three types, based on what fields they define. Different methods are used to define "bval" (the baseline value), "pval" (the positive control value), and "resp" (the final response value).

Although it does not say so specifically in each description, all methods are applied by acid.

More information about the level 3 single-concentration processing is available in the package vignette, "Data_processing."

bval Methods:

bval.apid.nwlls.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the raw values (rval) for neutral control wells (wllt = n).

bval.apid.twlls.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the raw values (rval) for test compound wells (wllt = t).

bval.apid.tn.med Calculate the baseline value (bval) as the plate-wise median, by assay plate ID (apid), of the raw values (rval) for test compound wells (wllt = t) and neutral control wells (wllt = n).

bval.nwlls.med Calculate the baseline value (bval) as the median of the raw values (rval) for neutral control wells (wllt = n) by assay endpoint id (aeid).

pval Methods:

pval.apid.pwlls.med Calculate the positive control value (pval) as the plate-wise median, by assay plate ID (apid), of the raw values (rval) for single-concentration gain-of-signal positive control wells (wllt = p).

pval.apid.mwlls.med Calculate the positive control value (pval) as the plate-wise median, by assay plate ID (apid), of the raw values (rval) for multiple-concentration loss-of-signal negative control wells (wllt = m).

pval.apid.medpcbyconc.max Calculate the positive control value (pval) as the plate-wise maximum, by assay plate ID (apid), of the medians of the raw values (rval) for gain-of-signal single- or multiple-concentration positive control wells (wllt = p or c) by apid, well type, and concentration.

pval.apid.medpcbyconc.min Calculate the positive control value (pval) as the plate-wise minimum, by assay plate ID (apid), of the medians of the raw values (rval) for gain-of-signal single- or multiple-concentration positive control wells (wllt = p or c) by apid, well type, and concentration.

pval.apid.medncbyconc.min Calculate the positive control value (pval) as the plate-wise minimum, by assay plate ID (apid), of the medians of the raw values (rval) for gain-of-signal single- or multiple-concentration negative control wells (wllt = m or o) by apid, well type, and concentration.

pval.zero Set the positive control value (pval) to 0; pval = 0.

pval.apid.or.aeid.pwlls.med Calculate the positive control value (pval) as the plate-wise median, by assay plate ID (apid), of the raw values (rval) for single-concentration gain-of-signal positive control wells (wllt = p). For plates without p wells, set the pval as the median pval calculated from all plates.

resp Methods:

- resp.pc** Calculate the normalized response (resp) as a percent of control, i.e. the ratio of the difference between the raw (rval) and baseline (bval) values divided by the difference between positive control (pval) and baseline (bval) values multiplied by 100; $resp = (rval - bval) / (pval - bval) * 100$.
- resp.fc** Calculate the normalized response (resp) as fold change, i.e. the ratio of the raw (rval) and baseline (bval) values; $resp = rval / bval$.
- resp.logfc** Calculate the normalized response (resp) as the fold change of logged, i.e. the difference between raw (rval) and baseline (bval) log-scale values.
- resp.log2** Transform the response values to log-scale (base 2).
- resp.multneg1** Multiply the normalized response value (resp) by -1; $-1 * resp$.
- none** Use raw value (rval) as is. This may be necessary for additional endpoint-specific adjustments, or where no additional sc1 methods are needed.
- resp.incr.zerocenter.fc** Calculate the normalized response (resp) as a zero center fold change, i.e. the ratio of the raw (rval) and baseline (bval) values minus 1; $resp = rval / bval - 1$. Typically used for increasing responses.

Note

This function is not exported and is not intended to be used by the user.

See Also

[sc1](#), [Method functions](#) to query what methods get applied to each acid

sc2

Perform level 2 single-concentration processing

Description

sc2 loads level 1 data from the tcpl database for the given id and performs level 2 single-concentration processing. The processed data is then loaded into the sc2 table and all subsequent data is deleted with [tcplCascade](#). See details for more information.

The individual processing functions are no longer exported, as it is typically more convenient and suggested to use the [tcplRun](#) wrapper function.

Usage

```
sc2(ae, wr = FALSE)
```

Arguments

- | | |
|----|--|
| ae | Integer of length 1, assay endpoint id (aeid) for processing. |
| wr | Logical, whether the processed data should be written to the tcpl database |

Details

Level 2 single-concentration processing defines the bmad value, and uses the activity cutoff methods from `sc2_aeid` and `sc2_methods` to make an activity call.

Value

A boolean of length 1, indicating the success of the processing, or when 'wr' is FALSE, a list where the first element is a boolean indicating the success of processing and the second element is a `data.table` containing the processed data

See Also

[Method functions, SC2_Methods](#)

Other single-concentration: [sc1\(\)](#)

SC2_Methods

List of level 2 single-concentration hit-call functions

Description

`sc2_mthds` returns a list of functions to be used during level 2 single-concentration processing.

Usage

```
sc2_mthds()
```

Details

The functions contained in the list returned by `sc2_mthds` return a list of expressions to be executed in the `sc2` (not exported) function environment. The functions are described here for reference purposes. The `sc2_mthds` function is not exported, nor is it intended for use.

All available methods are described in the Available Methods section, listed by the function/method name.

Value

A list functions

Available Methods

The methods are broken down into four categories based on the type of cutoff they assign. Different methods are used to define cutoffs for "bmad" (baseline median absolute value), "pc" (percent of control), "pc or bmad", "log" (\log_2 or \log_{10}), and "other" (uncategorized methods).

All methods are applied by `aeid`.

Although there are method exceptions (notably within the "other" category), only highest calculated cutoff value based on assigned methods will be selected for hitcalling. Therefore, only the largest cutoff method per method type should be assigned.

More information about the level 2 single-concentration processing is available in the package vignette, "Data_processing."

BMAD Methods:

bmad1 Add a cutoff value of 1 multiplied by baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad1.5 Add a cutoff value of 1.5 multiplied by the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad2 Add a cutoff value of 2 multiplied by the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad3 Add a cutoff value of 3 multiplied by the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad5 Add a cutoff value of 5 multiplied the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad6 Add a cutoff value of 6 multiplied by the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

bmad10 Add a cutoff value of 10 multiplied by the baseline median absolute deviation (bmad). By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

Percent of Control Methods:

pc0.88 Add a cutoff value of 0.88. Typically for percent of control data.

pc16 Add a cutoff value of 16. Typically for percent of control data.

pc20 Add a cutoff value of 20. Typically for percent of control data.

pc25 Add a cutoff value of 25. Typically for percent of control data.

pc30 Add a cutoff value of 30. Typically for percent of control data.

pc40 Add a cutoff value of 40. Typically for percent of control data.

Percent of Control or BMAD Methods:

pc30orbmad3 Add a cutoff value of either 30 or 3 multiplied by the baseline median absolute deviation (bmad), whichever is less. By default, bmad is calculated using test compound wells (wlIt = t) for the endpoint.

Log Methods: Log Base 2

log2_0.76 Add a cutoff value of 0.76 for log2-transformed data. This was a custom threshold value set for endpoint id 1690 (formerly acid 1691).

log2_1.2 Add a cutoff value of $\log_2 1.2$. Typically for fold change data.

log2_1.5 Add a cutoff value of $\log_2 1.5$. Typically for fold change data.

Log Base 10

log10_1.2 Add a cutoff value of $\log_{10} 1.2$. Typically for fold change data.

Other Methods:

ow_bmad_nwells Overwrite the default baseline median absolute value (bmad) with a bmad calculated using neutral control wells (wlIt = n).

ow_bidirectional_gain Where responses only in the positive direction are biologically relevant, overwrite the max_med and max_tmp values, which were calculated using absolute value, to a calculation using a true maximum for uni-directional data.

ow_bidirectional_loss Where responses only in the negative direction are biologically relevant, overwrite the max_med and max_tmp values, which were calculated using absolute value, to a calculation using a true minimum for uni-directional data.

Note

This function is not exported and is not intended to be used by the user.

See Also

[sc2](#), [Method functions](#) to query what methods get applied to each acid

smdat	<i>A subset of ToxCast data showing changes in transcription factor activity for multiple targets.</i>
-------	--

Description

The example dataset is used to illustrate how the user can pipeline single-concentration data from chemical screening using tcpl.

Usage

```
smdat
```

Format

A data frame with 320 rows and 10 variables:

spid sample ID
apid assay plate ID
rowi well-plate row number (N/A)
coli well-plate column number (N/A)
wllt well type (N/A)
wllq well quality (N/A)
conc concentration in micromolar
rval raw assay component readout value
srcf source file containing the data
acsn assay component source name

Source

ToxCast database

sc_test	<i>List of lists containing queries sent to tcplQuery associated with each test case. Each list also contains the associated ids with each case. Only meant to be used with automated testing with mocking for sc data.</i>
---------	---

Description

List of lists containing queries sent to tcplQuery associated with each test case. Each list also contains the associated ids with each case. Only meant to be used with automated testing with mocking for sc data.

Usage

sc_test

Format

A list with 20 items:

tcplConfQuery Data table with 1 row and 2 columns used for each test case for establishing connection using tcplConf. This data table mocks the response one would get from connecting with invitrodb.

sc0_by_s0id List containing the queries used for loading sc0 data by s0id via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 's0id' labeled item storing the id used to load the data, for use in tests.

sc0_by_acid List containing the queries used for loading sc0 data by acid via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'acid' labeled item storing the id used to load the data, for use in tests.

sc1_by_s1id List containing the queries used for loading sc1 data by s1id via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 's1id' labeled item storing the id used to load the data, for use in tests.

sc1_by_acid List containing the queries used for loading sc1 data by acid via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'acid' labeled item storing the id used to load the data, for use in tests.

sc2_by_s2id List containing the queries used for loading sc2 data by s2id via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 's2id' labeled item storing the id used to load the data, for use in tests.

sc2_by_aeid List containing the queries used for loading sc2 data by aeid via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.

scagg_by_aeid List containing the queries used for loading sc 'agg' data by aeid via tcplLoadData. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.

- plot_single_s2id** List containing the queries used for loading and plotting sc data by s2id via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains one 's2id' labeled item storing the id used to load the data, for use in tests.
- plot_multiple_s2id** List containing the queries used for loading and plotting sc data by multiple s2ids via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains one 's2id' labeled item storing the ids used to load the data, for use in tests.
- plot_single_aeid** List containing the queries used for loading and plotting sc data by aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the id used to load the data, for use in tests.
- plot_multiple_aeid** List containing the queries used for loading and plotting sc data by multiple auids via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains one 'aeid' labeled item storing the ids used to load the data, for use in tests.
- plot_single_spid** List containing the queries used for loading and plotting sc data by spid/aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'spid' and 'aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_multiple_spid** List containing the queries used for loading and plotting sc data by multiple spids/aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'spid' and 'aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_single_s2id_compare** List containing the queries used for loading and plotting compared sc data by s2id via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 's2id' and 'compare.s2id' labeled items storing the ids used to load the data, for use in tests.
- plot_multiple_s2id_compare** List containing the queries used for loading and plotting compared sc data by multiple s2ids via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 's2id' and 'compare.s2id' labeled items storing the ids used to load the data, for use in tests.
- plot_single_aeid_compare** List containing the queries used for loading and plotting compared sc data by aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'aeid' and 'compare.aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_multiple_aeid_compare** List containing the queries used for loading and plotting compared sc data by multiple auids via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'aeid' and 'compare.aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_single_spid_compare** List containing the queries used for loading and plotting compared sc data by spid/aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'spid', 'compare.spid', 'aeid', and 'compare.aeid' labeled items storing the ids used to load the data, for use in tests.
- plot_multiple_spid_compare** List containing the queries used for loading and plotting compared sc data by multiple spids/aeid via tcplPlot. Each query has an associated data table response for mocking an actual connection. Contains 'spid', 'compare.spid', 'aeid', and 'compare.aeid' labeled items storing the ids used to load the data, for use in tests.

Source

ToxCast database

sc_vignette

List with single-concentration data for the vignette

Description

This dataset is a list with 3 data.tables (sc0,sc1,sc2).

Usage

```
sc_vignette
```

Format

1. **sc0** A data frame with 10 rows and 18 columns containing level 0 formatted raw data.

spid Sample ID
chid Unique chemical ID number for tcpl
casn Chemical Abstract Service(CAS) number
chnm Chemical name
dsstox_substance_id Chemical-specific DTXSID
code CAS number compressed into numeric string
acid Assay Component ID
acnm Assay Component Name
s0id Level 0 (sc0) ID
apid Assay plate ID
rowi Row Index
coli Column Index
wllt Well Type
wllq Well Quality (0 or 1)
conc Concentration in micromolar
rval Raw assay component readout value
srcf Source file containing the raw data
conc_unit Concentration Units

2. **sc1** A data frame with 10 rows and 20 columns containing level 1 normalized data.

spid Sample ID
chid Unique chemical ID number for tcpl
casn Chemical Abstract Service(CAS) number
chnm Chemical name
dsstox_substance_id Chemical-specific DTXSID
code CAS number compressed into numeric string

- aecid** Assay Component Endpoint ID
 - aenm** Assay endpoint name (i.e., assay_component_endpoint_name)
 - acid** Assay Component ID
 - acnm** Assay Component Name
 - s0id** Level 0 (sc0) ID
 - s1id** Level 1 (sc1) ID
 - apid** Assay plate ID
 - rowi** Row Index
 - coli** Column Index
 - wllt** Well Type
 - logc** Log base 10 concentration
 - resp** Normalized response value
 - resp_unit** Response Units
 - conc_unit** Concentration Units
3. **sc2** A data frame with 10 rows and 15 columns containing level 2 efficacy/hit designation data.
- spid** Sample ID
 - chid** Unique chemical ID number for tcpl
 - casn** Chemical Abstract Service(CAS) number
 - chnm** Chemical name
 - dsstox_substance_id** Chemical-specific DTXSID
 - code** CAS number compressed into numeric string
 - aecid** Assay Component Endpoint ID
 - aenm** Assay endpoint name (i.e., assay_component_endpoint_name)
 - s2id** Level 2 (sc2) ID
 - bmad** The median absolute deviation of all treatment wells (default option) or blank wells
 - max_med** Maximum median response
 - hitc** Hitcall
 - coff** Cutoff
 - resp_unit** Response Units
 - conc_unit** Concentration Units

sink.reset

Reset all sinks

Description

sink.reset resets all sinks and returns all output to the console.

Usage

```
sink.reset()
```

Details

`sink.reset` identifies all sinks with `sink.number` then returns all output and messages back to the console.

See Also

[sink](#), [sink.number](#)

Other tcpl abbreviations: [is.odd\(\)](#), [lu\(\)](#), [lw\(\)](#)

tcplAddModel

Draw a tcpl Model onto an existing plot

Description

`tcplAddModel` draws a a line for one of the tcpl Models (see [Models](#) for more information) onto an existing plot.

Usage

```
tcplAddModel(pars, modl = NULL, adj = NULL, ...)
```

Arguments

<code>pars</code>	List of parameters from level 4 or 5 output
<code>modl</code>	Character of length 1, the model to plot: 'cnst,' 'hill,' or 'gnls'
<code>adj</code>	Numeric of length 1, an adjustment factor, see details for more information
<code>...</code>	Additional arguments passed to curve

Details

`tcplAddModel` draws the model line assuming the x-axis represents log base 10 concentration.

If `modl` is `NULL`, the function checks `pars$modl` and will return an error if `pars$modl` is also `NULL`.

`adj` is intended to scale the models, so that models with different response units can be visualized on a single plot. The recommended value for `adj` is $1/(3 \cdot \text{bmad})$ for level 4 data and $1/\text{coeff}$ for level 5 data. If `adj` is `NULL` the function will check `pars$adj` and set `adj` to 1 if `pars$adj` is also `NULL`.

See Also

[Models](#), [tcplPlotFits](#)

Examples

```
## Create some dummy data to plot
logc <- 1:10
r1 <- sapply(logc, tcplHillVal, ga = 5, tp = 50, gw = 0.5)
r2 <- log2(sapply(logc, tcplHillVal, ga = 4, tp = 30, gw = 0.5))
p1 <- tcplFit(logc = logc, resp = r1, bmad = 10)
p2 <- tcplFit(logc = logc, resp = r2, bmad = log2(1.5))

## In the dummy data above, the two plots are on very different scales
plot(r1 ~ logc, pch = 16, ylab = "raw response")
tcplAddModel(pars = p1, modl = "hill")
points(r2 ~ logc)
tcplAddModel(pars = p2, modl = "hill", lty = "dashed")

## To visualize the two curves on the same plot for comparison, we can
## scale the values to the bmad, such that a scaled response of 1 will equal
## the bmad for each curve.
plot(r1/10 ~ logc, pch = 16, ylab = "scaled response")
tcplAddModel(pars = p1, modl = "hill", adj = 1/10)
points(r2/log2(5) ~ logc)
tcplAddModel(pars = p2, modl = "hill", adj = 1/log2(5), lty = "dashed")
```

tcplAICProb

*Calculate the AIC probabilities***Description**

tcplAICProb Calculates the probability that the model best represents the data based on the AIC value for each model.

Usage

```
tcplAICProb(...)
```

Arguments

... Numeric vectors of AIC values

Details

The function takes vectors of AIC values. Each vector represents the model AIC values for multiple observation sets. Each vector must contain the same number and order of observation sets. The calculation assumes every possible model is accounted for, and the results should be interpreted accordingly.

Value

A vector of probability values for each model given, as a list.

See Also

[tcplFit](#), [AIC](#) for more information about AIC values.

Examples

```
## Returns the probability for each model, given models with AIC values
## ranging from 80 to 100
tcplAICProb(80, 85, 90, 95, 100)

## Also works for vectors
m1 <- c(95, 195, 300) ## model 1 for three different observations
m2 <- c(100, 200, 295) ## model 2 for three different observations
tcplAICProb(m1, m2)
```

tcplAppend	<i>Append rows to a table</i>
------------	-------------------------------

Description

tcplAppend takes a data.table (dat) and appends the data.table into a database table.

Usage

```
tcplAppend(dat, tbl, db, lvl = NULL)
```

Arguments

- dat data.table, the data to append to a table
- tbl Character of length 1, the table to append to
- db Character of length 1, the database containing tbl
- lvl Usually Integer to indicate what level to auto-increment

Note

This function is not exported and not intended to be used by the user.

tcplCascade	<i>Do a cascading delete on tcpl screening data</i>
-------------	---

Description

tcplCascade deletes the data for the given id(s) starting at the processing level given. The delete will cascade through all subsequent tables.

Usage

```
tcplCascade(lvl, type, id)
```

Arguments

lvl	Integer of length 1, the first level to delete from
type	Character of length 1, the data type, "sc" or "mc"
id	Integer, the id(s) to delete. See details for more information.

Details

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the level tables, whereas the single concentration will be loaded into the single tables.

If lvl is less than 3, id is interpreted as acid(s) and if lvl is greater than or equal to 3, id is interpreted as aacid(s).

Note

This function is not exported and not intended to be used by the user.

tcplCode2CASN	<i>Convert chemical code to CAS Registry Number</i>
---------------	---

Description

tcplCode2CASN takes a code and converts it CAS Registry Number.

Usage

```
tcplCode2CASN(code)
```

Arguments

code	Character of length 1, a chemical code
------	--

Details

The function checks for the validity of the CAS Registry Number. Also, the ToxCast data includes chemicals for which there is no CASRN. The convention for these chemicals is to give them a CASRN as NOCAS_chid; the code for these compounds is CNOCASchid. The function handles the NOCAS compounds as they are stored in the database, as shown in the example below.

Value

A CAS Registry Number.

Examples

```
tcplCode2CASN("C80057")
tcplCode2CASN("C09812420") ## Invalid CASRN will give a warning
tcplCode2CASN("CNOCAS0015") ## The underscore is reinserted for NOCAS codes
```

tcplCytoPt

Calculate the cytotoxicity point based on the "burst" endpoints

Description

tcplCytoPt calculates the cytotoxicity point and average cytotoxicity distribution based on the activity in the "burst" assay endpoints.

Usage

```
tcplCytoPt(
  chid = NULL,
  aeid = NULL,
  flag = TRUE,
  min.test = TRUE,
  default.pt = 3
)
```

Arguments

chid	Integer, chemical ID values to subset on
aeid	Integer, assay endpoint ID values to override the "burst assay" definitions
flag	Integer, mc6_mthd_id values to be passed to tcplSubsetChid
min.test	Integer or Boolean, the number of tested assay endpoints required for a chemical to be used in calculating the "global MAD."
default.pt	Numeric of length 1, the default cytotoxicity point value

Details

tcplCytoPt provides estimates for chemical-specific cytotoxicity distributions (more information available in the vignette.) Before calculating the cytotoxicity distributions, the level 5 data is subsetted by the `tcplSubsetChid` function.

The 'chid' parameter specifies a subset of chemicals to use in the calculations, given by chemical ID (chid). The 'aeid' parameter specifies which assays to use in calculating the cytotoxicity point and distribution. By default tcplCytoPt will use all available chemicals and the assay endpoints defined by the 'burst_assay' field in the "assay_component_endpoint" table. The examples show how to identify the "burst" endpoints.

tcplCytoPt returns the cytotoxicity point (the AC50 values of the active "burst" endpoints), the corresponding MAD, and the global MAD (median of the calculated MAD values). Not every chemical must be tested in every "burst" endpoint. The 'min.test' parameter allows the user to specify a minimum number of tested assay endpoints as a requirement for MAD values to be included in the global MAD calculation. For example, suppose the user supplies 10 "burst" assays. The user can choose to require a chemical to be tested in at least 5 of those assays for its MAD value to be included in the global MAD calculation. Having chemicals with many less "burst" endpoints tested may inflate or deflate the global MAD calculation. By default (values of TRUE or NULL), tcplCytoPt requires a chemical to be tested in at least 80% of the given "burst" assays. The user can also provide 'min.test' values of FALSE (indicating to include all MAD values), or a number (indicating a specific number of endpoints).

Chemicals without at least 2 active "burst" assays do not have a MAD value, and the cytotoxicity point is defined by the 'default.pt' parameter. The default value for 'default.pt' is 3.

The resulting data.table has the following fields:

1. "chid" – The chemical ID.
2. "code" – The chemical code.
3. "chnm" – The chemical name.
4. "casn" – The chemical CASRN.
5. "med" – The median of the "burst" endpoint log(AC50)
6. "mad" – The MAD of the "burst" endpoint log(AC50) values.
7. "ntst" – The number of "burst" endpoints tested.
8. "nhit" – The number of active "burst" endpoints.
9. "used_in_global_mad_calc" – TRUE/FALSE, whether the mad value was used in the global MAD calculation.
10. "global_mad" – The median of the "mad" values where "used_in_global_mad_calc" is TRUE.
11. "cyto_pt" – The cytotoxicity point, or the value in "med" when "nhit" is at least 2.
12. "cyto_pt_um" – $10^{\text{cyto_pt}}$
13. "lower_bnd_um" – $10^{\text{cyto_pt} - 3\text{global_mad}}$

Value

A data.table with the cytotoxicity distribution for each chemical. The definition of the field names are listed under "details."

Examples

```
## Not run:
## Can only calculate the cytotox burst if using the MySQL database and
## TCPL_DRVR == 'MySQL'

if (getOption("TCPL_DRVR") == "MySQL") {

  ## Load the "burst" endpoints -- none are defined in the example dataset
  tcplloadAeid(fld = "burst_assay", val = 1)

  ## Calculate the cytotoxicity distributions using both example endpoints
  tcplCytoPt(aeid = 1:2)

  ## The above example does not calculate a global MAD, because no chemical
  ## hit both endpoints. (This makes sense, because both endpoints are
  ## derived from one component, where one endpoint is activity in the
  ## up direction, and the other is activity in the down direction.)
  ## Note, the cyto_pt is also 3 for all chemicals, because the function
  ## requires at least two endpoints to calculate a cytotoxicity point. If
  ## the user wishes to use one assay, this function is not necessary.

  ## Changing 'default.pt' will change cyto_pt in the resulting data.table
  tcplCytoPt(aeid = 1:2, default.pt = 6)
}

## End(Not run)
```

tcpldbStats

Get summary statistics for the database

Description

tcpldbStats takes a string(type) and an optional parameter(val) to return the summary statistics on the entire tcpl database. When type = "all" the val is ignored. the function returns the number of distinct spid and aeids in the database at each level. When type = "aeid", the val parameter has to be a valid aeid in the database. The function returns a table consisting of the number of distinct spids at each level of processing for the aeid given in 'val'. When type = "spid", the val parameter has to be a valid spid in the database. The function returns a table consisting of the number of distinct aeids at each level of processing for the given spid in 'val'.

Usage

```
tcpldbStats(type = "all", val = NULL)
```

Arguments

type	String either "all", "aeid" or "spid"
val	integer if type = "aeid" , string if type = "spid"

tcplDefine*Load data dictionary descriptions*

Description

tcplDefine queries the tcpl databases and returns field descriptions from the data dictionary.

Usage

```
tcplDefine(val = NULL)
```

Arguments

val	The values to query on. Can be any combination of table names (to return all of its field descriptions) and field names
-----	---

Details

Short descriptions of fields for different tables are stored in a data dictionary. Query by table name to retrieve descriptions of each field in the given table, and/or query by field name to retrieve descriptions on every field with the given name, regardless of which table they belong to.

Value

A data.table with the data dictionary information for the given parameters.

Examples

```
## Store the current config settings, so they can be reloaded at the end
## of the examples
conf_store <- tcplConfList()
tcplConf(drvr = "example")

## Passing no parameters returns all of the fields described in the data
## dictionary
tcplDefine()

## Specifying table names of 'chemical' and 'sample' yields all of the
## fields from the 'chemical' and 'sample' tables
tcplDefine(c("chemical", "sample"))

## Specifying a field of 'wllt' yields all of the fields from any table that
## contains 'wllt' as a field
tcplDefine("wllt")

## Specifying a combination of table and field names results in all of the
## fields which are contained in the given tables and all of the given fields
## found in any table
tcplDefine(c("chemical", "spid", "wllt"))
```

```
## Reset configuration
options(conf_store)
```

tcplDelete	<i>Delete rows from tcpl databases</i>
------------	--

Description

tcplDelete deletes rows from the given table and database.

Usage

```
tcplDelete(tbl, fld, val, db)
```

Arguments

tbl	Character, length 1, the table to delete from
fld	Character, the field(s) to query on
val	List, vectors of values for each field to query on. Must be in the same order as 'fld'.
db	Character, the database containing the table

Note

This function is not exported and not intended to be used by the user.

See Also

[tcplSendQuery](#)

tcplFit	<i>Fit the data with the constant, hill, and gain-loss models</i>
---------	---

Description

tcplFit fits the constant, hill, and gain-loss models to the given data and returns some summary statistics and the fit parameters in a list.

Usage

```
tcplFit(
  logc,
  resp,
  bmad,
  force.fit = FALSE,
  bidirectional = FALSE,
  verbose = FALSE,
  ...
)
```

Arguments

logc	Numeric, log concentration values
resp	Numeric, normalized response values
bmad	Numeric, the baseline median absolute deviation for the entire assay
force.fit	Logical, TRUE indicates to attempt fitting every concentration series
bidirectional	Boolean If TRUE, bidirectional negative data before fitting (default=FALSE) The original version of the code required the data to start at small values and rise, so that negative curves had to be bidirectionalped outside the function, and TOP was always positive. Setting bidirectional to TRUE allows both rising and falling curves
verbose	Boolean If TRUE print warning messages
...	Any other data to be included in list output.

Details

when at least one median value is greater than $3 \times \text{bmad}$.

Value

List of summary values and fit parameters for the given data.

See Also

[tcplObjCnst](#), [tcplObjHill](#), [tcplObjGnls](#), [constrOptim](#)

Examples

```
logc <- 1:10
resp <- sapply(1:10, tcplHillVal, ga = 5, tp = 50, gw = 0.5)
params <- tcplFit(logc = logc, resp = resp, bmad = 10)
plot(resp ~ logc)
tcplAddModel(pars = params, modl = "hill")
```

tcplFit2	<i>tcpl Wrapper for tcplfit2_core including additional calculations to fit into new schema</i>
----------	--

Description

tcpl Wrapper for tcplfit2_core including additional calculations to fit into new schema

Usage

```
tcplFit2(
  dat,
  fitmodels = c("cnst", "hill", "gnls", "poly1", "poly2", "pow", "exp2", "exp3", "exp4",
    "exp5"),
  bmed = NULL
)
```

Arguments

dat	output from level 3 processing
fitmodels	list of the models that should be fit with the data
bmed	baseline value, typically should be 0

Value

Data.table with an additional column fitparams that includes all of the fitting parameters

tcplfit2_fun	<i>tcplfit2_fun Returns tcplfit2 function for drawing models given input</i>
--------------	--

Description

tcplfit2_fun Returns tcplfit2 function for drawing models given input

Usage

```
tcplfit2_fun(row, model, x)
```

Arguments

row	data table with all required conc/resp data
model	string of model to draw from tcplfit2
x	int vector of x values to calculate curve (y) values for

Value

a tcplfit2 function call

tcplFit2_nest	<i>Nest dataframe into a list that is readable by tcplfit2</i>
---------------	--

Description

Nest dataframe into a list that is readable by tcplfit2

Usage

```
tcplFit2_nest(dat)
```

Arguments

dat	a dataframe that has all of the fitting parameters in the style of tcplloaddata
-----	---

Value

a list of fitting parameters that can be consumed by tcplfit2

tcplFit2_unnest	<i>Unnest tcplfit2 parameters into a dataframe</i>
-----------------	--

Description

Unnest tcplfit2 parameters into a dataframe

Usage

```
tcplFit2_unnest(output)
```

Arguments

output	list of output from tcplfit2
--------	------------------------------

Value

list of parameters unnested and compiled into a dataframe

tcplGetAeid	<i>get Aeid for endpoint name</i>
-------------	-----------------------------------

Description

tcplGetAeid takes a string(name) and finds the assay component endpoint names that match the string and the auids associated with those names. The function performs a regular expression like matching for strings in the assay component endpoint name column in the assay component endpoint table.

Usage

```
tcplGetAeid(name)
```

Arguments

name	A string that will be matched to the assay component endpoint name
------	--

Examples

```
## Not run:
## Search for aenm (assay name) case insensitive
tcplGetAeid("TOX21")
tcplGetAeid("tox21")

## End(Not run)
```

tcplggplot2	<i>tcplggplot2</i>
-------------	--------------------

Description

tcplggplot2

Usage

```
tcplggplot2(
  dat,
  type = "mc",
  compare = "m4id",
  verbose = TRUE,
  flags = FALSE,
  yrange = c(NA, NA),
  group.fld = NULL,
  group.threshold = 9,
  hide_losing_models = FALSE
)
```

Arguments

<code>dat</code>	data table with all required conc/resp data; each row will extend comparison
<code>type</code>	string type of data that should be plotted 'sc' plotting - max medians plotted with hitcall 'mc' plotting - all fit models and winning model with hitcall
<code>compare</code>	Character vector, the field(s) samples were joined on for comparison
<code>verbose</code>	boolean should plotting include table of values next to the plot
<code>flags</code>	boolean should plotting include level 6 flags in plot caption
<code>yrange</code>	Integer of length 2, for overriding the y-axis range, <code>c(<min>,<max>)</code> . By default, <code>c(NA,NA)</code> .
<code>group.fld</code>	Character, column name to group curves by when number of comparison curves exceeds <code>group.threshold</code> . By default 'modl' for MC and 'hitc' for SC.
<code>group.threshold</code>	Integer of length 1, minimum number of samples in a given plot where comparison plots, instead of coloring models by sample, should delineate curve color by a given <code>group.fld</code> . By default 9.
<code>hide_losing_models</code>	Boolean, by default FALSE. For individual mc plots only, should the losing models be hidden?

Value

A ggplot object or grob with accompanied table depending on verbose option

 tcplHit2

Hitcalling with tcplfit2

Description

Hitcalling with tcplfit2

Usage

```
tcplHit2(mc4, coff)
```

Arguments

<code>mc4</code>	data.table with level 4 data
<code>coff</code>	cutoff value for hitcalling

Value

Data.table with key value pairs of hitcalling parameters

tcplLegacyPlot	<i>tcplLegacyPlot</i>
----------------	-----------------------

Description

tcplLegacyPlot

Usage

tcplLegacyPlot()

Value

a ggplot based on old plotting methodology

tcplListFlds	<i>Load the field names for a table</i>
--------------	---

Description

tcplListFlds loads the column names for the given table and database.

Usage

tcplListFlds(tbl, db = getOption("TCPL_DB"))

Arguments

tbl	Character of length 1, the tcpl database table
db	Character of length 1, the tcpl database

Details

This function can be particularly useful in defining the 'fld' param in the tcplLoad- functions.

Value

A string of field names for the given table.

Examples

```
## Not run:
## Gives the fields in the mc1 table
tcplListFlds("mc1")

## End(Not run)
```

tcplLoadChem	<i>Load sample/chemical information</i>
--------------	---

Description

tcplLoadChem queries the tcpl database and returns the chemical information for the given field and values.

Usage

```
tcplLoadChem(field = NULL, val = NULL, exact = TRUE, include.spid = TRUE)
```

Arguments

field	Character of length 1, the field to query on
val	Vector of values to subset on
exact	Logical, should chemical names be considered exact?
include.spid	Logical, should spid be included?

Details

The 'field' parameter is named differently from the 'fld' parameter seen in other functions because it only takes one input.

In the MySQL environment the user should be able to give partial chemical name strings, to find chemicals with similar names. For example, setting 'val' to "phenol" when 'field' is "chnm" and 'exact' is FALSE might pull up the chemicals "Bisphenol A" and "4-Butylphenol". More technically, setting 'exact' to FALSE passes the string in 'val' to an RLIKE statement within the MySQL query.

Value

A data.table with the chemical information for the given parameters

Examples

```
## Not run:
## Passing no parameters gives all of the registered chemicals with their
## sample IDs
tcplLoadChem()

## Or the user can exclude spid and get a unique list of chemicals
tcplLoadChem(include.spid = FALSE)

## In addition, the user can retrieve only the registered chemicals from the chemical table
tcplLoadChem(field = 'chem.only')

## Other examples:
tcplLoadChem(field = "chnm", val = "Bisphenol A")
```

```
tcplLoadChem(field = "chid", val = 20182)

## End(Not run)
```

tcplLoadChemList	<i>Load chemical list information</i>
------------------	---------------------------------------

Description

tcplLoadChemList queries the tcpl databases and returns information about the chemical lists.

Usage

```
tcplLoadChemList(field = NULL, val = NULL)
```

Arguments

field	Character of length 1, 'chid', 'dsstox_substance_id' or 'list_acronym', whether to search by chemical id (chid), dsstox_substance_id, or list_acronym
val	The values to query on

Details

Chemicals are stored in different lists by chemical ID. Therefore, it is not possible to delineate samples with the same chemical ID into two distinct chemical lists. However, it is possible for a chemical ID to belong to more than one (or no) chemical lists.

When chemicals belong to more than one list, the chemical is listed multiple times (one for each distinct list).

Value

A data.table with the chemical list information for the given parameters.

Examples

```
## Not run:
## Passing no parameters gives all of the chemical IDs that have a chemical
## list registered
clist <- tcplLoadChemList()

## Notice there are different number of rows in tcplLoadChemList than in tcplLoadChem,
## indicating some chemicals must belong to more than list (or no lists).
chem <- tcplLoadChem(include.spid = TRUE)
nrow(chem)
nrow(clist)
```

```
## Show the unique chemical lists
clist[ , unique(list_acronym)]

## Specifying a chemical list will not show what other libraries a
## chemical might belong to.
tcplLoadChemList(field = "list_acronym", val = "CPDBAS")
tcplLoadChemList(field = "chid", val = 20182)
tcplLoadChemList(field = "dsstox_substance_id", val = "DTXSID7020182")

## End(Not run)
```

tcplLoadConcUnit	<i>Load concentration units for assay endpoints</i>
------------------	---

Description

tcplLoadUnit queries the tcpl databases and returns a data.table with the concentration units for the given assay endpoint ids (spid).

Usage

```
tcplLoadConcUnit(spid)
```

Arguments

spid	Integer, assay endpoint ids
------	-----------------------------

Value

A data.table containing level 3 correction methods for the given spids.

See Also

[tcplQuery](#), [data.table](#)

tcplLoadData	<i>Load tcpl data</i>
--------------	-----------------------

Description

tcplLoadData queries the tcpl databases and returns a data.table with data for the given level and data type.

Usage

```
tcplLoadData(
  lvl,
  fld = NULL,
  val = NULL,
  type = "mc",
  add.fld = TRUE,
  exact = TRUE
)
```

Arguments

lvl	Integer of length 1, the level of data to load
fld	Character, the field(s) to query on
val	List, vectors of values for each field to query on. Must be in the same order as 'fld'.
type	Character of length 1, the data type, "sc" or "mc"
add.fld	Boolean if true we want to return the additional parameters fit with tcplfit2
exact	Logical, passed to tcplLoadChem – should chemical names be considered exact?

Details

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the 'mc' tables, whereas the single concentration will be loaded into the 'sc' tables.

Setting 'lvl' to "agg" will return an aggregate table containing the m4id with the concentration-response data and m3id to map back to well-level information.

If tcplConf() was set with "API" as the driver, then tcplLoadData will return data from the CCTE Bioactivity API. API data is available for type = 'mc' and lvl = c(3,4,5,6) and 'agg'. Only fields relating to the requested level are returned, but not all fields that usually return from invitrodb are available from the API. To have all fields available from the API return, regardless of what lvl is set to, set add.fld to TRUE. API query-able fields include "aeid", "spid", "m4id", and "dtxsid".

Leaving fld NULL will return all data.

Valid fld inputs are based on the data level and type:

type	lvl	Queried tables
sc	0	sc0
sc	1	sc0, sc1
sc	agg	sc1, sc2_agg
sc	2	sc2
mc	0	mc0
mc	1	mc0, mc1
mc	2	mc0, mc1, mc2
mc	3	mc0, mc1, mc3
mc	agg	mc3, mc4_agg

mc	4	mc4
mc	5	mc4, mc5
mc	6	mc4, mc6
mc	7	mc4, mc7

Value

A data.table containing data for the given fields.

See Also

[tcplQuery](#), [data.table](#)

Examples

```
## Not run:
## Load all of level 0 for multiple-concentration data, note 'mc' is the
## default value for type
tcplLoadData(lvl = 0)

## Load all of level 1 for single-concentration
tcplLoadData(lvl = 1, type = "sc")

## List the fields available for level 1, coming from tables mc0 and mc1
tcplListFlds(tbl = "mc0")
tcplListFlds(tbl = "mc1")

## Load level 0 data where the well type is "t" and the concentration
## index is 3 or 4
tcplLoadData(lvl = 1, fld = c("wllt", "cndx"), val = list("t", c(3:4)))

## Load level 4 data using a chemical name
tcplLoadData(lvl = 4, fld = "chnm", val = "Bisphenol A")

## Load level 3 data using a partial chemical name
tcplLoadData(lvl = 3, fld = "chnm", val = "phenol", exact = FALSE)

## End(Not run)
```

tcplLoadUnit

Load response units for assay endpoints

Description

tcplLoadUnit queries the tcpl databases and returns a data.table with the response units for the given assay endpoint ids (aeid).

Usage

```
tcplLoadUnit(aeid)
```

Arguments

aeid Integer, assay endpoint ids

Value

A data.table containing level 3 correction methods for the given aeid.

See Also

[tcplQuery](#), [data.table](#)

tcplLvlCount	<i>Load tcpl level counts</i>
--------------	-------------------------------

Description

tcplLvlCount queries the tcpl databases and returns a data frame with count totals for the given levels and data type.

Usage

```
tcplLvlCount(lvls = NULL, type = "mc")
```

Arguments

lvls Integer or list of Integers, The levels of data to load
type Character of length 1, the data type, "sc" or "mc"

Details

The data type can be either 'mc' for mutiple concentration data, or 'sc' for single concentration data.
Leaving lvls NULL will return all data.

Value

A data.table containing data for the given fields.

See Also

[tcplQuery](#), [data.table](#)

Examples

```
## Not run:
## Get all counts for level 1 for multiple-concentration
tcplLvlCount(lvls = 1)

## Get all counts for levels 4 through 7 for multiple-concentration
tcplLvlCount(lvls = 4:7)

## Get all counts for multiple-concentration data, note 'mc' is the
## default value for type
tcplLvlCount()

## End(Not run)
```

tcplMakeAeidMultiPlts *Create a .pdf with all dose-response plots for a given aeid, 6 per page*

Description

tcplMakeAeidMultiPlts Create a .pdf with all dose-response plots for a given aeid

Usage

```
tcplMakeAeidMultiPlts(
  aeid,
  lvl = 4L,
  fname = NULL,
  odir = getwd(),
  hitc.all = TRUE
)
```

Arguments

aeid	Integer of length 1, the assay endpoint id
lvl	Integer of length 1, the data level to use (4-7)
fname	Character, the filename
odir	The directory to save the .pdf file in
hitc.all	If FALSE, only plots with hitc==1 will be displayed

Details

tcplMakeAeidMultiPlts provides a wrapper for [tcplMultiplot](#), allowing the user to produce PDFs with the curve plots without having to separately load all of the data and establish the PDF device.

If 'fname' is NULL, a default name is given by concatenating together assay information.

tcplMakeAeidPlts

Create a .pdf with dose-response plots

Description

tcplMakeAeidPlts creates a .pdf file with the dose-response plots for the given aeid.

Usage

```
tcplMakeAeidPlts(
  aeid,
  compare = F,
  lvl = 4L,
  fname = NULL,
  odir = getwd(),
  ordr.fitc = TRUE,
  cnst = NULL
)
```

Arguments

aeid	Integer of length 1 or 2, the assay endpoint id
compare	Boolean to for comparison of aeids if length(aeid)>1
lvl	Integer of length 1, the data level to use (4-7). Only level 5-6 valid for compare aeids.
fname	Character, the filename
odir	The directory to save the .pdf file in
ordr.fitc	Logical, should the fits be ordered by fit category?
cnst	Constant hline to draw on plot

Details

tcplMakeAeidPlts provides a wrapper for [tcplPlotFits](#), allowing the user to produce PDFs with the curve plots without having to separately load all of the data and establish the PDF device.

If 'fname' is NULL, a default name is given by concatenating together assay information.

Note, the default value for ordr.fitc is TRUE in tcplMakeAeidPlts, but FALSE in tcplPlotFits

Note, only level 5 or level 6 is valid for comparing 2 aeids.

Examples

```
## Not run:
## Will produce the same result as the example for tcplPlotFits
tcplMakeAeidPlts(aeid = 1, lvl = 6, ordr.fitc = FALSE)

## End(Not run)
```



```
## Not run:  
## Compare two aeids on same plots  
tcplMakeAeidPlts(aeid = c(1,2), compare=T, lvl = 6)  
  
## End(Not run)
```

tcplMakeChidMultiPlts *Create a .pdf with all dose-response plots for a given chid, 6 per page*

Description

tcplMakeChidMultiPlts Create a .pdf with all dose-response plots for a given chid

Usage

```
tcplMakeChidMultiPlts(  
  chid,  
  lvl = 4L,  
  fname = NULL,  
  odir = getwd(),  
  hitc.all = TRUE  
)
```

Arguments

chid	Integer of length 1, the chemical id
lvl	Integer of length 1, the data level to use (4-7)
fname	Character, the filename
odir	The directory to save the .pdf file in
hitc.all	If FALSE, only plots with hitc==1 will be displayed

Details

tcplMakeChidMultiPlts provides a wrapper for [tcplMultiplot](#), allowing the user to produce PDFs with the curve plots without having to separately load all of the data and establish the PDF device.

If 'fname' is NULL, a default name is given by concatenating together assay information.

tcplMultiplot	<i>Plot summary fits based on fit and dose-response data</i>
---------------	--

Description

tcplMultiplot takes the dose-response and fit data and produces summary plot figures.

Usage

```
tcplMultiplot(dat, agg, flg = NULL, boot = NULL, browse = FALSE, hitc.all)
```

Arguments

dat	data.table, level 4 or level 5 data, see details.
agg	data.table, concentration-response aggregate data, see details.
flg	data.table, level 6 data, see details.
boot	data.table, level 7 data, see details.
browse	Logical, should browser() be called after every plot?
hitc.all	Logical, if FALSE, only plots with hitc==1 will be displayed

Details

The data for 'dat', 'agg', and 'flg' should be loaded using the [tcplLoadData](#) function with the appropriate 'lvl' parameter. See help page for tcplLoadData for more information.

If dat contains only one aeid, plots will be ordered by chemical name (chnm). Otherwise, plots are ordered by assay endpoint name (aenm). ## While it is most likely the user will want to just save all of the plots ## to view in a PDF, the 'browse' parameter can be used to quickly view ## some plots.

tcplPlot	<i>Generic Plotting Function for tcpl</i>
----------	---

Description

tcplPlot queries the tcpl databases and returns or saves customizable plots for the given data type, field(s), and value(s).

Usage

```
tcplPlot(
  dat = NULL,
  type = "mc",
  fld = "m4id",
  val = NULL,
  compare = "m4id",
  by = NULL,
  output = c("ggplot", "console", "pdf", "png", "jpg", "svg", "tiff"),
  fileprefix = paste0("tcplPlot_", Sys.Date()),
  multi = NULL,
  verbose = TRUE,
  nrow = NULL,
  ncol = NULL,
  dpi = 600,
  flags = FALSE,
  yuniform = FALSE,
  yrange = c(NA, NA),
  group.fld = NULL,
  group.threshold = 9,
  hide_losing_models = FALSE
)
```

Arguments

<code>dat</code>	data.table or list of data.tables containing plot-prepared data, not required. Used for stand-alone (ToxCast or other tcplfit2-fit data) plotting or advanced plotting (generating comparison plots across multiple database configurations). Pass a data.table for default behavior, which will split data by 'compare'. Pass a list of data.tables to directly specify the groupings for comparison plots where each list item (data.table) will be printed on a single plot. See tcplPlotLoadData.
<code>type</code>	Character of length 1, the data type, "sc" or "mc".
<code>fld</code>	Character vector, the field(s) to query on.
<code>val</code>	List containing vectors of values for each field to query on. Must be in the same order as 'fld'.
<code>compare</code>	Character vector, the field(s) to join samples on to create comparison plots. By default "m4id", "s2id" if 'type' = "sc". As every endpoint-sample will always have its own m4id, this will create individual plots. To create a comparison plot across the same chemicals, use a chemical identifier like "dsstox_substance_id". Likewise, to create a comparison plot across the same sample ids, use "spid". Use "aeid" to create a comparison plot across the same assay component endpoints, which will likely trigger the large compare plot style; for more info see 'group.threshold'. To use a custom field to create comparison, 'dat' should be supplied as a data.table generated from tcplPlotLoadData with the custom column added. If 'dat' is instead a list of data.tables, setting 'compare' will be ignored in favor of the list groups.
<code>by</code>	Parameter to divide files into e.g. "aeid".

output	How should the plot be presented. To work with the plot in environment, use "ggplot"; to interact with the plot in application, use "console"; or to save as a file type, use "pdf", "jpg", "png", "svg", or "tiff".
fileprefix	Prefix of file when saving.
multi	Boolean, by default TRUE for "pdf" if the number of plots exceeds one. Prints variable number of plots per page depending on 'verbose' and 'type' settings.
verbose	Boolean, by default TRUE. If TRUE, a table with fitting parameters is included with the plot. To return or save simple plot, set to FALSE. In comparison plotting, verbose as TRUE will also return annotation information in a table, which is hidden using verbose = FALSE.
nrow	Integer, number of rows in multiplot. By default 2.
ncol	Integer, number of columns in multiplot. By default 3, 2 if verbose, 1 for verbose compare plots.
dpi	Integer, image print resolution. By default 600.
flags	Boolean, by default FALSE. If TRUE, level 6 flags are displayed within output.
yuniform	Boolean, by default FALSE. If TRUE, all plots will have uniform y axis scaling, automatically determined.
yrange	Integer of length 2, for directly setting the y-axis range, c(<min>,<max>). By default, c(NA,NA). Plots containing points or curves outside this range will be expanded to fit.
group.fld	Character, column name to group curves by when number of comparison curves exceeds group.threshold. By default 'modl' for MC and 'hite' for SC.
group.threshold	Integer of length 1, minimum number of samples in a given plot where comparison plots, instead of coloring models by sample, should delineate curve color by a given group.fld. By default 9.
hide_losing_models	Boolean, by default FALSE. For individual mc plots only, should the losing models be hidden?

Details

The data 'type' can be either "mc" for multiple concentration data, or "sc" for single concentration data. 'fld' can be any field(s) available at or before level 5 for 'mc' or level 2 for 'sc'. 'val' must be contain values for each 'fld'. If 'fld' is just length 1, 'val' can be a regular number or character vector of any length. If 'fld' is greater than length 1, 'val' must be a list where each item matches each 'fld'.

Use 'dat' to supply custom tcplfit2 data or pre-loaded (and manipulated) tcpl pipe-lined data from tcplPlotLoadData for more flexibility. 'dat' must contain all columns required by tcplPlot by the specified type, such as sample info, chemical info, assay endpoint info, all conc/resp data, all level 4 model fitting parameters, all level 5 hitcall and potency values, and level 6 cautionary flags if 'flags' = TRUE. 'dat' can be used to add custom columns for use with the 'compare', 'by', or 'group.fld' parameters.

There are three plotting styles tcplPlot provides. First, individual concentration response plots, by sample (more specifically, the distinct "m4id"), which contain response points, winning and losing

models, cut-off lines, AC50 lines, BMD/BMR lines, and a potency table. Second are traditional compare plots, by default generated when the number of samples on one plot is between 2 and 8 inclusive. These plots contain response points, winning models, and cut-offs, all color-delineated by sample ("m4id"). It also contains tables for viewing similar and differing annotation info as well as hit-calls, BMDs, and AC50s. Finally are tcplPlot's comparison plots for large comparisons, greater than 8 curves by default. These plots strip the annotation tables, response points, and curve color grouping by sample in favor of a more holistic view which emphasizes the winning models altogether grouped for color by 'group.fld'. Setting 'flags' = TRUE here provides a view of the flag counts across the entire plot's samples.

Examples

```
## Not run:
# Requires a database or API connection
tcplConfDefault()

# Simple plot returned as ggplot object for viewing or manipulating
tcplPlot(fld = "m4id", val = 10000000)

# Simple single conc plot returned as ggplot object for viewing or manipulating
# Every example below works for single conc, as long as type = "sc" and any use
# of "m4id" is replaced with "s2id". Single conc not supported under API connection.
tcplPlot(type = "sc", fld = "s2id", val = 6500000)

# Simple plot returned to console using plotly package
tcplPlot(fld = "m4id", val = 10000000, output = "console")

# Simple plot saved to image file
tcplPlot(fld = "m4id", val = 10000000, output = "png")

# Simple plot saved to pdf
tcplPlot(fld = "m4id", val = 10000000, output = "pdf", multi = FALSE)

# Multiple plots saved to pdf, by assay endpoint
tcplPlot(fld = "aeid", val = 1750, output = "pdf")

# Multiple plots by assay endpoint saved to pdf, specifying a file name prefix
tcplPlot(fld = "aeid", val = 1750, output = "pdf", fileprefix = "example")

# Multiple plots by assay endpoint saved to pdf, adjusting the number of rows and columns
tcplPlot(fld = "aeid", val = 1750, output = "pdf", nrow = 3, ncol = 4)

# Multiple plots saved to pdf, split into files by aeid
tcplPlot(fld = "aeid", val = c(1746, 1748, 1750, 1752, 1754), by = "aeid", output = "pdf")

# To use more than one field, be sure 'val' is given as a list with an element for each 'fld'.
# Note - using more than one field is not supported under an API connection.
tcplPlot(fld = c("aeid", "spid"), val = list(1750, c("WaterSample3", "WaterSample20")),
        output = "pdf")

# Simple plot saved to image file, without verbose table
tcplPlot(fld = "m4id", val = 10000000, output = "png", verbose = FALSE)
```

```

# Simple plot saved to image file, with level 6 cautionary flags
tcplPlot(fld = "m4id", val = 10000000, output = "png", flags = TRUE)

# Simple plot saved to image file, with decreased dpi
tcplPlot(fld = "m4id", val = 10000000, output = "png", dpi = 200)

# Multiple plots by assay endpoint saved to pdf, with uniform y-range across all plots
tcplPlot(fld = "aeid", val = 1750, output = "pdf", yuniform = TRUE)

# Multiple plots by assay endpoint saved to pdf, with specific uniform y-range across
# all plots. Any plots with elements outside this range will be expanded to fit.
tcplPlot(fld = "aeid", val = 1750, output = "pdf", yrange = c(-1,2))

# Two plots created by comparing all curves across one assay endpoint saved to pdf.
# "conc_unit" included since this endpoint contains multiple ("uM" and "CF"), so we
# should split them up.
tcplPlot(fld = "aeid", val = 1750, compare = c("aeid", "conc_unit"), output = "pdf")

# Change group.fld to alter the binning field for curve color in this large comparison plot
tcplPlot(fld = "aeid", val = 1750, compare = c("aeid", "conc_unit"), output = "pdf",
  group.fld = "fitc")

# If you'd rather not have curves binned by 'group.fld', set group.threshold to a
# value greater than the greatest number of curves contained on one plot.
# NOTE - it is not recommended to do this for plots with more than 10-12 curves.
tcplPlot(fld = "aeid", val = 1750, compare = c("aeid", "conc_unit"), output = "pdf",
  group.threshold = 25)

# Multiple plots created by comparing all spids across five assay endpoints saved to pdf
tcplPlot(fld = "aeid", val = c(1746, 1748, 1750, 1752, 1754), compare = "spid", output = "pdf")

# Multiple plots created by comparing all spids across five assay endpoints saved to pdf,
# split into files by "conc_unit"
tcplPlot(fld = "aeid", val = c(1746, 1748, 1750, 1752, 1754), compare = "spid",
  by = "conc_unit", output = "pdf")

# Multiple plots created by using a custom field to compare across five assay
# endpoints saved to pdf.
dat <- tcplPlotLoadData(fld = "aeid", val = c(1746, 1748, 1750, 1752, 1754), flags = TRUE)
dat$cmp.fld <- rep("", nrow(dat)) # some logic for setting column data
tcplPlot(dat = dat, compare = "cmp.fld", output = "pdf")

# Multiple plots created by using a custom binning field to compare across five
# assay endpoints saved to pdf.
dat <- tcplPlotLoadData(fld = "aeid", val = c(1746, 1748, 1750, 1752, 1754), flags = TRUE)
dat$grp.fld <- rep("", nrow(dat)) # some logic for setting column data
tcplPlot(dat = dat, compare = c("aeid", "conc_unit"), output = "pdf", group.fld = "grp.fld")

# Multiple plots created by supplying dat as a list of data.tables.
dat <- tcplPlotLoadData(fld = "aeid", val = c(1746, 1748, 1750, 1752, 1754), flags = TRUE)
dat <- split(dat, by = "conc_unit") # or likely some more complex logic for splitting
tcplPlot(dat = dat, output = "pdf")

```

```
## End(Not run)
```

```
tcplPlotCalcAspectRatio
      tcplPlotCalcAspectRatio
```

Description

tcplPlotCalcAspectRatio

Usage

```
tcplPlotCalcAspectRatio(
  type = "mc",
  verbose = FALSE,
  multi = FALSE,
  nrows = 0,
  output = c("ggplot", "console", "pdf", "png", "jpg", "svg", "tiff"),
  group.threshold = 9,
  nrow = NULL,
  ncol = NULL,
  flags = FALSE
)
```

Arguments

type	string of mc or sc indicating if it is single or multi conc
verbose	should the plot return a table with parameters
multi	Boolean, by default TRUE for "pdf" if the number of plots exceeds one. Prints variable number of plots per page depending on 'verbose' and 'type' settings.
nrows	Integer, number of rows each compare plot uses
output	How should the plot be presented. To work with the plot in environment, use "ggplot"; to interact with the plot in application, use "console"; or to save as a file type, use "pdf", "jpg", "png", "svg", or "tiff".
group.threshold	Integer of length 1, minimum number of samples in a given plot where comparison plots, instead of coloring models by sample, should delineate curve color by a given group.fld. By default 9.
nrow	Integer, number of rows in multiplot. By default 2.
ncol	Integer, number of columns in multiplot. By default 3, 2 if verbose, 1 for compare plots.
flags	Boolean, by default FALSE. If TRUE, level 6 flags are displayed within output.

Value

a list of validated parameters for plotting

tcplPlotFitc	<i>Plot the fit category tree</i>
--------------	-----------------------------------

Description

tcplPlotFitc makes a plot showing the level 5 fit categories.

Usage

```
tcplPlotFitc(fitc = NULL, main = NULL, fitc_sub = NULL)
```

Arguments

fitc	Integer, the fit categories
main	Character of length 1, the title (optional)
fitc_sub	Integer, a subset of fit categories to plot

Note

Suggested device size (inches): width = 10, height = 7.5, pointsize = 9

Examples

```
## Not run:
## Plot visualization of fit categories for all level 5 data
tcplPlotFitc(fitc = tcplLoadData(5)$fitc)

## End(Not run)
```

tcplPlotFits	<i>Plot summary fits based on fit and dose-response data</i>
--------------	--

Description

tcplPlotFits takes the dose-response and fit data and produces summary plot figures.

Usage

```
tcplPlotFits(
  dat,
  agg,
  flg = NULL,
  boot = NULL,
  ordr.fitc = FALSE,
  browse = FALSE,
```



```

    cnst = NULL,
    orig.aeid = NULL,
    compare = F
  )

```

Arguments

<code>dat</code>	data.table, level 4 or level 5 data, see details.
<code>agg</code>	data.table, concentration-response aggregate data, see details.
<code>flg</code>	data.table, level 6 data, see details.
<code>boot</code>	data.table, level 7 data, see details.
<code>ordr.fitc</code>	Logical, should the fits be ordered by fit category?
<code>browse</code>	Logical, should <code>browser()</code> be called after every plot?
<code>cnst</code>	Constant hline to draw on plot
<code>orig.aeid</code>	Original aeid list from <code>tcplMakeAeidPlts</code> to maintain order
<code>compare</code>	boolean to determine if aeids should be compared on same plot

Details

The data for 'dat', 'agg', and 'flg' should be loaded using the [tcplLoadData](#) function with the appropriate 'lvl' parameter. See help page for `tcplLoadData` for more information.

Supplying level 4 data for the 'dat' parameter will result in level 4 plots. Similarly, supp

If fits are not ordered by fit category, they will be ordered by chemical ID. Inputs with multiple assay endpoints will first be ordered by assay endpoint ID.

Examples

```

## Not run:
## tcplPlotFits needs data.tables supplying the concentration/response
## data stored in mc4_agg, as well as the fit information from mc4 or mc5.
## Additionally, tcplPlotFits can take level 6 data from mc6 and add the
## flag information to the plots. The following shows how to make level 5
## plots. Adding the 'flg' parameter would result in level 6 plots, and
## loading level 4, rather than level 5 data, would result in level 4 plots.

l5 <- tcplLoadData(lvl = 5, fld = "m4id", val = 18609966)
l4_agg <- tcplLoadData(lvl = "agg", fld = "m4id", val = 18609966)

pdf(file = "tcplPlotFits.pdf", height = 6, width = 10, pointsize = 10)
tcplPlotFits(dat = l5, agg = l4_agg)
graphics.off()

## While it is most likely the user will want to just save all of the plots
## to view in a PDF, the 'browse' parameter can be used to quickly view
## some plots.

## Start by identifying some sample IDs to plot, then call tcplPlotFits with
## a subset of the data. This browse function is admittedly clunky.

```

```

bpa <- tcplLoadChem(field = "chnm", val = "Bisphenol A")[, spid]
l5_sub <- l5[spid %in% bpa]
tcplPlotFits(dat = l5_sub,
             agg = l4_agg[m4id %in% l5_sub$m4id],
             browse = TRUE)

## End(Not run)

```

tcplPlotLoadData	<i>Utility function to load data for tcplPlot</i>
------------------	---

Description

tcplPlotLoadData queries the tcpl databases and returns a data.table with data for the given field, value, level, and data type prepared in a format tcplPlot can use to generate plots.

Usage

```
tcplPlotLoadData(type = "mc", fld = "m4id", val, flags = FALSE)
```

Arguments

type	Character of length 1, the data type, "sc" or "mc"
fld	Character, the field(s) to query on.
val	List, vectors of values for each field to query on. Must be in the same order as 'fld'.
flags	Boolean, by default FALSE. If TRUE, level 6 flags are loaded for use in tcplPlot. Must be set to TRUE if tcplPlot 'flags' also is/will be set to TRUE

Details

This utility function is used by tcplPlot to load and prepare data from tcplLoadData for use in generating plots. It is exported for use in advanced comparison plots where users create plots using multiple data sources. After saving the response from tcplPlotLoadData, switch data source config and pass the data to tcplPlot dat parameter.

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data.

Value

A data.table containing plot-ready data for the given fields.

See Also

[tcplPlot](#)

Examples

```
## Not run:
## load mc plot data for an entire endpoint
dat <- tcplPlotLoadData(fld = "aeid", val = 703)

## load sc plot data for an entire endpoint
dat <- tcplPlotLoadData(type = "sc", fld = "aeid", val = 703)

## load plot data for two endpoint-samples and include loading of flags
## flags must equal TRUE if tcplPlot will/does
dat <- tcplPlotLoadData(fld = c("spid", "aeid"),
                        val = list(c("TP0000269F11", "TP0000395A09"),703),
                        flags = TRUE)

## if desired, switch connections
tcplConf()

## use dat in tcplPlot
tcplPlot(dat = dat,
         fld = c("spid", "aeid"),
         val = list(c("TP0000269F11", "TP0000395A09"),703),
         compare.val = list(c("LEGTV002B01", "LEGTV003A06"),703),
         output = "pdf",
         flags = TRUE,
         fileprefix="example")

## End(Not run)
```

tcplPlotLoadWllt	<i>tcplPlotLoadWllt Replaces NA dtxsid and chnm with a string description of the sample's well type(s)</i>
------------------	--

Description

tcplPlotLoadWllt Replaces NA dtxsid and chnm with a string description of the sample's well type(s)

Usage

```
tcplPlotLoadWllt(dat = NULL, type = "mc")
```

Arguments

dat	dataset
type	mc or sc

Value

dat with updated dtxsid/chnm if they are NA

tcplPlotlyPlot	<i>tcplPlotlyPlot</i>
----------------	-----------------------

Description

tcplPlotlyPlot

Usage

tcplPlotlyPlot(dat, lvl = 5, hide_losing_models = FALSE)

Arguments

- dat data table with all required conc/resp data
- lvl integer level of data that should be plotted
- hide_losing_models Boolean, by default FALSE. For individual mc plots only, should the losing models be hidden? level 2 - for 'sc' plotting level 5 - for 'mc' plotting, all fit models and winning model with hitcall

Value

A plotly plot

tcplPlotM4ID	<i>Plot fit summary plot by m4id</i>
--------------	--------------------------------------

Description

tcplPlotM4ID creates a summary plots for the given m4id(s) by loading the appropriate data from the tcpl databases and sending it to [tcplPlotFits](#)

Usage

tcplPlotM4ID(m4id, lvl = 4L)

Arguments

- m4id Integer, m4id(s) to plot
- lvl Integer, the level of data to plot

Details

A level 4 plot ('lvl' = 4) will plot the concentration series and the applicable curves, without an indication of the activity call or the winning model. Level 4 plots can be created without having done subsequent processing.

Level 5 plots include the level 4 information with the activity call and model selection. The winning model will be highlighted red in the side panel containing the summary statistics. Level 6 plots, in addition the all of the level 4 and 5 information, include the positive flag IDs. If the flag has an associated value, the value will be in parentheses following the flag ID. Level 7 plots in addition to all of the level 4, 5, and 6 information, include the AC50 confidence interval and hit percentage information from bootstrapping.

See Also

[tcplPlotFits](#), [tcplMakeAeidPlts](#)

Examples

```
## Not run:
tcplPlotM4ID(m4id = 18609966, lvl = 4) ## Create a level 4 plot
tcplPlotM4ID(m4id = 18609966, lvl = 5) ## Create a level 5 plot
tcplPlotM4ID(m4id = 18609966, lvl = 6) ## Create a level 6 plot

## End(Not run)
```

tcplPlotPlate	<i>Plot plate heatmap</i>
---------------	---------------------------

Description

tcplPlotPlate generates a heatmap of assay plate data

Usage

```
tcplPlotPlate(dat, apid, id = NULL, quant = c(0.001, 0.999))
```

Arguments

dat	data.table containing tcpl data
apid	Character of length 1, the apid to plot
id	Integer of length 1, the assay component id (acid) or assay endpoint id (aeid), depending on level. Only need to specify for multiplexed assays when more than one acid/aeid share an apid.
quant	Numeric vector, the range of data to include in the legend

Details

The legend represents the range of the data supplied to `dat`, for the applicable ID. The additional horizontal lines on the legend indicate the range of the plotted plate, to show the relation of the plate to the assay as a whole. A plot with a legend specific for the given apid can be created by only supplying the data for the apid of interest to `'dat'`.

The `quant` parameter, by default including 99.8 allows for extreme outliers without losing resolution. Outliers in either direction will be highlighted with a dark ring, as seen in the example. A NULL value for `'quant'` will not restrict the data at all, and will use the full range for the legend.

Wells with a well quality of 0 (only applicable for level 1 plots), will have an "X" through their center.

Note

For the optimal output size, use `width = 10`, `height = 10*(2/3)`, `pointsize = 10`, `units = "in"`

Examples

```
## Not run:
d1 <- tcplLoadData(lvl = 1, fld = "acid", val = 1)
tcplPlotPlate(dat = d1, apid = "09Apr2014.Plate.17")

## End(Not run)
```

<code>tcplPlotSetYRange</code>	<i>tcplPlotSetYRange</i>
--------------------------------	--------------------------

Description

`tcplPlotSetYRange`

Usage

```
tcplPlotSetYRange(dat, yuniform, yrange, type)
```

Arguments

<code>dat</code>	dataset
<code>yuniform</code>	should the yrange be uniform
<code>yrange</code>	length 2 of the yrange
<code>type</code>	mc or sc

Value

yrange of the data

tcplPlotValidate	<i>tcplPlotValidate</i>
------------------	-------------------------

Description

tcplPlotValidate

Usage

```
tcplPlotValidate(  
  dat = NULL,  
  type = "mc",  
  compare = "m4id",  
  by = NULL,  
  flags = NULL,  
  output = c("ggplot", "console", "pdf", "png", "jpg", "svg", "tiff"),  
  multi = NULL,  
  verbose = FALSE  
)
```

Arguments

dat	data.table containing plot-prepared data
type	string of mc or sc indicating if it is single or multi conc
compare	Character vector, the field(s) to join samples on to create comparison plots
by	Parameter to divide files into e.g. "aeid".
flags	bool - should we return flags
output	how should the plot be formatted
multi	are there multiple plots
verbose	should the plot return a table with parameters

Value

a list of validated parameters for plotting

tcplPrepOtppt*Map assay/chemical ID values to annotation information*

Description

tcplPrepOtppt queries the chemical and assay information from the tcpl database, and maps the annotation information to the given data.

Usage

```
tcplPrepOtppt(dat, ids = NULL)
```

Arguments

dat	data.table, output from tcplLoadData
ids	Character, (optional) a subset of ID fields to map

Details

tcplPrepOtppt is used to map chemical and assay identifiers to their respective names and annotation information to create a human-readable table that is more suitable for an export/output.

By default the function will map sample ID (spid), assay component id (acid), and assay endpoint ID (aeid) values. However, if 'ids' is not null, the function will only attempt to map the ID fields given by 'ids.'

Value

The given data.table with chemical and assay information mapped

Examples

```
## Not run:
## Load some example data
d1 <- tcplLoadData(1)

## Check for chemical name in 'dat'
"chnm" %in% names(d1) ## FALSE

#' ## Map all annotations
d2 <- tcplPrepOtppt(d1) ##
"chnm" %in% names(d2) ## TRUE
"acnm" %in% names(d2) ## TRUE

## Map chemical annotation only
d3 <- tcplPrepOtppt(d1, ids = "spid")
"chnm" %in% names(d3) ## TRUE
"acnm" %in% names(d3) ## FALSE
```



```
## End(Not run)
```

tcplRun	<i>Perform data processing</i>
---------	--------------------------------

Description

tcplRun is the function for performing the data processing, for both single-concentration and multiple-concentration formats.

Usage

```
tcplRun(  
  asid = NULL,  
  slvl,  
  elvl,  
  id = NULL,  
  type = "mc",  
  mc.cores = NULL,  
  outfile = NULL,  
  runname = NULL,  
  ready_only = FALSE  
)
```

Arguments

asid	Integer, assay source id
slvl	Integer of length 1, the starting level to process
elvl	Integer of length 1, the ending level to process
id	Integer, rather than assay source id, the specific assay component or assay end-point id(s) (optional)
type	Character of length 1, the data type, "sc" or "mc"
mc.cores	Integer of length 1, the number of cores to use, set to 1 when using Windows operating system
outfile	Character of length 1, the name of the log file (optional)
runname	Character of length 1, the name of the run to be used in the outfile (optional)
ready_only	Boolean, whether to only process endpoints marked as export_ready = 1.

Details

The tcp1Run function is the core processing function within the package. The function acts as a wrapper for individual processing functions, (ie. mc1, sc1, etc.) that are not exported. If possible, the processing is done in parallel by 'id' by utilizing the [mclapply](#) function within the parallel package.

If slvl is less than 4, 'id' is interpreted as acid and if slvl is 4 or greater 'id' is interpreted as aeid. Must give either 'asid' or 'id'. If an id fails no results get loaded into the database, and the id does not get placed into the cue for subsequent level processing.

The 'type' parameter specifies what type of processing to complete: "mc" for multiple-concentration processing, and "sc" for single-concentration processing.

Value

A list containing the results from each level of processing. Each level processed will return a named logical vector, indicating the success of the processing for the id.

tcp1SubsetChid	<i>Subset level 5 data to a single sample per chemical</i>
----------------	--

Description

tcp1SubsetChid subsets level 5 data to a single tested sample per chemical. In other words, if a chemical is tested more than once (a chid has more than one spid) for a given assay endpoint, the function uses a series of logic to select a single "representative" sample.

Usage

```
tcp1SubsetChid(dat, flag = TRUE, type = "mc", export_ready = FALSE)
```

Arguments

dat	data.table, a data.table with level 5 data
flag	Integer, the mc6_mthd_id values to go into the flag count, see details for more information
type	Character of length 1, the data type, "sc" or "mc"
export_ready	Boolean, default FALSE, should only export ready 1 values be included in calculation

Details

tcp1SubsetChid is intended to work with level 5 data that has chemical and assay information mapped with [tcp1Prep0tpt](#).

To select a single sample, first a "consensus hit-call" is made by majority rule, with ties defaulting to active. After the chemical-wise hit call is made, the samples corresponding to to chemical-wise

hit call are logically ordered using the fit category, the number of the flags, and AC50 (or modl_ga), then the first sample for every chemical is selected.

The flag param can be used to specify a subset of flags to be used in the flag count. Leaving flag TRUE utilize all the available flags. Setting flag to FALSE will do the subsetting without considering any flags.

Value

A data.table with a single sample for every given chemical-assay pair.

See Also

[tcplPrep0tpt](#)

Examples

```
## Not run:
## Load the example level 5 data
d1 <- tcplLoadData(lvl = 5, fld = "aeid", val = 797)
d1 <- tcplPrep0tpt(d1)

## Subset to an example of a duplicated chid
d2 <- d1[chid == 20182]
d2[, list(m4id, hitc, fitc, modl_ga)]

## Here the consensus hit-call is 1 (active), and the fit categories are
## all equal. Therefore, if the flags are ignored, the selected sample will
## be the sample with the lowest modl_ga.
tcplSubsetChid(dat = d2, flag = FALSE)[, list(m4id, modl_ga)]

## End(Not run)
```

tcplVarMat

Create chemical by assay matrices

Description

tcplVarMat creates chemical by assay matrices.

Usage

```
tcplVarMat(
  dsstox_substance_id = NULL,
  aeid = NULL,
  std.vars = c("ac50", "ac50_verbosc", "acc", "acc_verbosc", "hitc_mc", "hitc_sc",
    "zscore"),
  add.vars = NULL,
  flag = TRUE,
```

```
    cyto.pars = list()
  )
```

Arguments

<code>dsstox_substance_id</code>	Integer, chemical ID values to subset on
<code>aeid</code>	Integer, assay endpoint ID values to subset on
<code>std.vars</code>	Character, standard set of matrices; use this parameter to subset this list
<code>add.vars</code>	Character, mc4 or mc5 field(s) not included in the standard list to add additional matrices
<code>flag</code>	Integer or Logical of length 1, passed to tcp1SubsetChid
<code>cyto.pars</code>	List, named list of arguments passed to tcp1CytoPt for z-score matrix

Details

The `tcp1VarMat` function is used to create chemical by assay matrices for different parameters. The standard list of matrices returned includes:

1. "ac50" – The active concentration at 50 the winning model.
2. "ac50_verbose" – The AC50 for the winning model, with text describing some situations.
3. "acc" – The active concentration at user-defined cutoff for the winning model.
4. "acc_verbose" – The ACC for the winning model, with text describing some situations.
5. "hitc_mc" – The hit-call for the winning model in multiple-concentration (mc) screening.
6. "hitc_sc" – The hit-call in single concentration (sc) screening.
7. "zscore" – The z-score based on the output from `tcp1CytoPt`. The formula used for calculating the z-score is $-(ac50 - cyto_pt)/global_mad$

`tcp1VarMat` produces matrices of combined sc-mc output. For the ac50 and acc matrices specifically, values are inserted in place to show complete views of what was tested and what the results were. ac50 and acc values are:

- set to 1e6 when the chemical is tested but negative in mc. In `_verbose` matrices, these are indicated as "MC neg".
- set to 1e7 when the chemical is not tested in mc but was screened in sc with a positive hitcall for the same aeid. In `_verbose` matrices, these are indicated as "SC pos, No MC".
- set to 1e8 when the chemical is not tested in mc but was screened in sc with a negative hitcall for the same aeid. In `_verbose` matrices, these are indicated as "SC neg, No MC"
- not changed when chemical is tested in mc and positive, or not tested in either mc or sc

sc and mc data both are currently required to be included for these calculations. As a result, the "API" driver is not currently supported since it does not return sc data.

To add additional matrices, the `'add.vars'` parameter can be used to specify the fields from the mc4 or mc5 tables to create matrices for.

When more than one sample is included for a chemical/assay pair, `tcp1VarMat` aggregates multiple samples to a chemical level call utilizing [tcp1SubsetChid](#). The `tcp1VarMat` function calls both

tcplSubsetChid and tcplCytoPt (which separately calls tcplSubsetChid). The input for the tcplVarMat 'flag' parameter is passed to the tcplSubsetChid call and used to parse down the data to create the matrices. The tcplSubsetChid called within tcplCytoPt (to parse down the cytotoxicity data used to define the "zscore" matrix) can be modified by passing a separate 'flag' element in the list defined by the 'cyto.pars' parameter.

Value

A list of chemical by assay matrices (data.tables) where the rows are given by the dsstox_substance_id and corresponding chnm (chemical name) columns and the colnames are given by assay endpoint name (aenm).

See Also

[tcplSubsetChid](#)

Examples

```
## Not run:
## Demonstrate the returned values.
varmat <- tcplVarMat()

## Other changes can be made
aeids <- c(80)
dtxsid <- c("DTXSID4034653", "DTXSID2032683", "DTXSID6032358",
"DTXSID0032651", "DTXSID8034401")
varmat <- tcplVarMat(aeid = aeids, dsstox_substance_id = dtxsid)
varmat <- tcplVarMat(aeid = aeids, std.vars = c("ac50", "zscore"))
varmat <- tcplVarMat(aeid = aeids, add.vars = c("m4id", "resp_max", "max_med"))

## To save output to file
library(writexl)
write_xlsx(varmat, path = "varmat_output.xlsx")

## End(Not run)
```

tcplWriteData

Write screening data into the tcpl databases

Description

tcplWriteData takes a data.table with screening data and writes the data into the given level table in the tcpl databases.

Usage

```
tcplWriteData(dat, lvl, type)
```

Arguments

dat	data.table, the screening data to load
lvl	Integer of length 1, the data processing level
type	Character of length 1, the data type, "sc" or "mc"

Details

This function appends data onto the existing table. It also deletes all the data for any acids or aoids dat contains from the given and all downstream tables.

The data type can be either 'mc' for mutiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the level tables, whereas the single concentra-tion will be loaded into the single tables.

Note

This function is not exported and is not intended to be used by the user. The user should only write level 0 data, which is written with [tcplWriteLv10](#).

See Also

[tcplCascade](#), [tcplAppend](#), [tcplWriteLv10](#)

tcplWriteLv10	<i>Write level 0 screening data into the tcpl databases</i>
---------------	---

Description

tcplWriteLv10 takes a data.table with level 0 screening data and writes the data into the level 0 tables in the tcpl databases.

Usage

tcplWriteLv10(dat, type)

Arguments

dat	data.table, the screening data to load
type	Character of length 1, the data type, "sc" or "mc"

Details

This function appends data onto the existing table. It also deletes all the data for any acids or aoids that contains from the given and all downstream tables.

Before writing any data the function maps the assay component source name(s) (acsn) to assay component id (acid), ensures the proper class on each field and checks for every test compound sample id (spid where wllt == "t") in the tcpl chemical database. If field types get changed a warning is given listing the affected fields and they type they were coerced to. If the acsn(s) or spid(s) do not map to the tcpl databases the function will return an error and the data will not be written.

The data type can be either 'mc' for multiple concentration data, or 'sc' for single concentration data. Multiple concentration data will be loaded into the level tables, whereas the single concentration will be loaded into the single tables.

Note

This function should only be used to load level 0 data.

See Also

[tcplCascade](#), [tcplAppend](#)

test_api	<i>List containing ids used for different automated tests of tcpl integration with the CTX APIs, randomly selected from what is available via API.</i>
----------	--

Description

List containing ids used for different automated tests of tcpl integration with the CTX APIs, randomly selected from what is available via API.

Usage

```
test_api
```

Format

A list with 7 items:

aeid Randomly selected assay component endpoint id

acid Assay component id associated with the above aeid

aoid Assay id associated with the above aeid

asid Assay source id associated with the above aeid

dtxsid dsstox substance id of one sample from the above aeid

spid Sample id of one (the same) sample from the above aeid

m4id Level 4 id of one (the same) sample from the above aeid

Source

CTX Bioactivity API

write_lv1_4	<i>Write level 4 with updated schema</i>
-------------	--

Description

Write level 4 with updated schema

Usage

write_lv1_4(dat)

Arguments

dat output of tcplfit2 that has been unnested into a data.table

Index

* data processing functions

tcplRun, [105](#)

* datasets

invitrodb_dd, [15](#)

load_data_columns, [17](#)

mc_test, [36](#)

mc_vignette, [39](#)

mcdat, [35](#)

mthd_list_defaults, [51](#)

sc_test, [62](#)

sc_vignette, [64](#)

scdat, [61](#)

test_api, [111](#)

* multiple-concentration

mc1, [20](#)

mc2, [20](#)

mc3, [23](#)

mc4, [27](#)

mc5, [29](#)

mc6, [33](#)

* single-concentration

sc1, [55](#)

sc2, [58](#)

* tcpl abbreviations

is.odd, [16](#)

lu, [18](#)

lw, [19](#)

sink.reset, [65](#)

.ChemListQ, [5](#)

.ChemQ, [5](#)

.buildAssayQ, [4](#)

.convertNames, [6](#)

.load6DR, [6](#)

.plateHeat, [7](#)

.prepField, [7](#)

AIC, [68](#)

blineShift, [8](#)

check_tcpl_db_schema, [8](#)

Configure functions, [9](#)

constrOptim, [48, 75](#)

data.table, [83, 85, 86](#)

dynamic_table_trunc, [10](#)

flareFunc, [11](#)

get_plot_caption, [12](#)

get_plot_title, [12](#)

get_verbose_tables, [13](#)

Hill model utilites, [13](#)

interlaceFunc, [14](#)

invitrodb_dd, [15](#)

is.odd, [16, 19, 66](#)

length, [19](#)

Load assay information, [16](#)

load_data_columns, [17](#)

lu, [16, 18, 19, 66](#)

lw, [16, 19, 19, 66](#)

mc1, [20, 21, 23, 28, 30, 33](#)

mc2, [20, 20, 22, 23, 28, 30, 33](#)

MC2_Methods, [21, 21](#)

mc2_mthds (MC2_Methods), [21](#)

mc3, [8, 20, 21, 23, 27, 28, 30, 33](#)

MC3_Methods, [23, 24](#)

mc3_mthds, [8](#)

mc3_mthds (MC3_Methods), [24](#)

mc4, [20, 21, 23, 27, 29, 30, 33](#)

MC4_Methods, [28](#)

mc4_mthds (MC4_Methods), [28](#)

mc5, [20, 21, 23, 28, 29, 32, 33](#)

MC5_Methods, [30, 30](#)

mc5_mthds (MC5_Methods), [30](#)

mc6, [11, 15, 20, 21, 23, 28, 30, 33, 35](#)

MC6_Methods, [11, 15, 33, 34](#)

- mc6_mthds (MC6_Methods), 34
- mc_test, 36
- mc_vignette, 39
- mcdat, 35
- mclapply, 106
- Method functions, 47
- Models, 28, 48, 66
- MORELETTERS, 50
- mthd_list_defaults, 51
- Query functions, 52
- Register/update annotation, 53
- registerMthd, 54
- round_n, 55
- sc1, 55, 58, 59
- SC1_Methods, 56, 56
- sc1_mthds (SC1_Methods), 56
- sc2, 56, 58, 61
- SC2_Methods, 59, 59
- sc2_mthds (SC2_Methods), 59
- sc_test, 62
- sc_vignette, 64
- scdat, 61
- sink, 66
- sink.number, 66
- sink.reset, 16, 19, 65
- Startup, 10
- Sys.getenv, 10
- tcplAddModel, 66
- tcplAICProb, 67
- tcplAppend, 68, 110, 111
- tcplCascade, 20, 23, 27, 29, 33, 55, 58, 69, 110, 111
- tcplCode2CASN, 69
- tcplConf (Configure functions), 9
- tcplConfDefault (Configure functions), 9
- tcplConfList (Configure functions), 9
- tcplConfLoad (Configure functions), 9
- tcplConfReset (Configure functions), 9
- tcplConfSave (Configure functions), 9
- tcplCytoPt, 70, 108
- tcpldbStats, 72
- tcplDefine, 73
- tcplDelete, 74
- tcplFit, 28, 48, 68, 74
- tcplFit2, 76
- tcplfit2_fun, 76
- tcplFit2_nest, 77
- tcplFit2_unnest, 77
- tcplGetAeid, 78
- tcplggplot2, 78
- tcplHillACXX (Hill model utilites), 13
- tcplHillConc (Hill model utilites), 13
- tcplHillVal (Hill model utilites), 13
- tcplHit2, 79
- tcplLegacyPlot, 80
- tcplListFlds, 80
- tcplLoadAcid (Load assay information), 16
- tcplLoadAeid (Load assay information), 16
- tcplLoadAid (Load assay information), 16
- tcplLoadAsid (Load assay information), 16
- tcplLoadChem, 6, 81
- tcplLoadChemList, 5, 82
- tcplLoadConcUnit, 83
- tcplLoadData, 6, 83, 90, 97, 104
- tcplLoadUnit, 85
- tcplLvlCount, 86
- tcplMakeAeidMultiPlts, 87
- tcplMakeAeidPlts, 88, 101
- tcplMakeChidMultiPlts, 89
- tcplMthdAssign (Method functions), 47
- tcplMthdClear (Method functions), 47
- tcplMthdList (Method functions), 47
- tcplMthdLoad (Method functions), 47
- tcplMultiplot, 87, 89, 90
- tcplObjCnst, 75
- tcplObjCnst (Models), 48
- tcplObjGnls, 75
- tcplObjGnls (Models), 48
- tcplObjHill, 75
- tcplObjHill (Models), 48
- tcplPlot, 90, 98
- tcplPlotCalcAspectRatio, 95
- tcplPlotFitc, 96
- tcplPlotFits, 66, 88, 96, 100, 101
- tcplPlotLoadData, 98
- tcplPlotLoadWllt, 99
- tcplPlotlyPlot, 100
- tcplPlotM4ID, 100
- tcplPlotPlate, 101
- tcplPlotSetYRange, 102

tcplPlotValidate, [103](#)
tcplPrepOtp, [104](#), [106](#), [107](#)
tcplQuery, [83](#), [85](#), [86](#)
tcplQuery (Query functions), [52](#)
tcplQueryAPI (Query functions), [52](#)
tcplRegister (Register/update
annotation), [53](#)
tcplRun, [20](#), [23](#), [27](#), [29](#), [33](#), [55](#), [58](#), [105](#)
tcplSendQuery, [74](#)
tcplSendQuery (Query functions), [52](#)
tcplSubsetChid, [70](#), [71](#), [106](#), [108](#), [109](#)
tcplUpdate (Register/update
annotation), [53](#)
tcplVarMat, [107](#)
tcplWriteData, [109](#)
tcplWriteLvl0, [110](#), [110](#)
test_api, [111](#)

unique, [19](#)

which, [19](#)
write_lvl_4, [112](#)